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**Understanding the Involvement of Environmental Exposures,
Genetic Risk, and Epigenetic Mechanisms in the Course
of Severe Mental Illness**

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Abbreviations

CNV	copy number variant
CpG	cytosine-phosphate-guanine
DSM	Diagnostic and Statistical Manual of Mental Disorders
DST	Differential Susceptibility Theory
EWAS	epigenome-wide association study
G × E	gene-by-environment
GWAS	genome-wide association studies
HiTOP	Hierarchical Taxonomy of Psychopathology
ICD	International Classification of Diseases
iPSYCH	Lundbeck Foundation Initiative for Integrative Psychiatric Research
LD	linkage disequilibrium
PGC	Psychiatric Genomics Consortium
PRS	polygenic risk score
RDoC	Research Domain Criteria
SNP	single nucleotide polymorphism

List of gene names

<i>BDNF</i>	Brain-derived neurotrophic factor
<i>FKBP5</i>	FKBP Prolyl Isomerase 5
<i>IL-6</i>	Interleukin 6
<i>OXTR</i>	Oxytocin Receptor
<i>POU6F2</i>	POU Class 6 Homeobox 2
<i>SLC6A4</i>	Solute Carrier Family 6 Member 4

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1 Introductory summary

It is now necessary to turn away from arranging illness in orderly well-defined groups, and to set ourselves the undoubtedly higher and more satisfying goal of understanding their essential structure.

– EMIL KRAEPELIN

1.1 Context of the study

Psychiatric disorders like major depression, schizophrenia, and bipolar disorder affect a significant proportion of the global population, accounting for a third of disability worldwide (World Health Organization, 2008). The enormous personal, societal and economic burden of these disorders demands a better understanding of their etiology (Collins et al., 2011). Since the start of the 20th century, psychiatric nosology has been defined by clinical observations of categorical syndromes based on signs and symptoms. However, despite the clinical utility of current classification systems, our understanding of mental disorders through observable behaviors and self-reported feelings and thoughts alone has reached its limits (Clark, Cuthbert, Lewis-Fernandez, Narrow, & Reed, 2017).

With little evidence to support natural boundaries of many mental disorders, the validity of the current Diagnostic and Statistical Manual of Mental Disorders (DSM) and International Classification of Diseases (ICD), is undoubtedly limited (Jablensky, 2016). This is especially true given the arbitrary thresholds used to define psychopathology versus normality, ambiguous boundaries between distinct diagnoses and excessive comorbidity between putatively independent disorders, together with heterogeneity within and instability of psychiatric diagnoses (Clark et al., 2017; Hengartner & Lehmann, 2017; Van Os, 2015). Moreover, the genetic architecture of most common psychiatric disorders does not support diagnostic categories as discrete entities, especially considering their cross-disorder heritability (Lee et al., 2013; Smoller et al., 2019). Needless to say, the current diagnostic system lacks etiological and pathophysiological justification for these clinically and genetically heterogeneous disorders.

With hopes of circumventing these caveats, efforts continue to be made towards a psychiatric nosology informed by disease mechanisms. Unprecedented progress in psychiatric genetics has been made in the last decade with the help of improved molecular technologies and cooperation

among psychiatric researchers (Sullivan, Daly, & O'Donovan, 2012). Genome-wide association studies (GWAS) alongside innovative disciplines like systems biology and functional genomics have begun reporting robust and replicable findings (Howard et al., 2019; Pardiñas et al., 2018; Stahl et al., 2019; Sullivan et al., 2012) which hold potential towards converting these disorders into pathophysiologically-defined diseases. Additionally, new research frameworks using dimensional models of psychopathology have emerged with hopes of better understanding the nature of psychopathology. The Hierarchical Taxonomy of Psychopathology (HiTOP) (Kotov et al., 2017) and Research Domain Criteria (RDoC) (Insel et al., 2010) are just two examples.

Moving forward, continued optimization of existing methodologies in psychiatric genetics and complementary fields like epigenetics have potential to inform disease mechanisms and provide clues to the complex etiology of psychiatric disease. As first recognized by the pivotal work of Emil Kraepelin more than a century ago, longitudinal measures are a key element of psychiatric investigations and should continue to be paired with other approaches (Kraepelin, 1921; McInnis & Greden, 2016). Detailed accounts of environment and longitudinal moderators of disease course are particularly important to better understand the complex relationship between environmental influences and biology (Craddock et al., 2009). Such findings could supplement clinical ratings to form biologically-valid diagnostic criteria and lead to improved diagnostic, personalized therapeutic, and preventative measures. This thesis presents two approaches in alignment with these efforts.

1.2 An introduction to psychiatric genetics

Following the success of linkage studies for the identification of Mendelian disorders, early studies of common psychiatric disorders hoped to discover single genetic variants with large effects (Moreno-De-Luca, Ross, & Ross, 2018). Yet, despite early family and twin studies supporting their strong genetic basis and high heritability (Shih, Belmonte, & Zandi, 2004), empirical data did not support this monogenic theory proving candidate gene studies to be insufficient (Schulze, Fangerau, & Propping, 2004). Rather, it became evident that the genetic structure of these disorders was more likely polygenic (involving many genetic loci) and heterogenous (affected individuals do not all share the same combination of risk alleles) (Fullerton & Nurnberger, 2019). At the turn of the millennium, GWAS became possible for the identification of common single nucleotide polymorphisms (SNPs) on a genome-wide scale, moving the field away from unsuccessful candidate gene studies. With initial GWAS bearing little fruit as a result of their limited sample sizes, it was quickly realized that due to the very small effect sizes of

individual common SNPs and inherent multiple testing burden of GWAS, huge studies and mega analyses were required. This paradigm shift encouraged the formation of large consortia like the international Psychiatric Genomics Consortium (PGC; <https://www.med.unc.edu/pgc/>) and the Lundbeck Foundation Initiative for Integrative Psychiatric Research (iPSYCH; <https://ipsych.dk/en/>) with hopes of leveraging data from thousands of individuals to perform powerful analyses. Today, microarrays and array-based technologies along with rapid expansion in methodological approaches have supported this effort. Through unprecedented technological advancements in the last decade, we are now able to genotype millions of SNPs across the genome in large samples at a quick pace and affordable price. Thereby, the complex polygenic and genetically heterogeneous architecture of major psychiatric disorders have been uncovered. Accordingly, new methodologies continue to be developed to analyze the vast amounts of data coming from GWAS, especially towards a better understanding of the complex interplay between genetic variants.

1.3 A look at genome-wide polygenic scores

While evidence supports the substantial contribution of common SNPs towards risk for disease, overall these variants account for only a fraction of genetic risk and a remarkable part of heritability has yet to be explained. While GWAS have identified robustly associated genome-wide significant variants for bipolar disorder (30 loci) (Stahl et al., 2019), schizophrenia (145 loci) (Pardiñas et al., 2018) and major depression (102 loci) (Howard et al., 2019), alone they explain only a small fraction of liability and hold little predictive accuracy. For example, the genome-wide significant loci for schizophrenia explain just 1.1% of variance on the liability scale (Pardiñas et al., 2018). Much debate has revolved around the reason for this missing heritability, with hypotheses pointing towards unaccounted for small effect variants, gene-gene interactions, as well as rare copy number variants (CNVs), *de novo* mutations, and epigenetic variations (Avramopoulos, 2010; Maher, 2008; Manolio et al., 2009; McCarthy & Hirschhorn, 2008).

In 2007 and 2009, two seminal papers demonstrated that a substantial polygenic component of psychiatric disorders is not found in the strongly associated loci, but rather in the thousands of common alleles that individually do not achieve genome-wide significance (Purcell et al., 2009; Wray, Goddard, & Visscher, 2007). Accordingly, they introduced an approach to capture the cumulative effects of these genetic loci as a single quantitative metric and thus introduced polygenic risk scores (PRSs) to psychiatric genetics. PRSs are calculated as the sum of risk alleles carried by an individual weighted by corresponding genotype effect sizes, derived from summary

statistics from an independent GWAS. These scores are calculated across many p -value thresholds to determine, for example, the optimal threshold for case discrimination, i.e., the threshold with greatest predictive power (Fullerton & Nurnberger, 2019). PRSs have already shown success in real-world health care settings, with a recent study using a schizophrenia PRS to detect risk for schizophrenia and psychosis using health record data (Zheutlin et al., 2019).

In the last years, PRSs have been widely used for a range of applications including the investigation of pleiotropic effects, i.e., the genetic overlap between disorders, evaluating the predictive power of genetic data, differentiation of cases and controls, and to perform experiments comparing individuals at extremes of the PRS distribution (Wray et al., 2014). Another use of PRSs proving to be valuable is their application to the genetics of endophenotypes, also known as intermediate or proxy phenotypes. Endophenotypes, as introduced to psychiatry by Gottesman and Shields, are measurable constructs that lie on the path between phenotypic expression and genes (Gottesman & Shields, 1972; Walters & Owen, 2007). This concept assumes that variation in an endophenotype will be associated with a simpler genetic architecture than the complex disorder as a whole. By definition, endophenotypes are associated with illness in the population, heritable, primarily state-independent, co-segregate with the illness within families, and are found in unaffected relatives at higher rates than in the general population (Gottesman & Gould, 2003). One broadly studied presumed endophenotype is cognitive deficits in patients with severe mental illness (Bora, Yucel, & Pantelis, 2009; Burdick, Goldberg, Harrow, Faull, & Malhotra, 2006; Ivleva et al., 2012).

1.4 An application of PRSs: Introduction to Original Article 1

Comes, A.L., Senner, F., Budde, M., [...], Falkai, P., Schulze, T.G., Papiol, S. (2019). The genetic relationship between educational attainment and cognitive performance in major psychiatric disorders. *Transl Psychiatry*, 9(1), 210. doi:0.1038/s41398-019-0547-x.

Individuals with severe mental illness experience broad cognitive deficits that are often treatment resistant and persistent throughout the lifetime (Sheffield, Karcher, & Barch, 2018). The onset and severity of these cognitive symptoms are highly predictive of patients' long-term prognoses (Bowie & Harvey, 2006; Green, 1996). Given that cognitive deficits are a core feature of severe mental illness, understanding these impairments could have major implications for offering better insight into the psychiatric disorder itself and for targeting effective therapies. Given the characteristics of these deficits, i.e., heritability, presence in unaffected first-degree relatives and

in affected individuals even during periods of remission, they have been named a noteworthy endophenotype for psychotic disorders (Bora et al., 2009; Snitz, Macdonald, & Carter, 2006).

While GWAS for social-science genetics is often limited, proxy-phenotypes can be used to test for associations with well-measured endophenotypes such as cognitive functioning. Educational attainment is strongly correlated with cognition both phenotypically (0.50) and genetically (0.65) (Rietveld et al., 2014). In 2018, a GWAS on educational attainment was conducted in 1.1 million individuals from the general population (Lee et al., 2018). Using these summary statistics, the authors were able to conduct a multi-phenotype analysis generating polygenic scores that explained up to 10% of variance in cognitive performance.

The aim of Original Article 1 was to explore this association across several cognitive domains in a subset of patients from the longitudinal PsyCourse study (Budde et al., 2019). Considering the positive genetic correlations between education and risk for both schizophrenia and bipolar disorder (Hill et al., 2018), we also investigated the association of cognitive performance and PRSs for both disorders. Overall, we hoped this work would allow us to gain insight into the genetic underpinnings of distinct cognitive domains in patients with severe mental illness.

In our sample of patients with known cognitive deficits, educational attainment polygenic scores explained up to 2% of variance in cognitive domains related to learning and working memory. Specifically, the educational attainment polygenic score could explain variance in the number of correctly recalled words of the Verbal Learning and Memory Test (Helmstaedter, Lendt, & Lux, 2001), the backwards task of the Verbal Digit Span (Aster, Neubauer, & Horn, 2006), and crystallized intelligence measured with the Mehrfachwahl-Wortschatz-Intelligenztest (Lehrl, 2005). These findings were particularly interesting considering these cognitive domains are among the most impaired in patients with severe mental illness (Barbosa et al., 2018). Post hoc analyses showed these findings were robust to the effects of diagnosis and current medications. Considering rather inconsistent findings in the literature, it was not completely surprising that no significant effects were observed when investigating the associations between disorder specific PRSs and cognitive outcomes (Schaupp, Schulze, & Budde, 2018). To some extent, these findings support the notion that genetic determinants of cognitive variation within these disorders are at least partly independent from those common SNPs that predispose an individual to a schizophrenia or bipolar diagnosis, however, more evidence is needed.

Findings such as these could have important implications for identifying subgroups of patients with higher risk for a more burdened course of disease and could contribute towards predictive models for more personalized interventions. Further investigation of these associations longitudinally could help identify critical periods of genetic influence on cognitive abilities (Mistry, Harrison, Smith, Escott-Price, & Zammit, 2018). Furthermore, future studies could optimize the polygenic scores used to further reveal the genetic architecture of these deficits, for example, by taking into consideration intricate pleiotropic effects or using hypothesis-based polygenic scores (Lam et al., 2019). Lastly, this study supports the continued investigation of quantitative endophenotypes given their potential to inform the biological pathways lying between genetic variants and fuzzy, qualitative diagnoses. Given the prevalence of endophenotype deficits across many heritable domains, studies aimed at a better understanding of their nature are in alignment with efforts like RDoC with hopes to ‘bridge genomic complexity and disorder heterogeneity’ (Braff & Tamminga, 2016; Insel & Cuthbert, 2009).

1.5 Beyond traditional genetic data: The role of epigenetics

The fact that current PRS estimates explain only a small percentage of risk for psychiatric disorders, e.g., up to 5% in schizophrenia using the optimal p -value threshold of 0.05 (Pardiñas et al., 2018), is not unexpected given the major proportion of risk that environmental factors contribute to psychiatric disorders. Not only do distinct psychiatric disorders share some genetic risk factors, they also share environmental risks with factors like early life adversities playing a major role in risk for multiple psychiatric syndromes (Green et al., 2010; Kessler et al., 2010). Gene-by-environment ($G \times E$) interaction studies have therefore been an integral component to understanding complex psychiatric disorders. Such studies examine the effects of environmental and genetic determinants on a phenotype and further explore their joint effects. Traditionally, the Diathesis-Stress Model has been the guiding conceptual framework for $G \times E$ studies (Monroe & Simons, 1991), which assumes that one’s genetic predisposition makes them vulnerable to the development of a psychiatric disorder when exposed to a certain environmental adversity (Assary, Vincent, Keers, & Pluess, 2018). An opposing view, the Differential Susceptibility Theory (DST), however, proposes individuals vary more generally in their developmental plasticity thus in their susceptibility to both positive and negative environmental influences (Belsky, Bakermans-Kranenburg, & van IJzendoorn, 2007; Belsky & Pluess, 2009; Ellis, Boyce, Belsky, Bakermans-Kranenburg, & Van IJzendoorn, 2011). Despite their value, $G \times E$ studies have faced many limitations such as poor replication rates and high false discovery rates, therefore complementary

approaches have been suggested to further uncover the underlying biological mechanisms of these interactions (Halldorsdottir & Binder, 2017).

One approach to explain more variability in psychiatric phenotypes is to investigate the potential molecular mechanisms underlying $G \times E$ interactions, for example, through the study of epigenetics (Heim & Binder, 2012). Coined by Conrad Waddington in the early 40's, the term 'epigenetics' was first used to describe the involvement of gene-environment interaction during development. The literal meaning of epigenetics is 'on top of the genes' and refers to stable and heritable changes in gene activity or expression without change in the DNA sequence (Jiang, Bressler, & Beaudet, 2004). Epigenetic modifications, acquired throughout the lifetime, include DNA methylation and hydroxymethylation of cytosines, post-translational histone modifications, and expression of non-coding RNAs (Jaenisch & Bird, 2003; Ptak & Petronis, 2010). The epigenotype encompasses the signature of these epigenetic marks across the genome. DNA methylation, that is the addition of a methyl group to a cytosine primarily at a cytosine-phosphate-guanine (CpG) dinucleotide, is thus far the most accessible and widely studied epigenotype (Liu, Faraone, & Glatt, 2019). In general, DNA methylation disrupts binding of transcription factors and leads to transcriptional repression (Murgatroyd, Wu, Bockmuhl, & Spengler, 2010).

In the last decade, different approaches have been used to explore DNA methylation in relation to environmental exposures in patients with severe mental illness through global methylation, candidate gene, and epigenome-wide studies (Pishva et al., 2014). These studies have supported the role of DNA methylation as an adaptive mechanism by which environment, especially early life adversities, can become "embedded" in the genome and have lasting effects that can become maladaptive and lead to psychopathology in adulthood (Matosin, Cruceanu, & Binder, 2017; Nöthling, Malan-Müller, Abrahams, Hemmings, & Seedat, 2019; Szyf, 2011; Szyf & Bick, 2013; Vinkers et al., 2015). However, considering the high cost of epigenotyping, limited studies have conducted hypothesis-free, epigenome-wide investigations, and few have investigated methylation signatures longitudinally. Moreover, few studies have focused specifically on DNA methylation signatures in individuals with bipolar disorder, especially in relation to non-traumatic stressful life events. Considering that epigenetic modifications are sensitive to environment, stable, and reversible, they hold promise to better understand and treat psychiatric disorders (Kular & Kular, 2018).

1.6 Exploring DNA methylation in the course of bipolar disorder:

Introduction to Original Article 2

Comes, A.L., Czamara, D., Adorjan, K., [...], Falkai, P., Schulze, T.G., Heilbronner, U. (2020). The role of environmental stress and DNA methylation in the longitudinal course of bipolar disorder. *Int J Bipolar Disord*, 8(1), 9. doi:10.1186/s40345-019-0176-6.

Evidence indicates an important association between stressful life events and age of onset and the clinical course of affective disorders (Aldinger & Schulze, 2017). However, there is still much to be understood regarding the underlying mechanisms associated with these consequences. While studies have focused on DNA methylation as a trait and state marker of bipolar disorder, as previously reviewed in detail (Ludwig & Dwivedi, 2016), little has been done to investigate the effects of environment on methylation in bipolar individuals (Perroud et al., 2016).

The aim of Original Article 2 was to explore DNA methylation in association with self-reported stressful life events in a sample of bipolar patients from the PsyCourse study (Budde et al., 2019). Specifically, we determined the associations between DNA methylation and childhood trauma according to the Childhood Trauma Screener (Bernstein, Ahluvalia, Pogge, & Handelsman, 1997; Bernstein et al., 2003; Grabe et al., 2012), and impact ratings of positive and negative life events experienced in the last 6 months according to the Life Events Questionnaire (Norbeck, 1984; Sarason, Johnson, & Siegel, 1978). Whole blood samples from two time-points (1 year apart) were used to measure DNA methylation using the Infinium MethylationEPIC BeadChips from Illumina. First, we conducted a targeted analysis by interrogating CpG sites in the vicinity of candidate genes with established roles in the stress response, namely *BDNF*, *OXTR*, *IL6*, *SLC6A4*, and *FKBP5*. We then conducted an exploratory epigenome-wide association study (EWAS), taking into consideration the limited sample size of our study and therefore reducing our analyses to only the most variable CpG sites across the genome. Lastly, we explored epigenetic age-related measures (Horvath, 2013) in association with change in stress and symptom measures over time.

To the best of our knowledge, our study was the first to describe epigenome-wide methylation signatures of bipolar patients over time and in association with non-traumatic stress. Although not a single locus withstood correction for multiple testing, methylation at a single CpG site was suggestively associated with stressful life events. This CpG falls in proximity to the *POU6F2* gene, which has been associated with several psychiatric traits (Anney et al., 2010; Goes et al., 2015; Koshimizu et al., 2019; Nagel, Watanabe, Stringer, Posthuma, & van der Sluis, 2018) as well as educational attainment (Lee et al., 2018; Okbay et al., 2016), and intelligence (Davies et

al., 2018; Hill et al., 2018). The overlap in these findings highlights the importance of integrating and exploring data transdiagnostically to potentially uncover trends and perhaps shared mechanisms across different phenotypes. Additionally, DNA methylation in blood at this CpG site is correlated with methylation in the brain, and *POU6F2* is highly expressed in brain tissue. At the current sample size, however, our study provided limited evidence of the role of DNA methylation in the course of bipolar disorder. Larger, longitudinal studies of well-characterized bipolar patients are warranted to better understand the role of epigenetics in the course of mood disorders.

1.7 Where do we go from here?

Our work took advantage of the unique longitudinal features of the PsyCourse cohort (www.PsyCourse.de) to explore the role of genetics, epigenetics, and environment in the course of severe mental illness.

Our findings from Original Article 1 support the use of educational attainment polygenic scores to identify individuals who might be at greater risk of more severe cognitive impairments and consequently lower levels of functioning. Such information could one day be used to target earlier and perhaps more effective care for these individuals. This work also supports transdiagnostic approaches, showing that the genetic determinants of cognitive functioning were at least partially independent of disorder-specific risks.

As samples continue to grow and become more diverse with the help of large collaborations like the PGC, PRSs will continue to be optimized allowing for more comprehensive genomic risk prediction profiles. Moreover, continued discovery of common SNPs with smaller effects will allow for PRSs that explain greater variance with increased precision (Liu et al., 2019). Future PRSs could also consider weighting and calibration, for example, incorporating linkage disequilibrium (LD) structure, and inclusion of other contributing factors like low-frequency and high-penetrance genetic variants, rare CNVs, epigenetics, and epistatic relationships between variants. Efforts are already being made towards improved PRSs (Ge, Chen, Ni, Feng, & Smoller, 2019). With continued refinement of PRSs, they have potential to become a powerful tool with considerable clinical utility. Especially when paired with other data they could be used to assess the long-term prognosis of patients, subtype disease, and inform treatment response (Fullerton & Nurnberger, 2019). PRSs have already shown clinical utility in multimodal prediction models. For example, in the case of coronary heart disease, the addition of a PRS has been shown to

improve risk prediction relative to clinical risk predictors (Abraham et al., 2016). Furthermore, they have also proved to be useful for prediction in the case of type 1 diabetes, where a PRS could predict progression to type 1 diabetes in children with a family history of the disease (Redondo et al., 2018). Lastly, future studies should continue exploring endophenotypes with PRSs to uncover the level and functional significance of the genetic contributions of psychiatric disorders (Braff & Tamminga, 2016).

Considering the important role of environment in risk for severe mental illness, epigenetic mechanisms are likely to be an important puzzle piece towards precision psychiatry. In Original Article 2, we conducted the first, longitudinal epigenome-wide profiling of DNA methylation in patients with bipolar disorder. Despite the clear limitation of sample size, our study took into consideration critical confounding factors like methylation-based smoking estimates that are often unaccounted for and lead to spurious findings in methylation studies. To gain power, beyond increasing sample size, future studies should reduce the noise of confounding factors, for example, by taking into consideration the disorder-specific, sex, age and genotype-dependent, and tissue-specific nature of DNA methylation (Boks et al., 2015; Marzi et al., 2018; Mehta et al., 2017; Smith et al., 2011; Uddin et al., 2010; Vinkers et al., 2015).

In the future, molecular phenotypes like epigenetics will continue to contribute to our understanding of disease mechanisms, especially when paired with other laboratory and clinical endpoints. In particular, functional research should be paired with epigenetic studies to better understand the role of epigenetic modifications in psychiatric disorders (Mill & Heijmans, 2013). Integrating -omics data like genomics, epigenomics, transcriptomics, and proteomics could lead to the “identification of novel ‘multidimensional’ markers and [to] reveal novel insight in the classification of complex diseases” (Comes et al., 2018).

1.8 Conclusion

Translational studies which integrate multiple levels of data, for example into biopsychosocial models, have potential to reveal novel insight into the etiology and inheritance of severe mental illness. Moreover, such studies have potential to eventually lead to a more valid diagnostic framework of psychiatric disease which could ultimately improve patient prognosis, diagnosis, and treatment. This thesis used approaches to identify mechanisms related to a worse course of disease *within* patients with severe mental illness. In doing so, we showed a genome-wide polygenic score for educational attainment could be used to explain variance within some of the

most impaired cognitive domains associated with psychiatric illness. We also reported longitudinal, epigenome-wide measures of methylation related to stressful life events in bipolar patients, which, to the best of our knowledge, is the first study of its kind. With improved methodology and the integration of data from genetic, epigenetic, and environmental studies, we will continue to pave the way towards precision medicine. Taking advantage of unique resources like the transdiagnostic, longitudinal, PsyCourse study will continue to be invaluable towards this cause.

1.9 References

- Abraham, G., Havulinna, A. S., Bhalala, O. G., Byars, S. G., De Livera, A. M., Yetukuri, L., . . . Inouye, M. (2016). Genomic prediction of coronary heart disease. *European Heart Journal*, 37(43), 3267-3278. doi:10.1093/eurheartj/ehw450
- Aldinger, F., & Schulze, T. G. (2017). Environmental factors, life events, and trauma in the course of bipolar disorder. *Psychiatry and Clinical Neurosciences*, 71(1), 6-17. doi:10.1111/pcn.12433
- Anney, R., Klei, L., Pinto, D., Regan, R., Conroy, J., Magalhaes, T. R., . . . Hallmayer, J. (2010). A genome-wide scan for common alleles affecting risk for autism. *Human Molecular Genetics*, 19(20), 4072-4082. doi:10.1093/hmg/ddq307
- Assary, E., Vincent, J. P., Keers, R., & Pluess, M. (2018). Gene-environment interaction and psychiatric disorders: Review and future directions. *Seminars in Cell and Developmental Biology*, 77, 133-143. doi:10.1016/j.semcdb.2017.10.016
- Aster, M., Neubauer, A., & Horn, R. (2006). *Wechsler Intelligenztest für Erwachsene. Wechsler Intelligence Test for Adults (German revision and adaptation of the WAIS-III of David Wechsler)*. Frankfurt, Germany: Harcourt Test Services.
- Avramopoulos, D. (2010). Genetics of psychiatric disorders methods: Molecular approaches. *Psychiatric Clinics of North America*, 33(1), 1-13. doi:10.1016/j.psc.2009.12.006
- Barbosa, I. G., de Almeida, R. F., Rocha, N. P., Mol, G. C., da Mata Chiacchio Leite, F., Bauer, I. E., & Teixeira, A. L. (2018). Predictors of cognitive performance in bipolar disorder: The role of educational degree and inflammatory markers. *Journal of Psychiatric Research*, 106, 31-37. doi:10.1016/j.jpsychires.2018.09.003
- Belsky, J., Bakermans-Kranenburg, M. J., & van Ijzendoorn, M. H. (2007). For better and for worse: Differential susceptibility to environmental influences. *Current Directions in Psychological Science*, 16(6), 300-304. doi:10.1111/j.1467-8721.2007.00525.x
- Belsky, J., & Pluess, M. (2009). Beyond diathesis stress: Differential susceptibility to environmental influences. *Psychological Bulletin*, 135(6), 885.
- Bernstein, D. P., Ahluvalia, T., Pogge, D., & Handelsman, L. (1997). Validity of the Childhood Trauma Questionnaire in an adolescent psychiatric population. *Journal of the American Academy of Child and Adolescent Psychiatry*, 36(3), 340-348. doi:10.1097/00004583-199703000-00012

- Bernstein, D. P., Stein, J. A., Newcomb, M. D., Walker, E., Pogge, D., Ahluvalia, T., . . . Zule, W. (2003). Development and validation of a brief screening version of the Childhood Trauma Questionnaire. *Child Abuse and Neglect*, 27(2), 169-190. doi:10.1016/S0145-2134(02)00541-0
- Boks, M. P., van Mierlo, H. C., Rutten, B. P., Radstake, T. R., De Witte, L., Geuze, E., . . . Vermetten, E. (2015). Longitudinal changes of telomere length and epigenetic age related to traumatic stress and post-traumatic stress disorder. *Psychoneuroendocrinology*, 51, 506-512. doi:10.1016/j.psyneuen.2014.07.011
- Bora, E., Yucel, M., & Pantelis, C. (2009). Cognitive endophenotypes of bipolar disorder: A meta-analysis of neuropsychological deficits in euthymic patients and their first-degree relatives. *Journal of Affective Disorders*, 113(1-2), 1-20. doi:10.1016/j.jad.2008.06.009
- Bowie, C. R., & Harvey, P. D. (2006). Cognitive deficits and functional outcome in schizophrenia. *Neuropsychiatric Disease and Treatment*, 2(4), 531-536. doi:10.2147/ndt.2006.2.4.531
- Braff, D. L., & Tamminga, C. A. (2016). Endophenotypes, epigenetics, polygenicity and more: Irv Gottesman's dynamic legacy. *Schizophrenia Bulletin*, 43(1), 10-16. doi:10.1093/schbul/sbw157
- Budde, M., Anderson-Schmidt, H., Gade, K., Reich-Erkelenz, D., Adorjan, K., Kalman, J. L., . . . Heilbronner, U. (2019). A longitudinal approach to biological psychiatric research: The PsyCourse study. *American Journal of Medical Genetics. Part B: Neuropsychiatric Genetics*, 180(2), 89-102. doi:10.1002/ajmg.b.32639
- Burdick, K. E., Goldberg, J. F., Harrow, M., Faull, R. N., & Malhotra, A. K. (2006). Neurocognition as a stable endophenotype in bipolar disorder and schizophrenia. *Journal of Nervous and Mental Disease*, 194(4), 255-260. doi:10.1097/01.nmd.0000207360.70337.7e
- Clark, L. A., Cuthbert, B., Lewis-Fernandez, R., Narrow, W. E., & Reed, G. M. (2017). Three approaches to understanding and classifying mental disorder: ICD-11, DSM-5, and the National Institute of Mental Health's Research Domain Criteria (RDoC). *Psychological Science in the Public Interest*, 18(2), 72-145. doi:10.1177/1529100617727266
- Collins, P. Y., Patel, V., Joestl, S. S., March, D., Insel, T. R., Daar, A. S., . . . Walport, M. (2011). Grand challenges in global mental health. *Nature*, 475(7354), 27-30. doi:10.1038/475027a
- Comes, A. L., Papiol, S., Mueller, T., Geyer, P. E., Mann, M., & Schulze, T. G. (2018). Proteomics for blood biomarker exploration of severe mental illness: Pitfalls of the past and

- potential for the future. *Translational Psychiatry*, 8(1), 160. doi:10.1038/s41398-018-0219-2
- Craddock, N., Kendler, K., Neale, M., Nurnberger, J., Purcell, S., Rietschel, M., . . . Thapar, A. (2009). Dissecting the phenotype in genome-wide association studies of psychiatric illness. *British Journal of Psychiatry*, 195(2), 97-99. doi:10.1192/bjp.bp.108.063156
- Davies, G., Lam, M., Harris, S. E., Trampush, J. W., Luciano, M., Hill, W. D., . . . Deary, I. J. (2018). Study of 300,486 individuals identifies 148 independent genetic loci influencing general cognitive function. *Nature Communications*, 9(1), 2098. doi:10.1038/s41467-018-04362-x
- Ellis, B. J., Boyce, W. T., Belsky, J., Bakermans-Kranenburg, M. J., & Van IJzendoorn, M. H. (2011). Differential susceptibility to the environment: An evolutionary–neurodevelopmental theory. *Development and Psychopathology*, 23(1), 7-28. doi:10.1017/S0954579410000611
- Fullerton, J. M., & Nurnberger, J. I. (2019). Polygenic risk scores in psychiatry: Will they be useful for clinicians? *F1000Research*, 8. doi:10.12688/f1000research.18491.1
- Ge, T., Chen, C.-Y., Ni, Y., Feng, Y.-C. A., & Smoller, J. W. (2019). Polygenic prediction via Bayesian regression and continuous shrinkage priors. *Nature Communications*, 10(1), 1776. doi:10.1038/s41467-019-09718-5
- Goes, F. S., McGrath, J., Avramopoulos, D., Wolyniec, P., Pirooznia, M., Ruczinski, I., . . . Pulver, A. E. (2015). Genome-wide association study of schizophrenia in Ashkenazi Jews. *American Journal of Medical Genetics. Part B: Neuropsychiatric Genetics*, 168(8), 649-659. doi:10.1002/ajmg.b.32349
- Gottesman, I. I., & Gould, T. D. (2003). The endophenotype concept in psychiatry: Etymology and strategic intentions. *American Journal of Psychiatry*, 160(4), 636-645. doi:10.1176/appi.ajp.160.4.636
- Gottesman, I. I., & Shields, J. (1972). *Schizophrenia and genetics: A twin study vantage point*. Oxford, England: Academic Press.
- Grabe, H. J., Schulz, A., Schmidt, C. O., Appel, K., Driessen, M., Wingenfeld, K., . . . Freyberger, H. J. (2012). Ein Screeninginstrument für Missbrauch und Vernachlässigung in der Kindheit: der Childhood Trauma Screener (CTS). [A Brief Instrument for the Assessment of Childhood Abuse and Neglect: the Childhood Trauma Screener (CTS)]. *Psychiatrische Praxis*, 39(03), 109-115. doi:10.1055/s-0031-1298984

- Green, J. G., McLaughlin, K. A., Berglund, P. A., Gruber, M. J., Sampson, N. A., Zaslavsky, A. M., & Kessler, R. C. (2010). Childhood adversities and adult psychiatric disorders in the national comorbidity survey replication I: Associations with first onset of DSM-IV disorders. *Archives of General Psychiatry*, 67(2), 113-123.
doi:10.1001/archgenpsychiatry.2009.186
- Green, M. F. (1996). What are the functional consequences of neurocognitive deficits in schizophrenia? *American Journal of Psychiatry*, 153(3), 321-330.
doi:10.1176/ajp.153.3.321
- Halldorsdottir, T., & Binder, E. B. (2017). Gene x environment interactions: From molecular mechanisms to behavior. *Annual Review of Psychology*, 68, 215-241.
doi:10.1146/annurev-psych-010416-044053
- Heim, C., & Binder, E. B. (2012). Current research trends in early life stress and depression: Review of human studies on sensitive periods, gene–environment interactions, and epigenetics. *Experimental Neurology*, 233(1), 102-111.
doi:10.1016/j.expneurol.2011.10.032
- Helmstaedter, C., Lendt, M., & Lux, S. (2001). *Verbaler Lern- und Merkfähigkeitstest (VLMT)*. Göttingen, Germany: Beltz.
- Hengartner, M. P., & Lehmann, S. N. (2017). Why psychiatric research must abandon traditional diagnostic classification and adopt a fully dimensional scope: Two solutions to a persistent problem. *Frontiers in Psychiatry*, 8, 101. doi:10.3389/fpsyt.2017.00101
- Hill, W. D., Marioni, R. E., Maghzi, O., Ritchie, S. J., Hagenaars, S. P., McIntosh, A. M., . . . Deary, I. J. (2018). A combined analysis of genetically correlated traits identifies 187 loci and a role for neurogenesis and myelination in intelligence. *Molecular Psychiatry*, 24(2), 169-181. doi:10.1038/s41380-017-0001-5
- Horvath, S. (2013). DNA methylation age of human tissues and cell types. *Genome Biology*, 14(10), R115. doi:10.1186/gb-2013-14-10-r115
- Howard, D. M., Adams, M. J., Clarke, T.-K., Hafferty, J. D., Gibson, J., Shirali, M., . . . McIntosh, A. M. (2019). Genome-wide meta-analysis of depression identifies 102 independent variants and highlights the importance of the prefrontal brain regions. *Nature Neuroscience*, 22(3), 343-352. doi:10.1038/s41593-018-0326-7
- Insel, T., Cuthbert, B., Garvey, M., Heinssen, R., Pine, D. S., Quinn, K., . . . Wang, P. (2010). Research Domain Criteria (RDoC): Toward a new classification framework for research

- on mental disorders. *American Journal of Psychiatry*, 167(7), 748-751.
doi:10.1176/appi.ajp.2010.09091379
- Insel, T., & Cuthbert, B. N. (2009). Endophenotypes: Bridging genomic complexity and disorder heterogeneity. *Biological Psychiatry*, 66(11), 988-989.
doi:10.1016/j.biopsych.2009.10.008
- Ivleva, E. I., Morris, D. W., Osuji, J., Moates, A. F., Carmody, T. J., Thaker, G. K., . . . Tamminga, C. A. (2012). Cognitive endophenotypes of psychosis within dimension and diagnosis. *Psychiatry Research*, 196(1), 38-44. doi:10.1016/j.psychres.2011.08.021
- Jablensky, A. (2016). Psychiatric classifications: Validity and utility. *World Psychiatry*, 15(1), 26-31. doi:10.1002/wps.20284
- Jaenisch, R., & Bird, A. (2003). Epigenetic regulation of gene expression: How the genome integrates intrinsic and environmental signals. *Nature Genetics*, 33 Suppl, 245-254.
doi:10.1038/ng1089
- Jiang, Y.-h., Bressler, J., & Beaudet, A. L. (2004). Epigenetics and human disease. *Annual Review of Genomics and Human Genetics*, 5(1), 479-510.
doi:10.1146/annurev.genom.5.061903.180014
- Kessler, R. C., McLaughlin, K. A., Green, J. G., Gruber, M. J., Sampson, N. A., Zaslavsky, A. M., . . . Williams, D. R. (2010). Childhood adversities and adult psychopathology in the WHO World Mental Health Surveys. *British Journal of Psychiatry*, 197(5), 378-385.
doi:10.1192/bjp.bp.110.080499
- Koshimizu, H., Nogawa, S., Asano, S., Ikeda, M., Iwata, N., Takahashi, S., . . . Miyakawa, T. (2019). Genome-wide association study identifies a novel locus associated with psychological distress in the Japanese population. *Translational Psychiatry*, 9(1), 52-52.
doi:10.1038/s41398-019-0383-z
- Kotov, R., Krueger, R. F., Watson, D., Achenbach, T. M., Althoff, R. R., Bagby, R. M., . . . Zimmerman, M. (2017). The Hierarchical Taxonomy of Psychopathology (HiTOP): A dimensional alternative to traditional nosologies. *Journal of Abnormal Psychology*, 126(4), 454-477. doi:10.1037/abn0000258
- Kraepelin, E. (1921). Manic depressive insanity and paranoia. *The Journal of Nervous and Mental Disease*, 53(4), 350.
- Kular, L., & Kular, S. (2018). Epigenetics applied to psychiatry: Clinical opportunities and future challenges. *Psychiatry and Clinical Neurosciences*, 72(4), 195-211.
doi:10.1111/pcn.12634

- Lam, M., Hill, W. D., Trampush, J. W., Yu, J., Knowles, E., Davies, G., . . . Lencz, T. (2019). Pleiotropic meta-analysis of cognition, education, and schizophrenia differentiates roles of early neurodevelopmental and adult synaptic pathways. *The American Journal of Human Genetics*, 105(2), 334-350. doi:10.1016/j.ajhg.2019.06.012
- Lee, J. J., Wedow, R., Okbay, A., Kong, E., Maghzian, O., Zacher, M., . . . Cesarini, D. (2018). Gene discovery and polygenic prediction from a genome-wide association study of educational attainment in 1.1 million individuals. *Nature Genetics*, 50(8), 1112-1121. doi:10.1038/s41588-018-0147-3
- Lee, S. H., Ripke, S., Neale, B. M., Faraone, S. V., Purcell, S. M., Perlis, R. H., . . . Wray, N. R. (2013). Genetic relationship between five psychiatric disorders estimated from genome-wide SNPs. *Nature Genetics*, 45(9), 984-994. doi:10.1038/ng.2711
- Lehrl, S. (2005). *Mehrfachwahl-Wortschatz-Intelligenztest (MWT-B)*. Balingen, Germany: Spitta Verlag.
- Liu, C., Faraone, S. V., & Glatt, S. J. (2019). Psychiatric genetics, epigenetics, and cellular models in coming years. *Journal of Psychiatry and Brain Science*, 4, e190012. doi:10.20900/jpbs.20190012
- Ludwig, B., & Dwivedi, Y. (2016). Dissecting bipolar disorder complexity through epigenomic approach. *Molecular Psychiatry*, 21(11), 1490-1498. doi:10.1038/mp.2016.123
- Maher, B. (2008). Personal genomes: The case of the missing heritability. *Nature*, 456(7218), 18-21. doi:10.1038/456018a
- Manolio, T. A., Collins, F. S., Cox, N. J., Goldstein, D. B., Hindorff, L. A., Hunter, D. J., . . . Visscher, P. M. (2009). Finding the missing heritability of complex diseases. *Nature*, 461(7265), 747-753. doi:10.1038/nature08494
- Marzi, S. J., Sugden, K., Arseneault, L., Belsky, D. W., Burrage, J., Corcoran, D. L., . . . Caspi, A. (2018). Analysis of DNA methylation in young people: Limited evidence for an association between victimization stress and epigenetic variation in blood. *American Journal of Psychiatry*, 175(6), 517-529. doi:10.1176/appi.ajp.2017.17060693
- Matosin, N., Cruceanu, C., & Binder, E. B. (2017). Preclinical and clinical evidence of DNA methylation changes in response to trauma and chronic stress. *Chronic Stress (Thousand Oaks)*, 1. doi:10.1177/2470547017710764
- McCarthy, M. I., & Hirschhorn, J. N. (2008). Genome-wide association studies: Potential next steps on a genetic journey. *Human Molecular Genetics*, 17(R2), R156-R165. doi:10.1093/hmg/ddn289

- McInnis, M. G., & Greden, J. F. (2016). Longitudinal studies: An essential component for complex psychiatric disorders. *Neuroscience Research*, 102, 4-12.
doi:10.1016/j.neures.2015.05.004
- Mehta, D., Bruenig, D., Carrillo-Roa, T., Lawford, B., Harvey, W., Morris, C. P., . . . Voisey, J. (2017). Genomewide DNA methylation analysis in combat veterans reveals a novel locus for PTSD. *Acta Psychiatrica Scandinavica*, 136(5), 493-505.
doi:10.1111/acps.12778
- Mill, J., & Heijmans, B. T. (2013). From promises to practical strategies in epigenetic epidemiology. *Nature Reviews Genetics*, 14(8), 585-594. doi:10.1038/nrg3405
- Mistry, S., Harrison, J. R., Smith, D. J., Escott-Price, V., & Zammit, S. (2018). The use of polygenic risk scores to identify phenotypes associated with genetic risk of bipolar disorder and depression: A systematic review. *Journal of Affective Disorders*, 234, 148-155. doi:10.1016/j.jad.2018.02.005
- Monroe, S. M., & Simons, A. D. (1991). Diathesis-stress theories in the context of life stress research: Implications for the depressive disorders. *Psychological Bulletin*, 110(3), 406-425. doi:10.1037/0033-2909.110.3.406
- Moreno-De-Luca, D., Ross, M. E., & Ross, D. A. (2018). Leveraging the power of genetics to bring precision medicine to psychiatry: Too little of a good thing? *Biological Psychiatry*, 83(8), e45-e46. doi:10.1016/j.biopsych.2018.02.013
- Murgatroyd, C., Wu, Y., Bockmuhl, Y., & Spengler, D. (2010). Genes learn from stress: How infantile trauma programs us for depression. *Epigenetics*, 5(3), 194-199.
doi:10.4161/epi.5.3.11375
- Nagel, M., Watanabe, K., Stringer, S., Posthuma, D., & van der Sluis, S. (2018). Item-level analyses reveal genetic heterogeneity in neuroticism. *Nature Communications*, 9(1), 905. doi:10.1038/s41467-018-03242-8
- Norbeck, J. S. (1984). Modification of life event questionnaires for use with female respondents. *Research in Nursing and Health*, 7(1), 61-71.
doi:10.1002/nur.4770070110
- Nöthling, J., Malan-Müller, S., Abrahams, N., Hemmings, S. M. J., & Seedat, S. (2019). Epigenetic alterations associated with childhood trauma and adult mental health outcomes: A systematic review. *The World Journal of Biological Psychiatry*, 1-20.
doi:10.1080/15622975.2019.1583369

- Okbay, A., Beauchamp, J. P., Fontana, M. A., Lee, J. J., Pers, T. H., Rietveld, C. A., . . . Benjamin, D. J. (2016). Genome-wide association study identifies 74 loci associated with educational attainment. *Nature*, 533(7604), 539-542. doi:10.1038/nature17671
- Pardiñas, A. F., Holmans, P., Pocklington, A. J., Escott-Price, V., Ripke, S., Carrera, N., . . . Walters, J. T. R. (2018). Common schizophrenia alleles are enriched in mutation-intolerant genes and in regions under strong background selection. *Nature Genetics*, 50(3), 381-389. doi:10.1038/s41588-018-0059-2
- Perroud, N., Zewdie, S., Stenz, L., Adouan, W., Bavamian, S., Prada, P., . . . Dayer, A. (2016). Methylation of serotonin receptor 3A in ADHD, borderline personality, and bipolar disorders: Link with severity of the disorders and childhood maltreatment. *Depression and Anxiety*, 33(1), 45-55. doi:10.1002/da.22406
- Pishva, E., Kenis, G., van den Hove, D., Lesch, K. P., Boks, M. P., van Os, J., & Rutten, B. P. (2014). The epigenome and postnatal environmental influences in psychotic disorders. *Social Psychiatry and Psychiatric Epidemiology*, 49(3), 337-348. doi:10.1007/s00127-014-0831-2
- Ptak, C., & Petronis, A. (2010). Epigenetic approaches to psychiatric disorders. *Dialogues in Clinical Neuroscience*, 12(1), 25-35.
- Purcell, S. M., Wray, N. R., Stone, J. L., Visscher, P. M., O'Donovan, M. C., Sullivan, P. F., & Sklar, P. (2009). Common polygenic variation contributes to risk of schizophrenia and bipolar disorder. *Nature*, 460(7256), 748-752. doi:10.1038/nature08185
- Redondo, M. J., Geyer, S., Steck, A. K., Sharp, S., Wentworth, J. M., Weedon, M. N., . . . Oram, R. A. (2018). A type 1 diabetes genetic risk score predicts progression of islet autoimmunity and development of type 1 diabetes in individuals at risk. *Diabetes Care*, 41(9), 1887-1894. doi:10.2337/dc18-0087
- Rietveld, C. A., Esko, T., Davies, G., Pers, T. H., Turley, P., Benyamin, B., . . . Koellinger, P. D. (2014). Common genetic variants associated with cognitive performance identified using the proxy-phenotype method. *Proceedings of the National Academy of Sciences of the United States of America*, 111(38), 13790-13794. doi:10.1073/pnas.1404623111
- Sarason, I. G., Johnson, J. H., & Siegel, J. M. (1978). Assessing the impact of life changes: Development of the Life Experiences Survey. *Journal of Consulting and Clinical Psychology*, 46(5), 932-946. doi:10.1037//0022-006x.46.5.932
- Schaupp, S., Schulze, T., & Budde, M. (2018). Let's talk about the association between schizophrenia polygenic risk scores and cognition in patients and the general

- population: A review. *Journal of Psychiatry and Brain Science*, 3(6), 12.
doi:10.20900/jpbs.20180012
- Schulze, T. G., Fangerau, H., & Propping, P. (2004). From degeneration to genetic susceptibility, from eugenics to genethics, from Bezugsziffer to LOD score: The history of psychiatric genetics. *International Review of Psychiatry*, 16(4), 246-259.
doi:10.1080/09540260400014419
- Sheffield, J. M., Karcher, N. R., & Barch, D. M. (2018). Cognitive deficits in psychotic disorders: A lifespan perspective. *Neuropsychology Review*, 28(4), 509-533. doi:10.1007/s11065-018-9388-2
- Shih, R. A., Belmonte, P. L., & Zandi, P. P. (2004). A review of the evidence from family, twin and adoption studies for a genetic contribution to adult psychiatric disorders. *International Review of Psychiatry*, 16(4), 260-283. doi:10.1080/09540260400014401
- Smith, A. K., Conneely, K. N., Kilaru, V., Mercer, K. B., Weiss, T. E., Bradley, B., . . . Ressler, K. J. (2011). Differential immune system DNA methylation and cytokine regulation in post-traumatic stress disorder. *American Journal of Medical Genetics. Part B: Neuropsychiatric Genetics*, 156b(6), 700-708. doi:10.1002/ajmg.b.31212
- Smoller, J. W., Andreassen, O. A., Edenberg, H. J., Faraone, S. V., Glatt, S. J., & Kendler, K. S. (2019). Psychiatric genetics and the structure of psychopathology. *Molecular Psychiatry*, 24(3), 409-420. doi:10.1038/s41380-017-0010-4
- Snitz, B. E., Macdonald, A. W., 3rd, & Carter, C. S. (2006). Cognitive deficits in unaffected first-degree relatives of schizophrenia patients: A meta-analytic review of putative endophenotypes. *Schizophrenia Bulletin*, 32(1), 179-194. doi:10.1093/schbul/sbi048
- Stahl, E. A., Breen, G., Forstner, A. J., McQuillin, A., Ripke, S., Trubetskoy, V., . . . the Bipolar Disorder Working Group of the Psychiatric Genomics Consortium. (2019). Genome-wide association study identifies 30 loci associated with bipolar disorder. *Nature Genetics*, 51(5), 793-803. doi:10.1038/s41588-019-0397-8
- Sullivan, P. F., Daly, M. J., & O'Donovan, M. (2012). Genetic architectures of psychiatric disorders: The emerging picture and its implications. *Nature Reviews Genetics*, 13(8), 537-551. doi:10.1038/nrg3240
- Szyf, M. (2011). The early life social environment and DNA methylation: DNA methylation mediating the long-term impact of social environments early in life. *Epigenetics*, 6(8), 971-978. doi:10.4161/epi.6.8.16793

- Szyf, M., & Bick, J. (2013). DNA methylation: A mechanism for embedding early life experiences in the genome. *Child Development*, 84(1), 49-57. doi:10.1111/j.1467-8624.2012.01793.x
- Uddin, M., Aiello, A. E., Wildman, D. E., Koenen, K. C., Pawelec, G., de Los Santos, R., . . . Galea, S. (2010). Epigenetic and immune function profiles associated with posttraumatic stress disorder. *Proceedings of the National Academy of Sciences of the United States of America*, 107(20), 9470-9475. doi:10.1073/pnas.0910794107
- Van Os, J. (2015). The transdiagnostic dimension of psychosis: Implications for psychiatric nosology and research. *Shanghai Archives of Psychiatry*, 27(2), 82-86. doi:10.11919/j.issn.1002-0829.215041
- Vinkers, C. H., Kalafateli, A. L., Rutten, B. P., Kas, M. J., Kaminsky, Z., Turner, J. D., & Boks, M. P. (2015). Traumatic stress and human DNA methylation: A critical review. *Epigenomics*, 7(4), 593-608. doi:10.2217/epi.15.11
- Walters, J. T. R., & Owen, M. J. (2007). Endophenotypes in psychiatric genetics. *Molecular Psychiatry*, 12(10), 886-890. doi:10.1038/sj.mp.4002068
- World Health Organization. (2008). The global burden of disease: 2004 update. In. Geneva: World Health Organization.
- Wray, N. R., Goddard, M. E., & Visscher, P. M. (2007). Prediction of individual genetic risk to disease from genome-wide association studies. *Genome Research*, 17(10), 1520-1528. doi:10.1101/gr.6665407
- Wray, N. R., Lee, S. H., Mehta, D., Vinkhuyzen, A. A., Dudbridge, F., & Middeldorp, C. M. (2014). Research review: Polygenic methods and their application to psychiatric traits. *Journal of Child Psychology and Psychiatry and Allied Disciplines*, 55(10), 1068-1087. doi:10.1111/jcpp.12295
- Zheutlin, A. B., Dennis, J., Karlsson Linner, R., Moscati, A., Restrepo, N., Straub, P., . . . Smoller, J. W. (2019). Penetrance and pleiotropy of polygenic risk scores for schizophrenia in 106,160 patients across four health care systems. *American Journal of Psychiatry*, 176(10), 846-855. doi:10.1176/appi.ajp.2019.18091085

2 Original Article 1

2.1 Individual contributions and reference

The study “The genetic relationship between educational attainment and cognitive performance in major psychiatric disorders” was published in *Translational Psychiatry* in 2019. It was conducted under the supervision of T.G.S. and S.P. The research was designed by A.L.C. in consultation with F.S. and M.B. S.P. supported A.L.C in the calculation of polygenic risk scores. A.L.C performed the statistical analysis, wrote the manuscript, and accompanied the publication process as corresponding author. All co-authors critically revised and approved the manuscript.

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ARTICLE

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The genetic relationship between educational attainment and cognitive performance in major psychiatric disorders

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Abstract

Cognitive deficits are a core feature of psychiatric disorders like schizophrenia and bipolar disorder. Evidence supports a genome-wide polygenic score (GPS) for educational attainment (GPS_{EDU}) can be used to explain variability in cognitive performance. We aimed to identify different cognitive domains associated with GPS_{EDU} in a transdiagnostic clinical cohort of chronic psychiatric patients with known cognitive deficits. Bipolar and schizophrenia patients from the PsyCourse cohort ($N = 730$; 43% female) were used. Likewise, we tested whether GPSs for schizophrenia (GPS_{SZ}) and bipolar disorder (GPS_{BD}) were associated with cognitive outcomes. GPS_{EDU} explained 1.5% of variance in the backward verbal digit span, 1.9% in the number of correctly recalled words of the Verbal Learning and Memory Test, and 1.1% in crystallized intelligence. These effects were robust to the influences of treatment and diagnosis. No significant associations between GPS_{SZ} or GPS_{BD} with cognitive outcomes were found. Furthermore, these risk scores did not confound the effect of GPS_{EDU} on cognitive outcomes. GPS_{EDU} explains a small fraction of cognitive performance in adults with psychiatric disorders, specifically for domains related to linguistic learning and working memory. Investigating such a proxy-phenotype longitudinally, could give intriguing insight into the disease course, highlighting at what time genes play a more influential role on cognitive performance. Better understanding the origin of these deficits might help identify those patients at risk for lower levels of functioning and poor social outcomes. Polygenic estimates may in the future be part of predictive models for more personalized interventions.

Introduction

Cognitive deficits are a core and robust feature of psychiatric disorders like bipolar disorder and schizophrenia, present even during periods of remission^{1–3}. These deficits are key predictors of long-term functional and social outcomes and are difficult to treat with current pharmaceutical options or behavioral interventions^{4–6}.

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Considering the associated psychosocial burden and high prevalence of these deficits among patients, psychiatric researchers have put considerable effort towards understanding their underlying mechanisms. Thus far, genome-wide association studies (GWAS) have provided evidence supporting the polygenic architecture and remarkable heritability of cognitive performance in population-based cohorts^{7–10}. Furthermore, evidence supports the phenotypic and genetic stability of individual cognitive differences across the lifetime in domains including executive functioning, attention, and verbal memory^{11–14}. As studies have shown evidence of impairments even in unaffected first-degree relatives of individuals with psychiatric disorders, cognitive deficits have been hypothesized as a valuable endophenotype of interest for better understanding the genetic risk factors of psychiatric disorders^{15–17}.

Intelligence, encompassing cognition, is highly heritable and an imperative predictor of occupational and health outcomes¹⁸. Despite high heritability estimates of intelligence, indicated to be up to 80% in adulthood¹⁹, unraveling the underlying genetic contribution of intelligence differences using GWAS has been challenging and thus far little of the observed heritability has been explained^{11,20}. To date, studies on intelligence have been limited by insufficient sample sizes and further complicated by the challenge of precise and reliable measurements for this complex phenotype^{7–10,20}. The latest GWAS on intelligence identified 205 genomic loci implicating up to 1016 genes, which explained approximately five percent of the variance in intelligence²¹. Another large study reported a genome-wide polygenic score (GPS) that could explain 4.3% of variance in general cognitive function¹¹.

Educational attainment is moderately heritable and has been obtained as a demographic item in countless medical datasets and for cohorts of which genetic data is available²². In the last decade, educational attainment has been proposed as a proxy-phenotype for cognition, as it is highly associated with intelligence both on a phenotypic (0.50) and genetic level (0.65)^{18,20,23–26}. Notably, GPS based on GWAS summary statistics for years of education predict more variance in intelligence than the phenotype years of education per se^{18,25}, reflecting the substantial genetic correlation between both phenotypes. The largest GWAS of educational attainment published to date, based on 1.1 million individuals, identified 1271 lead single nucleotide polymorphisms (SNPs)²². Through a multi-phenotype analysis of educational attainment and three cognitive phenotypes, the authors were able to generate a GPS which explained 7–10% of variance in cognitive performance in the general population. The SNPs identified implicated genes involved in neurodevelopmental processes and neuron-to-neuron communication²². The

authors showed that the use of educational attainment as a proxy-phenotype could uncover genetic variants to be used as a set of “empirically-based candidate genes” for future studies, for example testing associations with important endophenotypes like cognition²⁷.

Studies have already demonstrated an important association of educational attainment GPS (GPS_{EDU}) with cognitive performance, showing that, in a general population, a higher GPS is associated with higher performance on neurocognitive tests²⁸. However, limited evidence exists supporting this association in patients with known cognitive deficits^{29,30}, and there remains a need to investigate this association across different cognitive domains. Here we analyzed whether GPS_{EDU} could be used to explain variability in different cognitive domains in chronic patients with schizophrenia and bipolar disorder from the PsyCourse cohort³¹. This transdiagnostic approach aligns with the growing evidence for dimensional models that cut across diagnostic categories in psychiatry and is supported by the large cognitive, clinical and genetic overlaps between both disorders^{32,33}. Particularly, the genetic overlap between both disorders has been firmly established by heritability estimates derived from population-based multi-generation registers³⁴ and by recent molecular studies that have reported an outstanding genetic correlation ($r_g = 0.70 \pm 0.02$)³⁵.

Considering the positive genetic correlations reported between education and both schizophrenia ($r_g = 0.10$) and bipolar risk ($r_g = 0.28$)³⁶, we further assessed how GPSs for both schizophrenia (GPS_{SZ}) and bipolar disorder (GPS_{BD}) were associated with cognitive performance in our sample.

Materials & methods

Participants

Data were used from the multicenter, PsyCourse study in Germany and Austria, consisting of participants of European ancestry (www.PsyCourse.de)³¹. Participants were phenotyped using a comprehensive battery including data on socio-demographics, psychopathology, cognition, and functioning assessed at each of four visits (6-month intervals). Recruitment strategies and characterization of all participants has been previously described in detail³¹. The sample selected for this project comprised a total of 730 participants with a DSM-IV³⁷ diagnosis of schizophrenia, schizoaffective disorder, or bipolar disorder (type I or II). Additionally, cognitive data available from 320 nonclinical (control) participants was used to give an orientation to the range of phenotypic data available in the PsyCourse cohort and to confirm general, well-replicated findings of lower cognitive performance in patients with psychiatric disorders compared to healthy controls. The study was approved by the local ethics committee for each study center and was carried out

following the rules of the Declaration of Helsinki. All individuals provided written informed consent as previously described³¹.

Psychopathology psychometric instrument

The Positive and Negative Syndrome Scale (PANSS) is a clinical instrument used to measure symptom severity in schizophrenia and routinely used to assess a variety of disorders including bipolar disorder³⁸. A continuous, total score of the three subscales, i.e., positive (e.g., hallucinations and delusions), negative (e.g., emotional withdrawal and blunted affect), and general symptoms (e.g., somatic concern and poor attention) was used as an indication of disease severity at the time of testing.

Cognitive performance psychometric instruments

Cognitive tests were administered at each study visit. The Verbal Learning and Memory Test (VLMT) was introduced at visit 2. For all other cognitive measures, scores from visit 1 were used for analyses.

Crystallized intelligence

The MWT-B (Mehrfachwahl-Wortschatz-Intelligenz test) was used to measure crystallized intelligence^{39,40}. In this test, subjects were presented with 37 sets of five words arranged according to the level of difficulty. Four words of each set were fictitious constructions of known vernacular (i.e., they do not exist in the German language), while one word really exists. Subjects were asked to cross out the word they know to exist. The total number of correctly marked lines was used as a score⁴⁰.

Trail-Making-Test (TMT)

The TMT is a measure of visual attention and task switching and is one of several executive functioning measures. The test consists of two parts, part A assesses psychomotor speed of the participant, and part B assesses switching between two automated tasks (counting and reciting the alphabet). The time taken to complete each part of the test was measured and the difference in time needed (part B-part A) was used, as it is considered a more accurate measure of the divided attention and alternating sequencing tasks tested in part B^{41–43}. In this case, a higher score meant worse cognitive performance.

Verbal digit span

The verbal digit span, from the Wechsler Adult Intelligence Scale, assesses short-term (forward digit-span) and working memory capacity (backward digit-span). Briefly, participants were asked to recall verbally a sequence of digits, with increasingly longer sequences in each trial. For each correctly recalled string of digits, one point was given. The test was ended when the participant was unable to correctly repeat two presented strings of the

same length. The difference between the forward and backward task is that the latter involves mental manipulation as the participant is required to repeat the digits in backward order⁴⁴. A score for each task was considered.

Digit-Symbol-Test (DST)

The DST is a subset of the Wechsler Adult Intelligence Scale⁴⁵ and measures processing speed, working memory, visuospatial processing and attention. In this test, the participant was asked to use a key of numbers 1–9 with coinciding symbols to draw the appropriate symbol that matched the number given. The participant was given 120 s to fill in as many corresponding symbols as possible. In the end, the correct number of symbols drawn was totaled to get an overall score.

Verbal Learning and Memory Test (VLMT)

The VLMT is the German version of the Auditory Verbal Learning Test⁴⁶. This word-list learning paradigm assesses several memory parameters through serial list learning with subsequent distraction, retrieval after distraction and half-hour time delay, and through a recognition task. The test consists of two different word lists which are each 15 independent words and a recognition list which includes 30 words from the two lists and 20 similar distractor words. Four VLMT scores were rated, the first for the number of correctly recalled words from the first list, a second score for the number of words lost after distraction, a third score of words lost after a time interval, and a fourth score of correctly recalled words from the recognition list⁴⁷.

Biological samples

Peripheral blood samples were used for DNA extraction using standard techniques. DNA samples were then used to genotype patients for calculation of GPSs. Genotype data for controls was not available at the time of this investigation and they have not been used for GPS analyses.

GPS estimation

DNA samples were genotyped using the Infinium PsychArray Beadchip (Psychip, Illumina, San Diego, CA, USA). Following standard quality control procedures, imputation was performed using the 1000 Genomes Phase 3 reference panel as previously described in detail^{31,48}. GPSs were calculated for all individuals using PLINK 1.90b5.3⁴⁹. Summary statistics for educational attainment were obtained from the Social Science Genetic Association Consortium (<https://www.thessgac.org/data>)²². These summary stats are derived from analyses excluding 23andMe samples. Summary statistics from the most recent Psychiatric Genomics Consortium GWASs for schizophrenia⁵⁰ and bipolar disorder⁵¹ were used. All

GPSs were calculated based on summary statistics from the discovery datasets, excluding low quality imputed variants (info score < 80%) in the test dataset, rare SNPs (minor allele frequency < 0.05), and ambiguous markers (A/T and C/G). Following the methodology of previous studies⁵², SNPs in the extended major histocompatibility complex region (chromosome 6: 25–34 Mbp) were completely excluded for the calculation of GPS_{EDU} while only the top-associated SNP in this region was included for the calculations of GPS_{SZ} and GPS_{BD}. Data was clumped in windows of 500 kbp, discarding variants in LD ($R^2 > .1$) with another more significantly associated marker.

GPSs were then calculated by multiplying the imputation dosage for each risk allele by the log(OR) of each genetic variant. The resulting values were summed to obtain an individual estimate of the genetic burden in each individual across different SNP p -value thresholds (p_T). Scores for GPS_{SZ} and GPS_{BD} were calculated based on best discrimination thresholds according to previous findings, i.e., $p_T < 0.05$ ⁵⁰ and $p_T < 0.01$ ⁵¹, respectively. GPS_{EDU} was calculated at four different p -value thresholds, from including only genome-wide significant SNPs to inclusion of all SNPs: $p_T < 5 \times 10^{-8}$, 0.05, 0.1, and 1. All GPSs were approximately normally distributed and standardized via z-score transformation.

Statistical analyses

Sample characteristics

As proof of concept, the effect of case status on cognitive performance was investigated using participants from the PsyCourse cohort. Visual inspection of boxplots comparing case versus control scores was performed and the effect of case status on cognitive domains was further determined through linear regression models, adjusting for age and sex. Socio-demographic and clinical characteristics were tested for between-group differences using the independent sample t-test for continuous data and Pearson's chi-squared tests for categorical variables. As an additional validation analysis, we investigated the relationship between GPS_{EDU} and educational attainment in our sample using ordinal logistic regression, adjusting for age, sex, the interaction between age and sex, and the first 10 PCs, according to previous work²². All analyses were performed using R statistical software version 3.4.0⁵³. An initial examination of the distributions of raw cognitive scores was performed to identify and exclude outliers based on Tukey's definition (removal of values beyond $3 \times$ the interquartile range)⁵⁴.

GPS analyses of cognitive performance

The effect of GPS_{EDU} on cognitive performance of cases was explored. Blockwise linear regression models were used to estimate the amount of variation in cognitive performance explained by the z-standardized GPS_{EDU} at

the four thresholds previously described. For each cognitive outcome, all base models were adjusted for confounding variables measured at the time of testing, i.e., age, age², sex, in/outpatient status, study center, and PANSS sum scores. Although our participants are chronic patients, duration of illness was considered an important covariate which could confound our results. However, as duration of illness proved to be well correlated with age ($r = 0.53$), ultimately only age was kept in the models. To guard against population stratification, the first 20 ancestry principal components (PCs) were included in our models and selected for each cognitive outcome tested using backward model selection ($p < 0.05$)⁵⁵. The significant PCs were as follows: PCs 12 and 17 for the Verbal digit span (forward task); PC 7 for the DST; PCs 1 and 18 for the MWT-B; PCs 1 and 5 for the VLMT- loss of words after time; and PC 16 for the VLMT- correctly recognized words. No significant PCs were found for the other cognitive outcomes. For each cognitive outcome of interest, we measured the incremental adjusted- R^2 , that is the gain in the coefficient of determination when the GPS_{EDU} was added as covariate to the regression model for each phenotype (cognition score) on a set of baseline covariates. Multiple testing was corrected for using the False Discovery Rate (FDR) method correcting for the polygenic profiles at all four thresholds and for all phenotypes investigated. Visual inspection of the residuals for each model was performed to be sure the requirement of normally distributed model residuals had been fulfilled.

GPS analyses of schizophrenia and bipolar disorder

Using blockwise linear regression models as described above, we tested whether polygenic scores for schizophrenia and bipolar disorder influenced cognitive outcomes. This was tested for both the GPS_{SZ} and GPS_{BD} separately. We then determined how the genetic risk for schizophrenia and bipolar disorder influenced the effect of GPS_{EDU} on cognitive outcomes. Both scores were included (separately) in those models in which GPS_{EDU} was significantly associated with the cognitive outcome tested.

Additional analyses

Post hoc analyses were performed to determine the robustness of our findings when correcting for diagnosis (bipolar-I disorder, bipolar-II disorder, schizophrenia, or schizoaffective disorder) and medication (number of antipsychotics, antidepressants, mood stabilizers, and tranquilizers at time of assessment). Furthermore, taking into consideration the significant correlation between memory and crystallized intelligence⁵⁶, we performed a mediation analysis introducing the DST and VLMT (number of correctly recalled words) as covariates in our model testing the association between GPS_{EDU} ($p_T < 1$)

and crystallized intelligence. Multicollinearity diagnostics were performed.

Results

A description of socio-demographic variables for participants is presented in Table 1. Seven-hundred and thirty patients with schizophrenia and bipolar disorder were used for analyses. The mean age of these participants was 43.19 years, the majority of which were male. The majority of cases (46.2%) were diagnosed with schizophrenia, 10.0% were schizoaffective, 35.1% were bipolar-I patients and 8.7% were bipolar-II patients. During baseline visits, 47.7% of patients were being treated as day/inpatients.

The correlations between cognitive domains were assessed (Supplementary Fig. 1). Boxplots depicting case versus control performance across all cognitive domains are shown in Supplementary Fig. 2. Investigation of linear models to test the effect of case status on cognitive performance, after adjusting for age and sex, showed a significant effect in the direction expected, i.e., a decreased performance for cases (Supplementary Table 1). Educational attainment was significantly associated with GPS_{EDU} in the direction expected (Supplementary Table 2).

Our investigation of the effect of GPS_{EDU} ($p_T < 1$) on cognitive performance in patients resulted in a significant increase in Nagelkerke's R^2 of 1.5% for the verbal digit span (backward; Fig. 1a), 1.9% for the VLMT number of

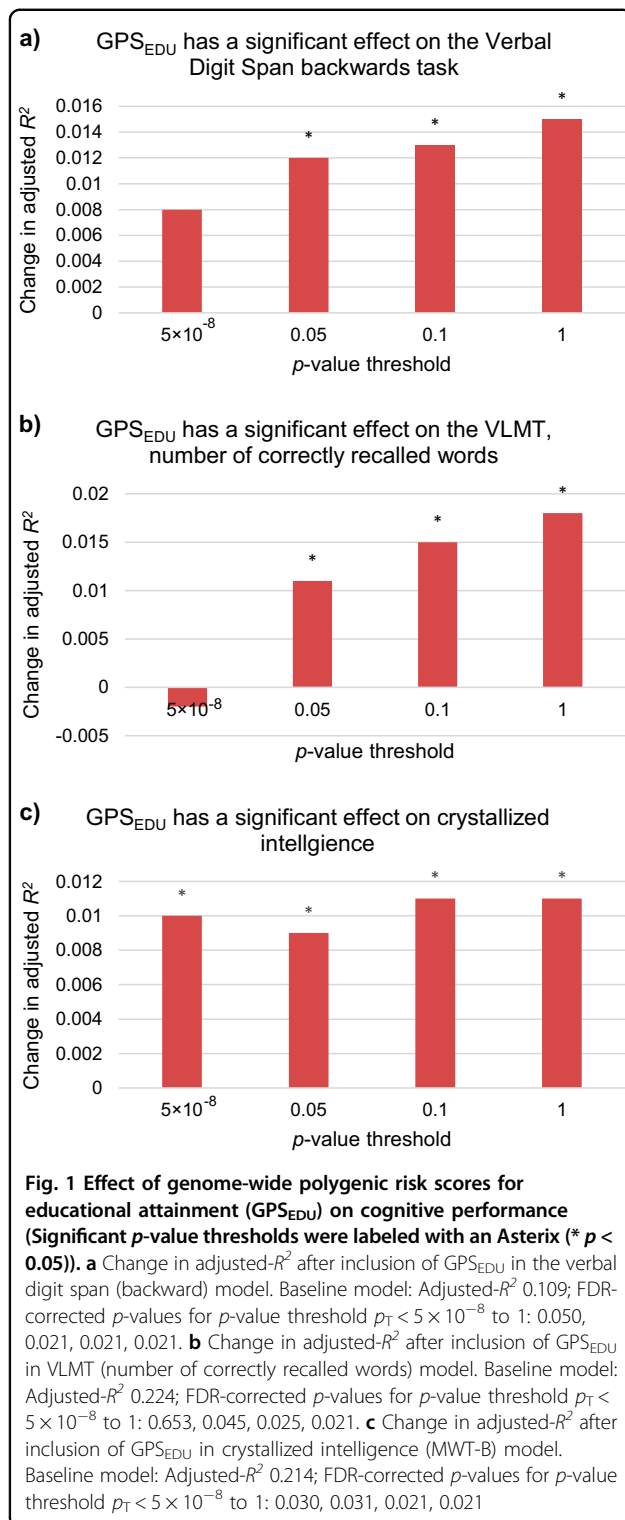
Table 1 Sample characteristics

	Cases ($n = 730$) ^b	Controls ($n = 320$) ^b	Test statistic	Degrees of freedom (df)	p -value
Age at baseline	43.19 (13.01)	37.53 (15.83)	5.62	516.11	<0.001
Sex			24.22	1	<0.001
Male	414 (56.7)	128 (40.0)			
Diagnosis		N/A	N/A	N/A	N/A
Schizophrenia	337 (46.2)				
Schizoaffective	73 (10.0)				
Bipolar-I disorder	256 (35.1)				
Bipolar-II disorder	64 (8.7)				
Education^a			154.11	6	<0.001
0	10 (1.4)	0 (0.0)			
1	46 (6.3)	2 (0.6)			
2	146 (20.0)	8 (2.5)			
3	179 (24.5)	98 (30.6)			
4	130 (17.8)	31 (9.7)			
5	87 (11.9)	35 (10.9)			
6	114 (15.6)	142 (44.4)			
Missing	18 (2.5)	4 (1.3)			
Duration of illness	12.93 (10.81)	N/A	N/A	N/A	N/A
Baseline treatment		N/A	N/A	N/A	N/A
None	23 (3.2)				
Outpatient	355 (48.6)				
Day patient	38 (5.2)				
Inpatient	310 (42.5)				
Missing	4 (0.5)				

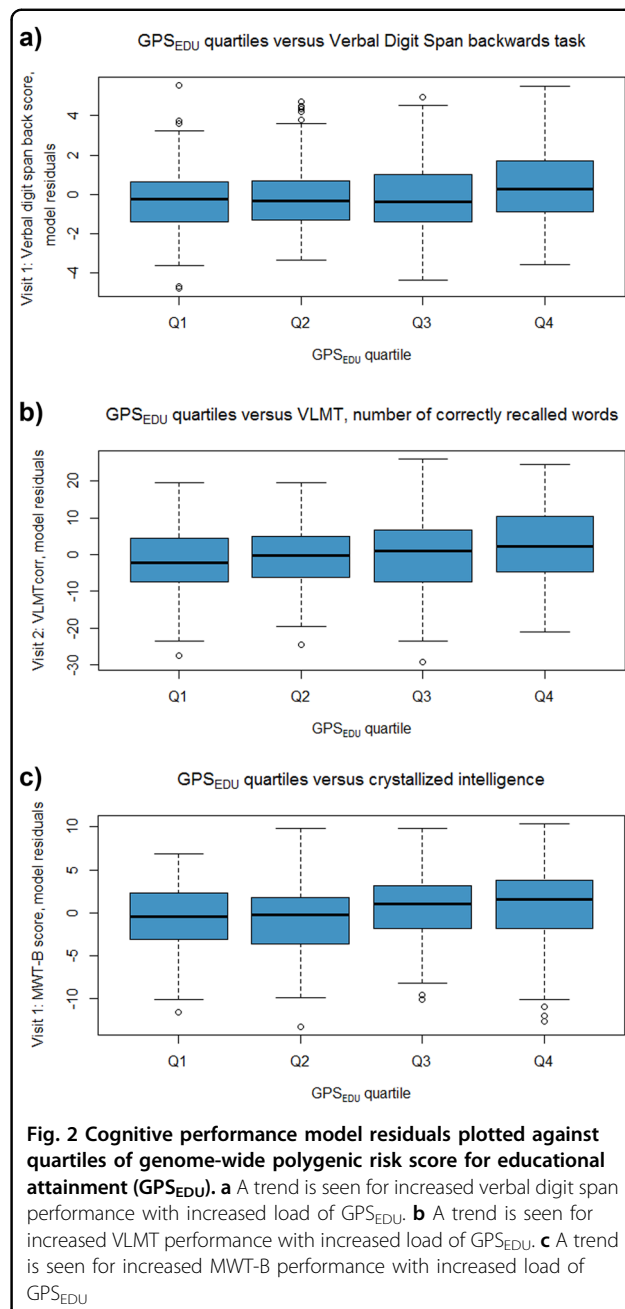
^aThe PsyCourse study measures status in the German educational system in detail. In order to make the German educational system comparable to English-speaking systems information on specialized schools, high school and professional education in Germany have been combined to form an ordinal educational scale with "6" being the highest level of education obtained

^bAge and duration of illness have been reported as mean (standard deviation), while all other categorical variables have been reported as n (%). A t -test was used for comparison of mean age and χ^2 -tests were used for all categorical comparisons

Socio-demographic information of participants



correctly recalled words (Fig. 1b) and 1.1% for crystallized intelligence (Fig. 1c). With more stringent *p*-value thresholds used, i.e., the inclusion of less SNPs, the change in adjusted-*R*² decreased. For the verbal digit span (backward) and the VLMT, the GPS_{EDU} based on the



p-value thresholds *p*_T < 0.05, 0.1, and <1 were significant (FDR-adjusted *p* < 0.05; Supplementary Table 3). The score was significant at all *p*-value thresholds for crystallized intelligence (FDR-adjusted *p* < 0.05). The examination of model residuals via quantile–quantile (QQ) plots did not show any extreme deviation from normality (Supplementary Figs. 3–5). Further inspection of model residuals against GPS_{EDU} quartiles showed evidence of increased performance on all three domains with increased GPS_{EDU} scores (Fig. 2). Our results remained robust after correcting for medication (Supplementary

Table 4) and diagnosis (Supplementary Table 5). Furthermore, our mediation analysis supports a robust association between GPS_{EDU} and crystallized intelligence that is not mediated by memory parameters (GPS_{EDU} $p < 0.05$; change in adjusted- $R^2 = 0.0091$).

No significant associations between cognitive outcomes and polygenic scores for schizophrenia or bipolar disorder were observed (Supplementary Table 6). Furthermore, neither risk score influenced the significant effects of GPS_{EDU} on the three cognitive domains reported above (Supplementary Table 7). Multicollinearity diagnostics showed no issues of collinearity in our regression analysis (variance inflation factor < 5 for all independent variables).

Discussion

Our study aimed to identify the influence of GPS_{EDU} on several cognitive domains in a transdiagnostic cohort of psychiatric patients. Confirming results of previous research, patients with bipolar disorder and schizophrenia in the PsyCourse cohort performed worse on tests of neurocognitive functioning in comparison to nonclinical controls. In patients, we observed a significant improvement in prediction of cognitive performance with inclusion of GPS_{EDU} for the backward verbal digit span, VLMT (correctly recalled words), and for crystallized intelligence. These findings confirm the ability of GPS_{EDU} to explain variability in linguistic cognitive performance related to working memory and learning in patients with known cognitive deficits. Furthermore, our findings show that cognitive performance measured for these domains were associated with the genetic underpinnings of GPS_{EDU} and not confounded by or associated with GPS_{SZ} or GPS_{BD} .

Previous studies have investigated the association between cognitive performance and GPS_{EDU} using summary statistics from an earlier GWAS on educational attainment by Okbay et al.²⁴. Our findings compliment earlier evidence supporting an association between cognitive performance and educational attainment, but not schizophrenia genetic risk, in clinical patients. For example, a study by Shafee et al. compared the effect of GPS_{SZ} on three cognitive phenotypes i.e. general cognitive function, premorbid intellectual potential, and years of education completed³⁰. The authors found that among healthy individuals, GPS_{SZ} was significantly associated with lower general cognitive functioning, however, found no association between GPS_{SZ} with any cognitive phenotype in patients with psychosis. Furthermore, the authors found significant positive correlations between GPS_{EDU} and both educational attainment and premorbid intelligence in patients with and without psychosis. Another study by Bansal et al. showed GPS_{EDU} could predict 2.09% of variance in premorbid IQ in a large schizophrenia sample²⁹. Our findings support earlier suggestions that different cognitive phenotypes vary in

their etiologic relationship with schizophrenia and in their genetic overlap with educational attainment³⁰. Furthermore, our findings are in line with evidence from the first educational attainment GWAS of 126,559 individuals which identified variants which implicated genes (including *BSN*, *GBX2*, *LRRN2*, and *PIK3C2B*) linked to processes such as learning and long-term memory²⁷. These findings are especially interesting given that learning and working memory are among some of the most impaired cognitive process for patients with psychiatric disorders⁵⁷.

While a polygenic score for educational attainment in the general population explained 7–10% of variance in cognitive performance, the score explained at most ~2% in our transdiagnostic cohort²². It is difficult to determine whether the smaller effect in our cohort was the result of a different phenotype being measured, i.e., specific cognitive domains and not a composite score, or whether this might reflect the cognitive performance of this unique, transdiagnostic sample being related to other complex genetic-environmental factors. Clearly, future investigations looking at other measures of cognition in large cohorts are warranted. Confounding variables such as acute symptoms may also contribute to the lack of variability explained in this case, although we have tried to capture this by controlling for current in/outpatient status and symptom severity. Furthermore, although based on big samples, polygenic scores “may not be sufficiently powerful to capture signs of disrupted neurodevelopment” in these patients as they exclude rare copy number variations and deleterious exonic mutations which may have important consequences⁵².

On both a phenotypic and genetic level, intelligence has been associated with psychiatric disorders. For example, individuals with a level of intelligence one standard deviation below the mean, have ~60% higher risk of hospitalization for schizophrenia⁵⁸. There is also evidence supporting an association between poorer school performance and higher risk for schizophrenia⁵². In addition, several longitudinal studies have linked deficits in premorbid IQ with subsequent schizophrenia development, which was also shown for mood disorders^{52,59}. The evidence, however, linking intelligence and affective disorders has been more inconsistent. For example, bipolar disorder has been associated with higher childhood IQ and an increased genetic risk of bipolar disorder has been associated with creativity and higher education^{60–62}. However, no such associations have been reported by studies of adolescent or adult IQ⁶⁰. Nevertheless, there are known genetic variants influencing both intelligence and psychiatric disorders which, in part, explain the phenotypic link between intelligence and these disorders⁵⁸.

We investigated the potential influence of GPS_{SZ} and GPS_{BD} on cognitive performance. These relationships

seem to be complex and while the genetic overlap between schizophrenia susceptibility with cognitive performance has been widely investigated in the literature with conflicting findings, less has been done in bipolar disorder^{8,63–65}. The lack of an association observed between either the GPS_{SZ} or GPS_{BD} with cognitive performance in our study emphasizes several issues inherent to these types of investigations. The first is that GPS_{EDU} is based on a much larger discovery sample than GPS_{SZ} and GPS_{BD}, meaning GPS_{EDU} had higher statistical power to capture smaller effect sizes and more accurate estimates for single SNPs of which the score is based on. Presuming the most optimistic estimate for variance explained in cognitive performance by the GPS_{SZ} of 1.6% that has previously been reported⁸, a sample of ~500 participants would be required to drive the effect of schizophrenia genetic risk scores on cognitive performance. However, given a more conservative estimate of 0.3%⁶⁶ variance explained, a sample size of over 2 600 participants would be required, suggesting that power may indeed be an issue in our study (Supplementary Fig. 6). This is also true with regards to GPS_{BD} in which genetic effects are likely to be at least as subtle. This again highlights the value of analyzing a proxy-phenotype such as educational attainment.

The second issue is in relation to the cognitive domains that were analyzed. As studies often use different cognitive tests from the wide variety that are available, it could be that the genetic risk of schizophrenia and bipolar disorder are more closely linked to domains that went unmeasured in our study. Perhaps if we had used a composite score across all domains or different neurocognitive tests in general, a significant effect would have been observed. Unfortunately, due to the longitudinal nature of our study which led to missing data across the different cognitive outcomes tested, a composite score analysis with adequate power was not feasible.

Our findings should be considered in light of a few limitations. The first is that our patients represent a chronic sample of heterogeneously treated patients. As these patients have been prescribed a wide range of medications at different dosages, correcting for the possible influence of medication is not an easy task. Not knowing how different drugs might interact with or influence cognition throughout the course of the disorder is a limitation that always must be considered in psychiatric research, and this problem has yet to have a perfect solution. A second limitation of our study is generalizability considering we investigated raw scores for several cognitive domains. As mentioned above, one of the major issues in the field at this time is the complexity in measuring this phenotype and with a plethora of tests that can be used, it is difficult to say how generalizable our findings are to other cognitive tests within the same cognitive domain in different cohorts. For example, while

crystallized intelligence was measured, our study failed to consider fluid intelligence which has a higher heritability component than crystallized intelligence⁶⁷. It is also important to note that while executive functions are related, they are also diverse⁶⁸. While the TMT used in our study is a measure of task switching, other executive functions like the updating process of working memory and inhibition should be explored. Lastly, we must acknowledge that our study has only assessed linguistic memory and not visuospatial memory. As these are two unique types of memory⁶⁹, future investigations are warranted to determine how the two might differ in association with GPS_{EDU}.

Although remarkable heterogeneity of cognitive deficits exists among individuals with psychiatric disorders, in general these deficits are, by a moderate degree, less severe in chronic bipolar patients in comparison to chronic schizophrenia patients. Furthermore, the trajectories of these impairments are quite different⁷⁰. Often, cognitive deficits are apparent before the onset of disease in individuals with schizophrenia⁷¹. Approximately 70% of bipolar patients exhibit cognitive deficits, especially related to verbal memory and attention⁵⁷, which often manifest in young adults⁶⁰. Despite these known differences for bipolar disorder and schizophrenia, we did not observe a significant effect of diagnosis on the effect of GPS_{EDU} related to cognitive performance. These diagnostic differences were most likely captured by the PANSS sum scores included in our models, which was highly significant. Evidence also supports an increase in the heritable component of intelligence with age⁷². Considering this knowledge, future studies, longitudinal in design, would be highly beneficial. It would be intriguing to see how the polygenic score for educational attainment can explain variability in cognitive performance throughout the course of the disorder. While our sample consisted of chronic mid-aged patients in which cognitive performance was rather stable across visits, it would be valuable to investigate younger cohorts of patients, even before the onset of disease, to determine how instability in cognitive performance throughout the disease course might influence the association between GPS_{EDU} and cognition. This would help determine at which points the underlying genetic components are most influential and help identify at which periods environmental influences might be more prominent in determining cognitive abilities.

Conclusions

Identifying a genetic component related to distinct neurocognitive profiles has potential to identify a more burdened subgroup of patients that in turn might be at risk for lower levels of functioning and poor social outcomes. This sort of information targets patients for more personalized interventions^{73,74}. Here we have explained

only a small fraction of variance in cognitive performance in patients with psychiatric disorders using the genetic variants associated with educational attainment. These findings highlight the importance of other uncaptured environmental exposures that have major influences on cognitive abilities and ultimately levels of functioning in these patients. Future studies, over the course of the disorder, would be informative to determine how this association changes over time, and at which periods environment may play the most influential role⁶⁰. Furthermore, future studies should factor in the complex pleiotropic relationships between these traits to generate enhanced polygenic scores to further clarify their genetic architecture⁷⁵. Moreover, hypothesis-based polygenic scores could help uncover biological pathways related to cognitive performance.

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Conflict of interest

Jens Reimer received honoraria from Otsuka-Lundbeck for participation in a speakers' bureau. The remaining authors declare that they have no conflict of interest.

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References

- Solé, B. et al. Cognitive impairment in bipolar disorder: treatment and prevention strategies. *Int. J. Neuropsychopharmacol.* **20**, 670–680 (2017).
- Sanchez, M., Bauer, I. E., Galvez, J. F., Zunta-Soares, G. B. & Soares, J. C. The management of cognitive impairment in bipolar disorder: current status and perspectives. *Am. J. Ther.* **22**, 477–486 (2015).
- Tripathi, A., Kar, S. K. & Shukla, R. Cognitive Deficits in Schizophrenia: Understanding the Biological Correlates and Remediation Strategies. *Clin. Psychopharmacol. Neurosci.* **16**, 7–17 (2018).
- Goff, D. C., Hill, M. & Barch, D. The treatment of cognitive impairment in schizophrenia. *Pharmacol. Biochem. Behav.* **99**, 245–253 (2011).
- Green, M. F. What are the functional consequences of neurocognitive deficits in schizophrenia? *Am. J. Psychiatry* **153**, 321–330 (1996).
- Bowie, C. R. & Harvey, P. D. Cognitive deficits and functional outcome in schizophrenia. *Neuropsychiatr. Dis. Treat.* **2**, 531–536 (2006).
- Davies, G. et al. Genome-wide association studies establish that human intelligence is highly heritable and polygenic. *Mol. Psychiatry* **16**, 996–1005 (2011).
- Lenz, T. et al. Molecular genetic evidence for overlap between general cognitive ability and risk for schizophrenia: a report from the Cognitive Genomics Consortium (COGENT). *Mol. Psychiatry* **19**, 168–174 (2014).
- Benyamin, B. et al. Childhood intelligence is heritable, highly polygenic and associated with FBNP1L. *Mol. Psychiatry* **19**, 253–258 (2014).
- Kirkpatrick, R. M., McGue, M., Iacono, W. G., Miller, M. B. & Basu, S. Results of a "GWAS plus" general cognitive ability is substantially heritable and massively polygenic. *PLoS ONE* **9**, e112390 (2014).

11. Davies, G. et al. Study of 300,486 individuals identifies 148 independent genetic loci influencing general cognitive function. *Nat. Commun.* **9**, 2098 (2018).
12. Deary, I. J. The stability of intelligence from childhood to old age. *Curr. Dir. Psychol. Sci.* **23**, 239–245 (2014).
13. Deary, I. J. et al. Genetic contributions to stability and change in intelligence from childhood to old age. *Nature* **482**, 212–215 (2012).
14. Lyons, M. J. et al. A longitudinal twin study of general cognitive ability over four decades. *Dev. Psychol.* **53**, 1170–1177 (2017).
15. Snitz, B. E., Macdonald, A. W. 3rd & Carter, C. S. Cognitive deficits in unaffected first-degree relatives of schizophrenia patients: a meta-analytic review of putative endophenotypes. *Schizophr. Bull.* **32**, 179–194 (2006).
16. Bora, E., Yucel, M. & Pantelis, C. Cognitive endophenotypes of bipolar disorder: a meta-analysis of neuropsychological deficits in euthymic patients and their first-degree relatives. *J. Affect. Disord.* **113**, 1–20 (2009).
17. Gottesman, I. I. & Gould, T. D. The endophenotype concept in psychiatry: etymology and strategic intentions. *Am. J. Psychiatry* **160**, 636–645 (2003).
18. Plomin, R. & von Stumm, S. The new genetics of intelligence. *Nat. Rev. Genet.* **19**, 148–159 (2018).
19. Deary, I. J., Johnson, W. & Houlihan, L. M. Genetic foundations of human intelligence. *Hum. Genet.* **126**, 215–232 (2009).
20. Trampush, J. W. et al. GWAS meta-analysis reveals novel loci and genetic correlates for general cognitive function: a report from the COGENT consortium. *Mol. Psychiatry* **22**, 336–345 (2017).
21. Savage, J. E. et al. Genome-wide association meta-analysis in 269,867 individuals identifies new genetic and functional links to intelligence. *Nat. Genet.* **50**, 912–919 (2018).
22. Lee, J. J. et al. Gene discovery and polygenic prediction from a genome-wide association study of educational attainment in 1.1 million individuals. *Nat. Genet.* **50**, 1112–1121 (2018).
23. Davies, G. et al. Genome-wide association study of cognitive functions and educational attainment in UK Biobank (N=112,151). *Mol. Psychiatry* **21**, 758–767 (2016).
24. Okbay, A. et al. Genome-wide association study identifies 74 loci associated with educational attainment. *Nature* **533**, 539–542 (2016).
25. Rietveld, C. A. et al. Common genetic variants associated with cognitive performance identified using the proxy-phenotype method. *Proc. Natl Acad. Sci. USA* **111**, 13790–13794 (2014).
26. Trampush, J. W. et al. Independent evidence for an association between general cognitive ability and a genetic locus for educational attainment. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **168B**, 363–373 (2015).
27. Rietveld, C. A. et al. GWAS of 126,559 individuals identifies genetic variants associated with educational attainment. *Science* **340**, 1467–1471 (2013).
28. Elliott, M. L. et al. A polygenic score for higher educational attainment is associated with larger brains. *Cereb. Cortex*. 2018; <https://doi.org/10.1093/cercor/bhy219>.
29. Bansal, V. et al. Genome-wide association study results for educational attainment aid in identifying genetic heterogeneity of schizophrenia. *Nat. Commun.* **9**, 3078 (2018).
30. Shafee, R. et al. Polygenic risk for schizophrenia and measured domains of cognition in individuals with psychosis and controls. *Transl. Psychiatry* **8**, 78 (2018).
31. Budde, M. et al. A longitudinal approach to biological psychiatric research: The PsyCourse study. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **180**, 89–102 (2019).
32. Forstner, A. J. et al. Identification of shared risk loci and pathways for bipolar disorder and schizophrenia. *PLoS ONE* **12**, e0171595 (2017).
33. Anttila, V. et al. Analysis of shared heritability in common disorders of the brain. *Science* **360**, 6395 (2018).
34. Lichtenstein, P. et al. Common genetic determinants of schizophrenia and bipolar disorder in Swedish families: a population-based study. *Lancet* **373**, 234–239 (2009).
35. Lee, P. H. et al. Genome wide meta-analysis identifies genomic relationships, novel loci, and pleiotropic mechanisms across eight psychiatric disorders. *bioRxiv* 2019; 528117.
36. Hill, W. D. et al. A combined analysis of genetically correlated traits identifies 187 loci and a role for neurogenesis and myelination in intelligence. *Mol. Psychiatry* **24**, 169–181 (2018).
37. American Psychiatric Association. Diagnostic and statistical manual of mental disorders (4th ed): Washington, DC, 2002.
38. Kay, S. R., Fiszbein, A. & Opler, L. A. The positive and negative syndrome scale (PANSS) for schizophrenia. *Schizophr. Bull.* **13**, 261–276 (1987).
39. Brown, R. E. Hebb and Cattell: The Genesis of the Theory of Fluid and Crystallized. *Intell. Front. Hum. Neurosci.* **10**, 606 (2016).
40. Lehl, S. *Mehrfachwahl-Wortschatz-Intelligenztest (MWT-B)*. Spitta Verlag: Baltingen, Germany, 2005.
41. Partington, J. E. & Leiter, R. G. Partington's Pathways Test. *Psychol. Serv. Cent. J.* **1**, 11–20 (1949).
42. Strauss, E., Sherman, E. M. & Spreen, O. *A compendium of neuropsychological tests: Administration, norms, and commentary*. 3 edn (Oxford University Press, New York, 2006).
43. Lamberty, G. J. et al. Derived Trail Making Test indices: A preliminary report. *Neuropsychiatry Neuropsychol. Behav. Neurol.* **7**, 230–234 (1994).
44. Aster, M., Neubauer, A. & Horn, R. *Wechsler Intelligenztest für Erwachsene. Wechsler Intelligence Test for Adults (German revision and adaptation of the WAIS-III of David Wechsler)*. Harcourt Test Services: Frankfurt, Germany, 2006.
45. Wechsler, D. *Manual for the Wechsler Adult Intelligence Scale*. Psychological Corp.: New York, 1955, vi, 110–vi, 110pp.
46. Lezak, M. D. *Neuropsychological Assessment (2nd ed.)*. Oxford University Press: New York, 1983.
47. Helmstaedter, C., Lendt, M. & Lux, S. Verbaler Lern- und Merkfähigkeitstest (VLMT). Beltz: Göttingen, Germany, 2001.
48. Kalman, J. L. et al. Investigating polygenic burden in age at disease onset in bipolar disorder: Findings from an international multicentric study. *Bipolar Disord.* **21**, 68–75 (2019).
49. Chang, C. C. et al. Second-generation PLINK: rising to the challenge of larger and richer datasets. *Gigascience* **4**, 7 (2015).
50. Pardiñas, A. F. et al. Common schizophrenia alleles are enriched in mutation-intolerant genes and in regions under strong background selection. *Nat. Genet.* **50**, 381–389 (2018).
51. Stahl, E. A. et al. Genome-wide association study identifies 30 loci associated with bipolar disorder. *Nat. Genet.* **51**, 793–803 (2019).
52. Sørensen, H. J. et al. Polygenic risk scores, school achievement, and risk for schizophrenia: a danish population-based study. *Biol. Psychiatry* **84**, 684–691 (2018).
53. R Core Team (2013). R: A language and environment for statistical computing. R Foundation for Statistical Computing: Vienna, Austria.
54. Tukey, J. W. *Exploratory data analysis*. Addison-Wesley Pub. Co.: Reading, Mass., 1977.
55. Price, A. L. et al. Principal components analysis corrects for stratification in genome-wide association studies. *Nat. Genet.* **38**, 904–909 (2006).
56. Unsworth, N. On the division of working memory and long-term memory and their relation to intelligence: a latent variable approach. *Acta Psychol.* **134**, 16–28 (2010).
57. Barbosa, I. G. et al. Predictors of cognitive performance in bipolar disorder: the role of educational degree and inflammatory markers. *J. Psychiatr. Res.* **106**, 31–37 (2018).
58. Hill, W. D., Harris, S. E. & Deary, I. J. What genome-wide association studies reveal about the association between intelligence and mental health. *Curr. Opin. Psychol.* **27**, 25–30 (2018).
59. Peyrot, W. J. et al. The association between lower educational attainment and depression owing to shared genetic effects? Results in ~25,000 subjects. *Mol. Psychiatry* **20**, 735–743 (2015).
60. Mistry, S., Harrison, J. R., Smith, D. J., Escott-Price, V. & Zammit, S. The use of polygenic risk scores to identify phenotypes associated with genetic risk of bipolar disorder and depression: a systematic review. *J. Affect. Disord.* **234**, 148–155 (2018).
61. Vreeker, A. et al. High educational performance is a distinctive feature of bipolar disorder: a study on cognition in bipolar disorder, schizophrenia patients, relatives and controls. *Psychol. Med.* **46**, 807–818 (2016).
62. MacCabe, J. H. et al. Excellent school performance at age 16 and risk of adult bipolar disorder: national cohort study. *Br. J. Psychiatry* **196**, 109–115 (2010).
63. Schaupp, S., Schulze, T. & Budde, M. Let's talk about the association between schizophrenia polygenic risk scores and cognition in patients and the general population: a review. *J. Psychiatry Brain Sci.* **3**, 12 (2018).
64. Mistry, S., Harrison, J. R., Smith, D. J., Escott-Price, V. & Zammit, S. The use of polygenic risk scores to identify phenotypes associated with genetic risk of schizophrenia: Systematic review. *Schizophr. Res.* **S0920-9964**, 30665–30665 (2017). pii.

65. Rønlund, S. et al. A polygenic risk score analysis of psychosis endophenotypes across brain functional, structural, and cognitive domains. *Am. J. Med. Genet. B Neuropsychiatr. Genet.* **177**, 21–34 (2018).
66. Riglin, L. et al. Schizophrenia risk alleles and neurodevelopmental outcomes in childhood: a population-based cohort study. *Lancet Psychiatry* **4**, 57–62 (2017).
67. Cattell, R. B., Schuberger, J. M., Ahern, F. M. & Kameoka, V. The heritability of fluid and crystallized intelligences: By the mava design and oses analysis. *Aust. J. Psychol.* **33**, 355–374 (1981).
68. Miyake, A. et al. The unity and diversity of executive functions and their contributions to complex “Frontal Lobe” tasks: a latent variable analysis. *Cogn. Psychol.* **41**, 49–100 (2000).
69. Baddeley, A. Working memory: looking back and looking forward. *Nat. Rev. Neurosci.* **4**, 829–839 (2003).
70. Bortolato, B., Miskowiak, K. W., Köhler, C. A., Vieta, E. & Carvalho, A. F. Cognitive dysfunction in bipolar disorder and schizophrenia: a systematic review of meta-analyses. *Neuropsychiatr. Dis. Treat.* **11**, 3111–3125 (2015).
71. Reichenberg, A. et al. Static and dynamic cognitive deficits in childhood preceding adult schizophrenia: a 30-year study. *Am. J. Psychiatry* **167**, 160–169 (2010).
72. Plomin, R. & Deary, I. J. Genetics and intelligence differences: five special findings. *Mol. Psychiatry* **20**, 98–108 (2014).
73. Tickell, A. M. et al. Neurocognitive clusters: a pilot study of young people with affective disorders in an inpatient facility. *J. Affect. Disord.* **242**, 80–86 (2019).
74. Kapur, S., Phillips, A. G. & Insel, T. R. Why has it taken so long for biological psychiatry to develop clinical tests and what to do about it? *Mol. Psychiatry* **17**, 1174–1179 (2012).
75. Lam, M. et al. Pleiotropic meta-analysis of cognition, education, and schizophrenia differentiates roles of early neurodevelopmental and adult synaptic pathways. *bioRxiv* 2019: 519967.

2.2 Original Article 1: Supplementary material

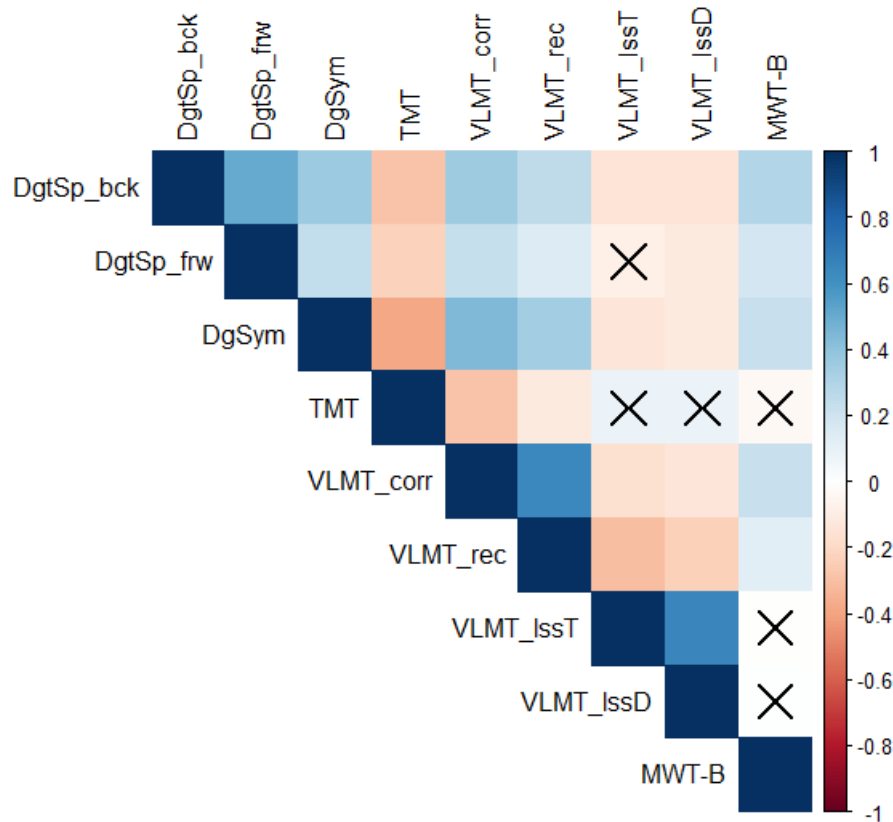


Figure S1. Pairwise correlations between cognitive outcomes in cases only. Shown are Pearson correlation coefficients, insignificant correlations labeled with “X” (uncorrected $p > 0.05$). *Note:* DgtSp_bck – Verbal digit span (backwards task); DgtSp_frwr – Verbal digit span (forwards task); DgSym – Digit symbol test; TMT – TMT reaction time difference; VLMT_corr – VLMT number of correctly recalled words; VLMT_issT – VLMT loss of recalled words after time; VLMT_issD – VLMT loss of recalled words after distraction; VLMT_rec – VLMT number of correctly recognized words

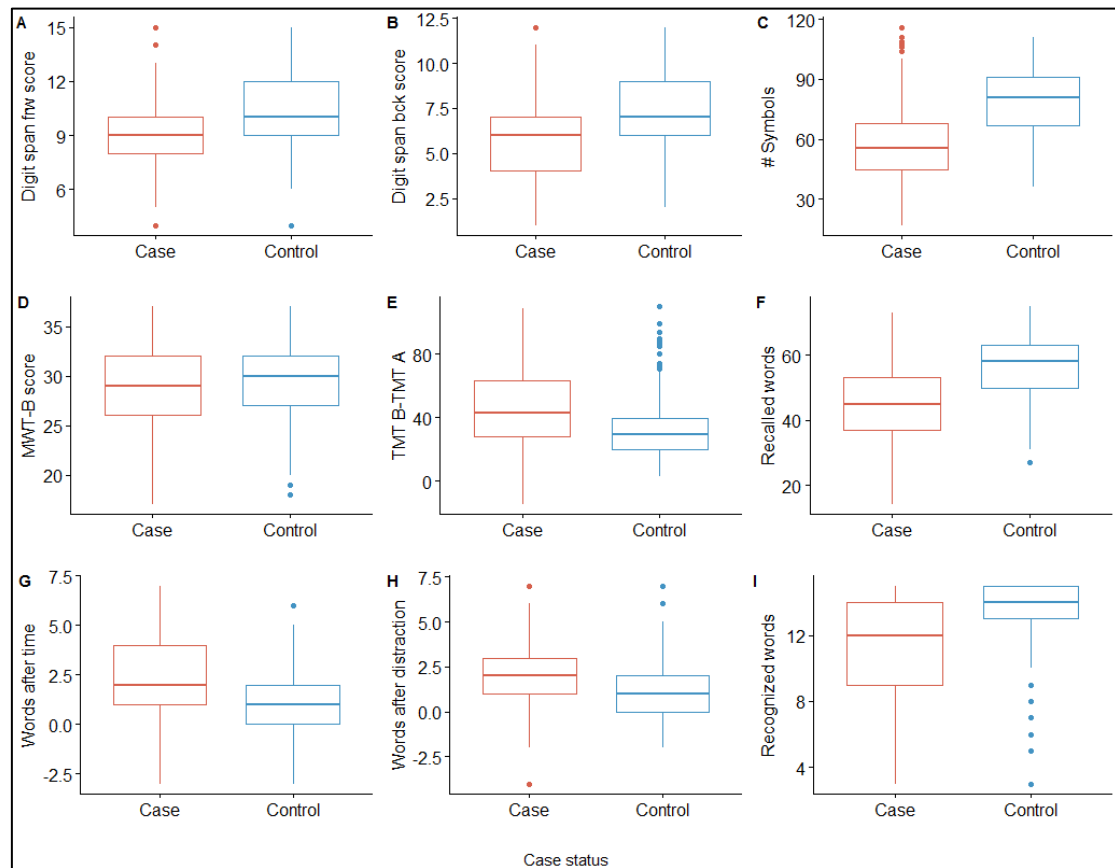


Figure S2. Boxplots depicting the distribution of cognitive performance for cases and controls across all cognitive domains. **A.** Verbal digit span, forward task score **B.** Verbal digit span, backward task score **C.** Digit symbol test, number of correct symbols **D.** MWT-B score **E.** Trail-Making-Test (TMT), difference in reaction time (TMT B-TMT A) **F.** Verbal Learning and Memory Test (VLMT): number of correctly recalled words **G.** VLMT: number of words lost after time **H.** VLMT: number of words lost after distraction **I.** VLMT: number of correctly recognized word

Table S1. The effect of case status on cognitive performance

Outcome	Cases mean (sd)	Controls mean (sd)	Estimate	Std. Error	T- value	p-value
Visit 1						
TMT reaction time difference	47.78 (24.80)	32.10 (17.91)	-12.55	1.49	-8.43	$< 2.00 \times 10^{-16}$ ***
Digit symbol test	56.37 (15.79)	79.03 (15.91)	18.39	1.03	17.86	$< 2.00 \times 10^{-16}$ ***
Verbal digit span backwards task	5.72 (1.95)	7.25 (2.14)	1.40	0.14	9.87	$< 2.00 \times 10^{-16}$ ***
Verbal digit span forwards task	9.01 (1.94)	10.32 (2.06)	1.18	0.15	8.15	1.13×10^{-15} ***
MWT-B (crystallized intelligence)	28.34 (4.56)	29.63 (3.61)	1.67	0.33	5.05	5.90×10^{-7} ***
Visit 2						
VLMT: correctly recalled words	44.99 (11.57)	56.35 (9.24)	8.87	0.83	10.70	$< 2 \times 10^{-16}$ ***
VLMT: loss of words after time	2.21 (2.01)	1.22 (1.77)	-0.80	0.16	-5.02	6.81×10^{-07} ***
VLMT: loss of words after distraction	2.07 (1.82)	1.26 (1.65)	-0.66	0.15	-4.41	1.21×10^{-05} ***
VLMT: correctly recognized words	11.28 (3.46)	13.44 (2.13)	1.45	0.23	6.20	1.06×10^{-09} ***

Note: Effect estimates and associated standard errors, T-values and p-values adjusted for age and sex

Table S2. The effect of GPS_{EDU} quartile on level of educational attainment in cases, adjusting for age, sex, the interaction between age and sex, and the first 10 principle components

	Proportional odds ratio	95% CI	p-value
GPS _{EDU} quartile 2	1.478	1.013 - 2.159	0.043*
GPS _{EDU} quartile 3	1.794	1.237 - 2.606	0.002*
GPS _{EDU} quartile 4	2.495	1.706 - 3.657	2.56×10 ⁻⁶ ***
Age (years)	0.866	0.695 - 1.078	0.200
Sex (male)	0.728	0.555 - 0.956	0.022*
PC1	1.138	1.002 - 1.303	0.046*
PC2	1.137	0.995 - 1.300	0.057
PC3	0.910	0.787 - 1.048	0.192
PC4	1.005	0.857 - 1.174	0.952
PC4	0.957	0.810 - 1.110	0.571
PC6	1.101	0.960 - 1.263	0.169
PC7	0.970	0.850 - 1.107	0.653
PC8	0.896	0.785 - 1.022	0.102
PC9	0.981	0.859 - 1.120	0.778
PC10	1.003	0.882 - 1.141	0.959
Age (years):Sex (M)	1.952	1.473 - 2.590	3.36×10 ⁻⁶ ***

* $p < 0.05$ ** $p < 0.005$ *** $p < 0.005$

Table S3. Effect of GPS_{EDU} on cognitive performance models adjusted for age, age², sex, in/outpatient status, center, PANSS sum scores, principle components

Model	GPS <i>p</i> -value threshold	<i>p</i> -value	FDR corrected <i>p</i> -value	Base model adjusted <i>R</i> ²	Adjusted <i>R</i> ² after GPS _{EDU} inclusion
Visit 1					
V1: Verbal digit span backwards task				0.109	
	1	0.001	0.021		0.124
	0.1	0.002	0.021		0.122
	0.05	0.004	0.021		0.121
	5×10 ⁻⁸	0.015	0.050		0.117
V1: Verbal digit span forwards task				0.066	
	1	0.477	0.553		0.066
	0.1	0.313	0.470		0.066
	0.05	0.384	0.522		0.065
	5×10 ⁻⁸	0.435	0.553		0.065
V1: Digit symbol test				0.292	
	1	0.206	0.417		0.293
	0.1	0.241	0.429		0.293
	0.05	0.469	0.553		0.292
	5×10 ⁻⁸	0.765	0.787		0.291
V1: MWT-B				0.214	
	1	0.003	0.021		0.225
	0.1	0.003	0.021		0.225
	0.05	0.008	0.030		0.223
	5×10 ⁻⁸	0.007	0.030		0.224
V1: TMT B-TMT A				0.091	
	1	0.454	0.464		0.091
	0.1	0.611	0.417		0.090
	0.05	0.560	0.417		0.090
	5×10 ⁻⁸	0.391	0.417		0.091
V2: VLMT correctly recalled words				0.224	
	1	0.002	0.021		0.243
	0.1	0.005	0.025		0.239
	0.05	0.013	0.045		0.235
	5×10 ⁻⁸	0.617	0.652		0.222
V2: VLMT loss of words after time				0.050	
	1	0.283	0.463		0.050
	0.1	0.3075	0.522		0.049
	0.05	0.296	0.464		0.050
	5×10 ⁻⁸	0.193	0.417		0.052
V2: VLMT loss of words after distraction				0.015	
	1	0.199	0.417		0.017
	0.1	0.220	0.417		0.017
	0.05	0.187	0.417		0.017
	5×10 ⁻⁸	0.035	0.104		0.025
V2: VLMT correctly recognized words				0.142	
	1	0.199	0.417		0.144
	0.1	0.210	0.417		0.144
	0.05	0.250	0.429		0.143
	5×10 ⁻⁸	0.936	0.936		0.139

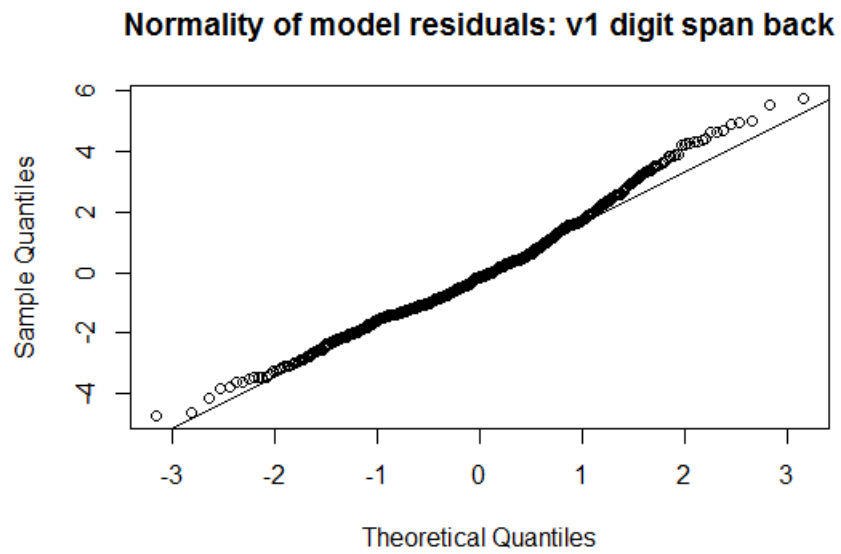


Figure S3. QQ plot for visual inspection of normality of model residuals – Visit 1 verbal digit span backward model

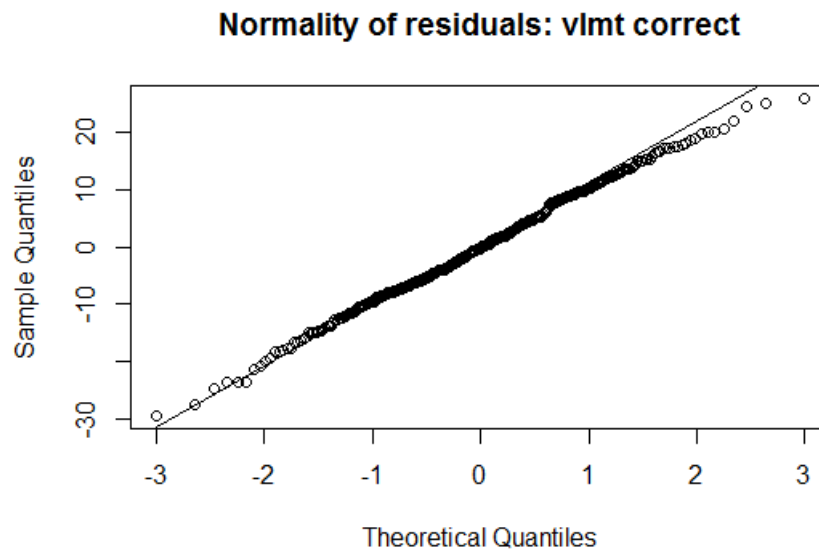


Figure S4. QQ plot for visual inspection of normality of residuals- Visit 2 VLMT correctly recalled words model

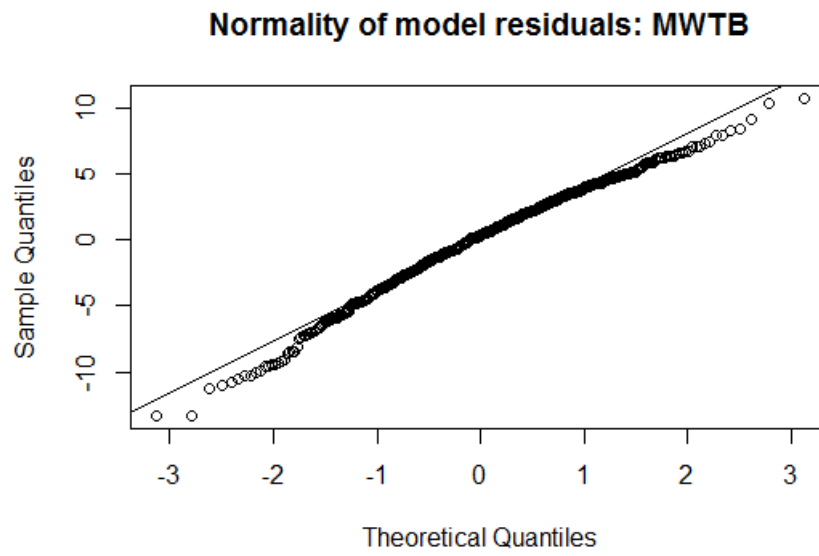


Figure S5. QQ plot for visual inspection of normality of residuals- Visit 1 MWT-B (crystallized intelligence) model

Table S4. Effect of GPS_{EDU} on cognitive performance models adjusted for age, age², sex, in/outpatient status, center, PANSS sum scores, principle components, and **medication**

Model	GPS <i>p</i> -value threshold	<i>p</i> -value	FDR corrected <i>p</i> -value	Base model adjusted <i>R</i> ²	Adjusted <i>R</i> ² after GPS _{EDU} inclusion
V1: Verbal digit span backwards task				0.109	
	1	0.001	0.010		0.124
	0.1	0.002	0.010		0.123
	0.05	0.003	0.014		0.121
	5×10 ⁻⁸	0.012	0.041		0.117
V1: Verbal digit span forwards task				0.066	
	1	0.387	0.478		0.066
	0.1	0.242	0.396		0.067
	0.05	0.300	0.446		0.066
	5×10 ⁻⁸	0.390	0.478		0.066
V1: Digit symbol test				0.301	
	1	0.184	0.346		0.302
	0.1	0.202	0.346		0.302
	0.05	0.398	0.478		0.301
	5×10 ⁻⁸	0.747	0.767		0.300
V1: MWT-B				0.213	
	1	0.001	0.010		0.226
	0.1	0.002	0.010		0.226
	0.05	0.004	0.015		0.224
	5×10 ⁻⁸	0.006	0.020		0.223
V1: TMT B-TMT A				0.098	
	1	0.526	0.592		0.097
	0.1	0.680	0.720		0.097
	0.05	0.623	0.680		0.097
	5×10 ⁻⁸	0.388	0.478		0.098
V2: VLMT correctly recalled words				0.253	
	1	0.001	0.010		0.276
	0.1	0.001	0.010		0.273
	0.05	0.004	0.015		0.269
	5×10 ⁻⁸	0.465	0.540		0.252
V2: VLMT loss of words after time				0.046	
	1	0.290	0.446		0.047
	0.1	0.391	0.478		0.046
	0.05	0.310	0.446		0.046
	5×10 ⁻⁸	0.141	0.338		0.050
V2: VLMT loss of words after distraction				0.014	
	1	0.182	0.346		0.016
	0.1	0.196	0.346		0.016
	0.05	0.166	0.346		0.017
	5×10 ⁻⁸	0.022	0.066		0.027
V2: VLMT correctly recognized words				0.148	
	1	0.125	0.321		0.152
	0.1	0.125	0.321		0.152
	0.05	0.151	0.340		0.151
	5×10 ⁻⁸	0.859	0.859		0.146

Table S5. Effect of GPS_{EDU} on cognitive performance models adjusted for age, age², sex, in/outpatient status, center, PANSS sum scores, principle components, and **diagnosis**

Model	GPS <i>p</i> -value threshold	<i>p</i> - value	FDR corrected <i>p</i> -value	Base model adjusted <i>R</i> ²	Adjusted <i>R</i> ² after GPS _{EDU} inclusion
V1: Verbal digit span backwards task				0.106	
	1	0.001	0.018		0.121
	0.1	0.002	0.018		0.119
	0.05	0.003	0.018		0.118
	5×10 ⁻⁸	0.015	0.050		0.114
V1: Verbal digit span forwards task				0.071	
	1	0.419	0.525		0.070
	0.1	0.262	0.450		0.071
	0.05	0.342	0.492		0.071
	5×10 ⁻⁸	0.365	0.506		0.071
V1: Digit symbol test				0.302	
	1	0.233	0.441		0.302
	0.1	0.290	0.474		0.302
	0.05	0.548	0.616		0.302
	5×10 ⁻⁸	0.725	0.746		0.302
V1: MWT-B				0.214	
	1	0.003	0.018		0.226
	0.1	0.003	0.018		0.226
	0.05	0.006	0.026		0.224
	5×10 ⁻⁸	0.006	0.026		0.224
V1: TMT B-TMT A				0.102	
	1	0.448	0.537		0.101
	0.1	0.603	0.638		0.100
	0.05	0.564	0.616		0.100
	5×10 ⁻⁸	0.409	0.525		0.101
V2: VLMT correctly recalled words				0.226	
	1	0.002	0.018		0.246
	0.1	0.004	0.019		0.242
	0.05	0.011	0.039		0.238
	5×10 ⁻⁸	0.559	0.616		0.224
V2: VLMT loss of words after time				0.054	
	1	0.316	0.492		0.054
	0.1	0.423	0.525		0.053
	0.05	0.331	0.492		0.054
	5×10 ⁻⁸	0.192	0.441		0.056
V2: VLMT loss of words after distraction				0.015	
	1	0.204	0.441		0.017
	0.1	0.231	0.441		0.017
	0.05	0.216	0.441		0.017
	5×10 ⁻⁸	0.037	0.111		0.025
V2: VLMT correctly recognized words				0.138	
	1	0.207	0.441		0.140
	0.1	0.222	0.441		0.140
	0.05	0.261	0.450		0.139
	5×10 ⁻⁸	0.893	0.893		0.135

Table S6. Effect of GPS_{BD} and GPS_{SZ} on cognitive performance, adjusted for age, age², sex, in/outpatient status, center, PANSS sum scores, and principle components

Model	Base model adjusted- R^2	p -value GPS _{SZ}	Adjusted R^2 after GPS _{SZ} inclusion	p -value GPS _{BD}	Adjusted- R^2 after GPS _{BD} inclusion
V1: Verbal digit span backwards task	0.109	0.268	0.109	0.312	0.109
V1: Verbal digit span forwards task	0.067	0.373	0.067	0.399	0.067
V1: Digit symbol test	0.291	0.838	0.290	0.786	0.290
V1: MWT-B	0.214	0.436	0.214	0.498	0.214
V1: TMT B-TMT A	0.093	0.174	0.094	0.208	0.094
V2: VLMT correctly recalled words	0.223	0.363	0.222	0.531	0.221
V2: VLMT words lost after time	0.038	0.399	0.037	0.050	0.046
V2: VLMT words lost after distraction	0.019	0.826	0.016	0.456	0.018
V2: VLMT correctly recognized words	0.143	0.60	0.150	0.965	0.140

Table S7. Effect of GPS_{BD} and GPS_{SZ} on the association between GPS_{EDU} and cognitive performance

Model	<i>p</i> -value GPS _{EDU} after adjusting for GPS _{SZ} ^a	Change in adjusted- <i>R</i> ² after GPS _{EDU} inclusion ^a	<i>p</i> -value GPS _{EDU} after adjusting for GPS _{BD} ^b	Change in adjusted- <i>R</i> ² after GPS _{EDU} inclusion ^b
V1: Verbal digit span backwards task	0.002	0.013	0.001	0.014
V1: MWT-B	0.004	0.011	0.003	0.011
V2: VLMT correctly recalled words	0.002	0.020	0.002	0.019

^a Adjusting for age, age², sex, in/outpatient status, center, PANSS sum scores, principle components and GPS_{SZ}

^b Base model including age, age², sex, in/outpatient status, center, PANSS sum scores, principle components and GPS_{BD}

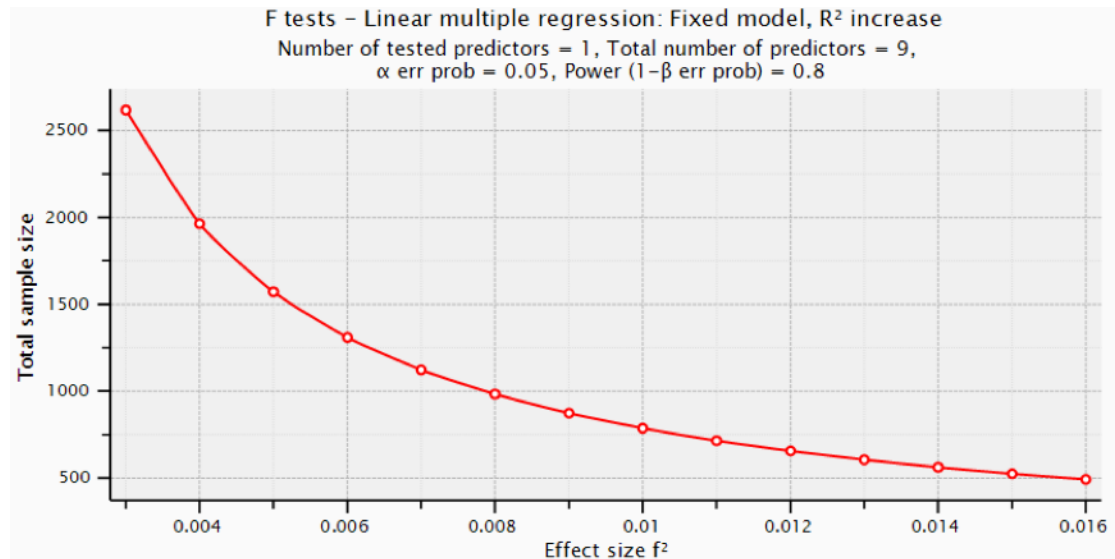


Figure S6. Post-hoc power calculation using the freely available *G* Power 3.1* (Faul, Erdfelder, Buchner, & Lang, 2009; Faul, Erdfelder, Lang, & Buchner, 2007). Plot depicting sample size required to drive the effect of GPS_{SZ} on cognitive performance to significance according to a range of effect sizes (change in adjusted R^2 between a model including only covariates and a model including covariates and GPS_{SZ}). *Note:* Alpha level 0.05, 80% power, number of tested predictors = 1, total number of predictors = 9.

Original Article 1: Supplementary references

- Faul, F., Erdfelder, E., Buchner, A., & Lang, A.-G. (2009). Statistical power analyses using g*power 3.1: Tests for correlation and regression analyses. *Behavior Research Methods*, 41(4), 1149-1160.
- Faul, F., Erdfelder, E., Lang, A.-G., & Buchner, A. (2007). G*power 3: A flexible statistical power analysis program for the social, behavioral, and biomedical sciences. *Behavior Research Methods*, 39(2), 175-191. doi:10.3758/bf03193146

3 Original Article 2

3.1 Individual contributions and reference

The study “The role of environmental stress and DNA methylation in the longitudinal course of bipolar disorder” was published in the *International Journal of Bipolar Disorders* in 2020. It was conducted under the supervision of T.G.S. and U.H. The research was designed by T.G.S. and A.L.C. Methylation data quality control and pre-processing was performed by A.L.C. with support from S.S. Data analysis was performed by A.L.C. with statistical guidance from U.H., D.C., and T.A. A.L.C. wrote the manuscript and accompanied the publication process as corresponding author. All co-authors critically revised and approved the manuscript.

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RESEARCH

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The role of environmental stress and DNA methylation in the longitudinal course of bipolar disorder

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Abstract

Background: Stressful life events influence the course of affective disorders, however, the mechanisms by which they bring about phenotypic change are not entirely known.

Methods: We explored the role of DNA methylation in response to recent stressful life events in a cohort of bipolar patients from the longitudinal PsyCourse study ($n = 96$). Peripheral blood DNA methylomes were profiled at two time points for over 850,000 methylation sites. The association between impact ratings of stressful life events and DNA methylation was assessed, first by interrogating methylation sites in the vicinity of candidate genes previously implicated in the stress response and, second, by conducting an exploratory epigenome-wide association analysis. Third, the association between epigenetic aging and change in stress and symptom measures over time was investigated.

Results: Investigation of methylation signatures over time revealed just over half of the CpG sites tested had an absolute difference in methylation of at least 1% over a 1-year period. Although not a single CpG site withstood correction for multiple testing, methylation at one site (cg15212455) was suggestively associated with stressful life events ($p < 1.0 \times 10^{-5}$). Epigenetic aging over a 1-year period was not associated with changes in stress or symptom measures.

Conclusions: To the best of our knowledge, our study is the first to investigate epigenome-wide methylation across time in bipolar patients and in relation to recent, non-traumatic stressful life events. Limited and inconclusive evidence warrants future longitudinal investigations in larger samples of well-characterized bipolar patients to give a complete picture regarding the role of DNA methylation in the course of bipolar disorder.

Keywords: DNA methylation, Bipolar disorder, Stressful life events, Longitudinal, Epigenomics, Epigenetic aging

Background

Bipolar disorder (BD) remains an interesting candidate for neurobiological analyses owing to its heterogeneous presentation and both genetic and environmental risk factors (Ludwig and Dwivedi 2016). While genome-wide

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association studies (GWAS) in BD have identified dozens of associated variants, they have explained only a small fraction of overall disease liability (Stahl et al. 2019). Therefore, the last decade has seen a shift towards investigating the complex interplay between genetic and environmental risk factors (Sharma et al. 2016). Advances in technologies have supported high-throughput investigations of biological markers representative of environmental modulation of the genome. These biomarkers hold promise for stratifying symptom-based phenotypes and assessing the prognosis of individual patients (Kobeissy et al. 2012). Moreover, these biomarkers could contribute to a more accurate multi-level diagnostic framework which relies on biological measures to supplement clinical ratings of symptoms (Meana and Mollinedo-Gajate 2017).

BD is a chronic, disabling, and severe mental illness characterized by recurrent depressive and manic episodes, somatic and psychiatric comorbidities, and functional impairments (Goodwin and Jamison 2007). Considering the high global burden and lifetime prevalence of bipolar spectrum disorders, estimated at approximately 2.4% (Rowland and Marwaha 2018), there is a need to better understand the factors affecting its onset and course. The significance of environment, especially childhood trauma and stressful life events on the trajectories of affective disorders, including vulnerability, onset, relapse and occurrence, has been well established (Aldinger and Schulze 2017; Lex et al. 2017; Johnson 2005; Alloy et al. 2005; Paykel 2003). However, little is known about the mechanisms involved in the consequences of such life events.

Recently, emphasis has been placed on the potential role of epigenetic variation in the etiopathogenesis of BD (Li et al. 2015). Epigenetics is an adaptive mechanism which can modulate the stress response through subtle gene expression modifications (Aas et al. 2016). In particular, DNA methylation (DNAm), the addition of a methyl group to DNA, primarily at cytosine-guanine dinucleotides (CpG), may pose a “mechanism by which life-experiences become ‘embedded’ in the genome” (Marzi et al. 2018).

Increasing evidence from both animal and human data supports the epigenetic programming of genes in response to trauma and chronic stress. Consistent findings have linked prenatal (Monk et al. 2012; Weaver et al. 2004) and early-life adversities to epigenetic modifications of genes, especially those involved in the hypothalamic–pituitary–adrenal (HPA) axis (Kular and Kular 2018; McGowan et al. 2009; Vinkers et al. 2015; Jaworska-Andrzejewska and Rybakowski 2019). While several studies have shown methylation changes associated with trauma during the adult period, few studies have

investigated non-traumatic chronic stress (Matosin et al. 2017) or acute stressful life events. Candidate gene approaches in the general population have reported differential methylation of CpGs in the vicinity of *SLC6A4* (Alasaari et al. 2012), *TH* (Myaki et al. 2015), and *BDNF* (Song et al. 2014) in association with sustained work-related stress. One study, which examined *LINE-1* as a proxy for global methylation, found no significant associations with chronic lifestyle stress (Duman and Canli 2015). To the best of our knowledge, not a single study has explored epigenome-wide signatures of DNAm in relation to acute, non-traumatic stress in humans. With regards to BD, studies have investigated methylation differences as both trait and state markers of the disorder in several promoter regions including *SLC6A4*, *PPIEL31*, *BDNF*, *HCG9*, *KCNQ3*, *5HTR1A* and *GPR24* (Ludwig and Dwivedi 2016; Fries et al. 2016; Pishva et al. 2014). Interestingly, evidence supports altered DNAm profiles for high-risk affected and even unaffected offspring of individuals with BD in comparison to low risk controls. Moreover, there seems to be a unique rate of change in DNAm over time for high risk individuals (Duffy et al. 2019). However, despite findings of differential epigenetic profiles, results have been inconsistent and there remains a need for genome-wide methylation studies, especially ones longitudinal in design.

This study aims to gain a better understanding of the role of epigenetic modifications, specifically DNAm, in relation to stress during the course of BD. Using repeated measures over a 1-year period, we explored the relationship between DNAm and stressful life events in chronic BD patients. We took a two-pronged approach, first by interrogating CpGs in the vicinity of candidate genes previously implicated in the stress response and, second, by conducting an exploratory epigenome-wide analysis. Furthermore, we determined whether changes in symptom and stress measures over time were associated with a DNAm-based age estimate and epigenetic aging.

Methods

Study sample

The study was conducted using data from the longitudinal PsyCourse cohort. PsyCourse has been described in detail (Budde et al. 2019). Briefly, PsyCourse is a multi-site, naturalistic study, based in the German and Austrian population. Psychopathology, pharmacological treatment, childhood trauma and current stressful life events were among other variables assessed at each of four visits (6-month intervals). Likewise, peripheral blood samples were collected at each visit, paving the way for a detailed analysis of the longitudinal correlation between disease status and peripheral biomarkers. For the purpose of this study, a subset of PsyCourse participants ($n=96$) was

selected according to a DSM-IV diagnosis (American Psychiatric Association 2002) of type I or II BD, availability of genotype data and biomaterial, and completed childhood trauma and stressful life events questionnaires. Demographic and clinical characteristics of these patients are reported in Table 1. The study was approved by the local ethics committee for each study center and was carried out following the rules of the Declaration of Helsinki. All individuals provided written informed consent.

Measures

Stressful life events

Current stressful life events were assessed with the Life Events Questionnaire (LEQ), a 79-item self-report instrument that has been described in detail (Norbeck 1984; Sarason et al. 1978). The LEQ covers a wide range of stressor exposure related to health, work, school, residence, love and marriage, family and friends, parenting, the personal sphere or social environment, finances, crime and legal matters. At each visit, participants reported whether they experienced any of the listed events in the last 6 months. When the patient experienced a specific event, they rated: (1) the nature of the

event (good/bad) and (2) the impact of the event on his/her life (0–3). At each time point, adverse life events were summed to yield a stress score that reflects the impact ratings of all “bad” events. The same was done for the impact ratings of “good” events. A total score was also summed including impact ratings of both “bad” and “good” events. These three LEQ scores were used as outcome measures in our association analyses.

Childhood trauma

The Childhood Trauma Screener (CTS) is a German, short version of the Childhood Trauma Questionnaire (Bernstein et al. 1997, 2003; Grabe et al. 2012). The screener includes five questions to assess sexual, physical and emotional abuse, as well as emotional and physical neglect. Validated threshold values (Glaesmer et al. 2013) were used to transform ratings for each item into a dichotomous scale in order to identify individuals with reported childhood trauma (yes/no). Details on reported childhood trauma and thresholds used can be found in Additional file 1: Table S1.

Symptom ratings

The Positive and Negative Syndrome Scale (PANSS) was used as a measure of psychopathology at the time of testing (Kay et al. 1987). A continuous total score of the three subscales, i.e. positive, negative, and general symptoms was used. The Global Assessment of Functioning (GAF) score was used as a measure of psychosocial functioning (Luborsky 1962; Endicott et al. 1976). The Young Mania Rating Scale (YMRS) was used as a measure of manic symptoms in the last 48 h (Young et al. 1978). Lastly, the Inventory of Depressive Symptomatology (IDS-C₃₀), a 30-item rating scale, was used to assess the severity of depressive symptoms (Trivedi et al. 2004).

Analysis of DNA methylation

DNA samples

Genomic DNA was extracted from whole blood using the PerkinElmer Chemagen Kit (chemagic DNA Blood10k prefilling VD120419.che) and all samples were subsequently stored in a Hamilton Bios M system at -80°C . DNA quality was assessed using the QIAxcl[®] system. DNA samples from baseline and 1-year follow-up visits were used to obtain methylation data. Prior to downstream analyses, potential population stratification was evaluated, and an initial step to remove European population outliers was taken (Budde et al. 2019). Thus, our sample consists of an ethnically homogenous population of Caucasians of European descent.

Table 1 Sample demographic and clinical characteristics

	Baseline (n = 96)	1-year follow-up (n = 95)	p-value
Sex			
Female	50	50	
Age, mean \pm SD	45.2 \pm 12.4	46.17 \pm 12.4	
Duration of illness, mean \pm SD	13.52 \pm 11.8	14.66 \pm 11.8	
DSM-IV diagnosis			
BD-I	79	78	
BD-II	17	17	
Medication			
Combo therapy	81	75	
Monotherapy	14	16	
No meds	1	4	
Childhood trauma (yes)	48	48	
LEQ scores, mean \pm SD			
Bad events	10.2 \pm 13.8	6.3 \pm 6.6	0.004 ^b
Good events	9.7 \pm 10.2	8.4 \pm 7.6	0.191 ^b
Total events	19.9 \pm 18.4	14.1 \pm 10.7	0.001 ^b
Symptom ratings			
GAF, mean \pm SD	61.5 \pm 12.6	65.8 \pm 12.4	0.032 ^a
YMRS sum, mean \pm SD	3.9 \pm 5.8	2.4 \pm 3.7	0.216 ^b
IDS-C ₃₀ , mean \pm SD	13.7 \pm 11.0	10.6 \pm 9.7	0.124 ^b
PANSS sum, mean \pm SD	42.8 \pm 11.8	39.2 \pm 9.6	0.063 ^b

^a Paired sample t-test

^b Wilcoxon signed rank test

Illumina EPIC chip processing

Bisulfide conversion of DNA and processing of methylation arrays was accomplished in collaboration with the Institute of Human Genetics, University of Bonn, Germany. Whole-blood genomic DNA diluted with water (50 ng/μl) was treated with sodium bisulfite using the EpiTect® Bisulfite Kit from QIAGEN® following the manufacturer's protocol. DNAm was assessed using the Illumina Infinium Human MethylationEPIC BeadChip array (Illumina Inc., San Diego, CA, USA) according to the manufacturer's instructions. To minimize batch effects during DNAm measurement, an algorithm for sample randomization was used for positioning samples onto 96-well plates according to exposures of interest and confounding variables (see Additional file 1).

Quality control and normalization

Quality control

The Bioconductor R package *minfi* was used to read raw intensity data files (.idat files) into R and for the subsequent quality control and normalization of methylation data (Aryee et al. 2014). Concordance between methylation-predicted and reported sex was confirmed. Filtering of poor-performing samples and probes was performed (see Additional file 1: Table S2). Probes with low detection *p*-values (>0.05 in $>10\%$ of samples) were excluded. Using the function *dropLociWithSnps()*, SNPs inside the probe body and at the nucleotide extension were removed according to a minor allele frequency $\geq 5\%$ based on dbSNP. To prevent a possible gender effect, X and Y chromosomes were removed. According to a list previously published (Chen et al. 2013), non-specific probes i.e. probes on the EPIC array that co-hybridize to alternate genomic sequences, were removed. Lastly, probes with a bead count <3 were removed.

Normalization

Data were normalized using functional normalization (FunNorm), an extension of quantile normalization. FunNorm uses internal control probes present on the array to infer between-array technical variation, by default using the first two principal components of the control probes (Fortin et al. 2014). Density plots were used to evaluate the distribution of *M*-values before and after functional normalization (see Additional file 1: Fig. S1).

Technical batch effects were then identified using linear regressions to inspect the association of principal components of the methylation values with possible technical batches. Additionally, the R package *shinyMethyl* was used for visual inspection of principle component analysis (PCA) plots. Identified batch effects (i.e., array and slide) were removed using the Empirical Bayes' method *ComBat* (Johnson et al. 2007). Batch corrected

M-values after *ComBat* were used for downstream analyses (see Additional file 1: Fig. S2). According to inspection of PCA plots, a single sample remained an outlier after batch correction and was excluded.

Confounders

Considering cell-type composition is a confounding factor in epigenome-wide association studies (EWAS), the *minfi* function *estimateCellcounts()* was used to estimate the cell type composition for our samples. This function uses a modified version of the Houseman algorithm to obtain a cell counts vector for the six cell-types (i.e., CD4T, CD8T, NK, B cells, monocytes, and granulocytes) for each sample (Houseman et al. 2012).

Active smoking is another established modifier of DNA methylation (Lee and Pausova 2013). Methylation-based smoking scores were calculated based on the methylation profile of the 187 CpG sites identified in Zeilinger et al. (2013). First, raw beta values were normalized using the Teschendorff et al. beta-mixture quantile dilation (BMIQ) strategy (Teschendorff et al. 2013). Adjusted beta-values were then used for calculation of methylation-based smoking scores using methods previously described (Elliott et al. 2014). The correlation between self-reported number of cigarettes smoked yearly and methylation-based smoking scores was assessed (Spearman's $\rho = 0.64$; $p < 0.001$).

To rule out possible confounding effects of medication, 5 samples were excluded in sensitivity analyses. These samples were participants who were not taking psychotropic drugs at the time of testing. All other participants were taking at least one (monotherapy) or a combination (combo therapy) of the following (1) antidepressants, (2) antipsychotics, (3) mood stabilizers, (4) tranquilizers, or (5) other psychiatric medications.

Statistical analyses

All statistical analyses were performed in R version 3.4.4 (<http://www.r-project.org/>) (R Core Team 2014).

Change in methylation over time

The general "stability" of methylation over time was investigated. First, the absolute change in methylation β -values between baseline and 1-year follow-up visits were calculated across all CpG sites. To determine whether differential methylation between visits remained significant after adjusting for known confounders, the package *lme4* (Bates et al. 2015) was used to fit a linear mixed-effects model (LMM) with the dependent variable "*M*-value" and the independent variable "time", adjusting for age, sex, DNAm smoking scores, and cell composition estimates. Patient ID was included as the random effect term.

Candidate gene analysis

The association between LEQ scores and the interaction between CT and total LEQ scores with DNAm was assessed via LMMs, adjusting for covariates as described above. We interrogated DNAm in the vicinity of genes previously implicated in the HPA-axis (i.e. *BDNF*, *FKBP5*, *IL6*, *SLC6A4*, and *OXTR*). All probes on the EPIC array annotated to each of these five genes were identified. The number of probes per gene ranged from 22 to 124. We corrected for multiple testing on a gene-level by applying the false discovery (FDR) correction (Benjamini and Hochberg 1995) per gene, with FDR-corrected p -values ≤ 0.05 deemed significant. Afterwards, Bonferroni-correction was used to correct overall for the number of candidate-genes tested.

Exploratory EWAS

An exploratory EWAS was conducted. As a means of noise reduction, the top 10% of the most variable CpGs of the normalized, batch corrected M -values were extracted according to median absolute deviation (MAD) scores i.e. the median of the absolute deviations from the data's median. Associations between the most variable sites and LEQ scores and the interaction between childhood trauma and total LEQ scores were then tested using LMMs, adjusting for covariates as described above.

Epigenetic aging

DNAm-based age prediction was performed using the Horvath age estimation algorithm (Horvath 2013) with a freely available online tool (<https://dnamage.genetics.ucla.edu/home>) which predicts DNAm-age based on the methylation of 353 CpGs using an elastic net penalized regression model. The difference between the estimated epigenetic age and chronological age (Δ age) and a measure of epigenetic age acceleration (AA), i.e., the residual from regressing DNAm age on chronological age, were calculated. LMMs were used to determine the effect of LEQ scores on Δ age, adjusting for chronological age, sex, DNAm smoking scores, cell composition estimates, and technical batch effects (sample slide and array). Additionally, the difference in symptom ratings and stress scores between visits were calculated. The association between the change in symptoms and LEQ scores between baseline and 1-year follow-up with AA at 1-year follow-up was determined via linear regression models, again controlling for chronological age, sex, DNAm smoking scores, cell composition estimates and technical batch effects.

Additional analyses

Nominally significant CpGs (unadjusted $p < 0.05$) associated with total LEQ scores were used for gene-based enrichment analysis using the *GOMeth* function from the Bioconductor package *missMethyl*. *GOMeth* maps a vector of CpG sites to Entrez Gene IDs, and tests for gene ontology (GO) term pathway enrichment using a hypergeometric test (Geeleher et al. 2013). Additionally, the correlation between DNAm in blood and four brain regions was explored for the most suggestive CpGs associated with total LEQ scores (see Additional file 1).

Results

Change in methylation over time

The mean absolute difference in methylation (β) between visits 1 and 3 ($|\Delta\beta|$) was calculated across all samples for all CpG sites (Fig. 1). Over the 1-year period, $|\Delta\beta|$ ranged from <0.001 to 0.299 with an average change of 0.014. Of 753,251 CpG sites, only 68 had an $|\Delta\beta|$ of 0.10 or more, while 8454 sites differed by at least 0.05 between visits. Just over half of the sites (428,610) showed an absolute difference in methylation of at least 1%. Investigation of the functional genomic distribution of the least stable CpGs over time ($|\Delta\beta| \geq 0.10$) revealed the majority of CpGs fell within Open Seas, while 12 fell within CpG Islands, and the remaining in CpG Shores and Shelves (Fig. 2). In summary, 34,776 CpG sites showed a nominally significant difference over time (unadjusted p -value < 0.05), after correcting for age, sex, smoking and cell composition estimates. However, not a single locus withstood correction for multiple testing (FDR-corrected p -value < 0.05).

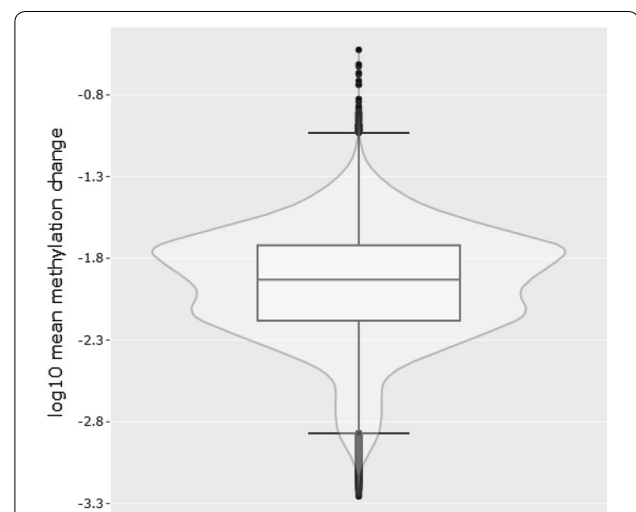
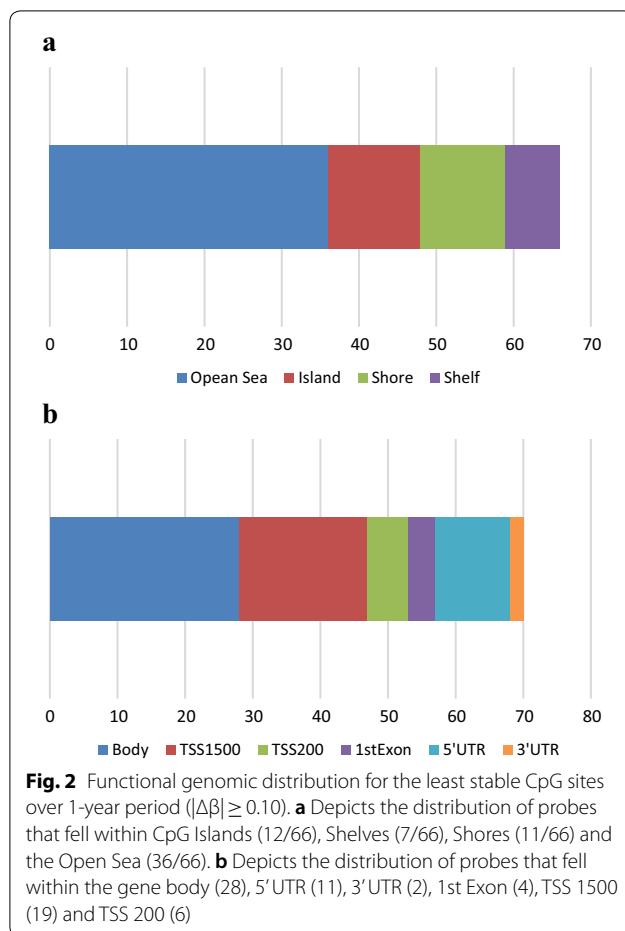
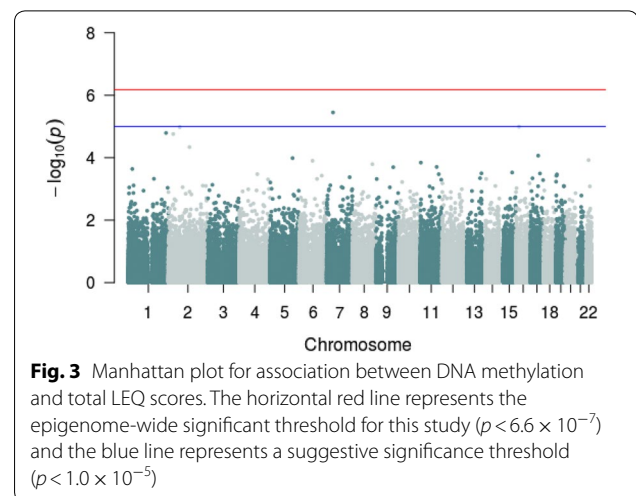


Fig. 1 Boxplot depicting the log10 mean change in methylation (β) between baseline and 1-year follow-up visit



Methylation association analysis

We performed an exploratory analysis looking for associations between LEQ scores and DNAm in individual CpG probes in the vicinity of candidate genes previously implicated in the stress response and in the most variable CpG sites across the epigenome. Methylation at a single CpG site (cg15212455; *POU6F2*; “POU class 6 homeobox 2”; chr 7) was associated with impact ratings of total LEQ scores with a suggestive significance of $p < 1.0 \times 10^{-5}$, although not a single locus withstood correction for multiple testing (FDR-corrected $p > 0.05$ for all comparisons). Figure 3 shows the Manhattan plot depicting all analyzed CpG sites with their calculated p -values for the association between DNAm and total LEQ scores. Table 2 lists the top 20 loci associated at nominal significance with total LEQ scores. Inspection of quantile–quantile (QQ) plots did not show evidence for inflation or bias (Fig. 4; Lambda factor = 0.98). Manhattan plots and associated QQ plots for additional association analyses can be found in Additional file 1: Fig. S3–S8. The sensitivity analysis, excluding subjects who did not take psychotropic drugs at the time of testing, did not yield significant



associations. These results, specific to modeling the association between DNAm and total LEQ scores, are presented in Additional file 1: Figs. S9 and S10.

Epigenetic aging

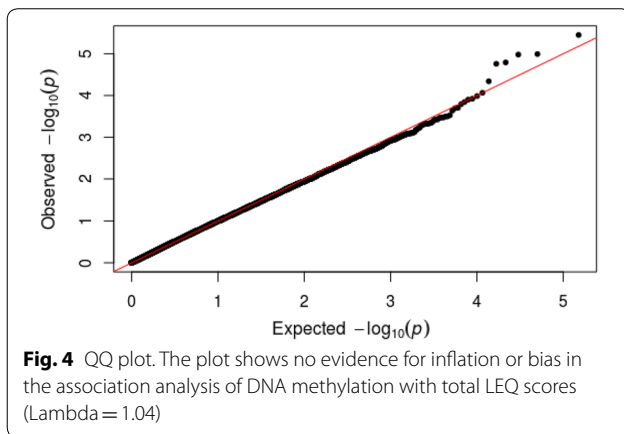
As expected, there was a strong positive correlation between individuals' DNAm age and chronological age ($r = 0.941$, $p < 0.001$; see Additional file 1: Fig. S11). According to Horvath's estimate, the mean (SD, range) AA was -0.23 years (3.71, range -9.94 to 9.86 years) at baseline and 0.25 years (3.95, range -8.12 to 9.43 years) at the 1-year follow-up. Between visits, the mean (SD, range) change in AA was 0.50 years (4.97, range -10.72 to 13.85 years). Overall, no statistically significant associations between epigenetic aging and symptom or stress measures were detected.

Additional analyses

We included genes mapped by the top CpG sites (unadjusted $p < 0.05$) associated with total LEQ scores in an enrichment analysis. No biological processes survived FDR correction (see Additional file 1: Table S3). Blood brain correlation coefficients for methylation of the top 20 loci associated with total LEQ scores (overlapping with the 450 K Beadchip array) are presented in Table 3. Eight of the top 20 most differentially methylated loci associated with total LEQ scores showed a significant correlation between methylation in the blood and methylation in at least one brain region. Methylation of the CpG site that was most strongly associated with total LEQ scores was significantly correlated with methylation in all four brain regions ($p < 0.001$; see Additional file 1: Fig. S12).

Table 2 Top 20 CpG sites associated with total LEQ scores

Probe	t value	p-value	FDR-corrected p-value	Chr	Relation to island	Annotated gene
cg15212455	− 4.87	3.56E−06	0.263	chr7	Open Sea	<i>POU6F2</i>
cg05335886	− 4.55	1.02E−05	0.263	chr16	Island	<i>TMC5</i>
cg09725915	4.54	1.05E−05	0.263	chr2	Island	
cg24511004	4.43	1.62E−05	0.263	chr1	Open Sea	
cg18110277	− 4.50	1.74E−05	0.263	chr2	Open Sea	
cg21516302	− 4.24	4.58E−05	0.575	chr2	Open Sea	
cg05180443	− 4.04	8.61E−05	0.927	chr17	Island	<i>CHAD; ACSF2</i>
cg01440452	− 3.97	1.03E−04	0.946	chr5	N Shore	<i>PURA</i>
cg26730347	− 4.00	1.20E−04	0.946	chr22	N Shore	<i>SLC5A1</i>
cg15869582	3.94	1.26E−04	0.946	chr6	S Shore	<i>IBTK</i>
cg05919744	3.95	1.44E−04	0.977	chr11	S Shore	<i>SLC22A18A5; SLC22A18</i>
cg26822318	3.86	1.61E−04	0.977	chr8	Open Sea	<i>FER1L6</i>
cg27296293	− 3.80	1.98E−04	0.977	chr11	Island	<i>RP11-748H22.1; TRPC6</i>
cg06334363	3.82	2.01E−04	0.977	chr9	S Shore	<i>RP11-235C23.5; FKTN</i>
cg00356897	− 3.79	2.30E−04	0.977	chr1	Open Sea	<i>RP4-594I10.2</i>
cg24795825	3.72	2.98E−04	0.977	chr15	N Shore	<i>MORF4L1</i>
cg17984201	3.69	3.17E−04	0.977	chr13	Open Sea	
cg18002447	− 3.67	3.20E−04	0.977	chr17	Island	
cg07349208	− 3.66	3.36E−04	0.977	chr4	Island	<i>RP11-380D23.2</i>
cg05705044	3.65	3.38E−04	0.977	chr11	S Shore	<i>RBM7</i>



Discussion

To the best of our knowledge, our study is the first to investigate epigenome-wide methylation changes over time in BD patients. Moreover, it is the first to explore methylation changes related to non-traumatic stressful life events on an epigenome-wide scale. Although no locus withstood correction for multiple testing, our suggestive findings and secondary analyses provide limited evidence supporting a role of DNAm in association with non-traumatic life events in chronic BD patients.

We identified a single, suggestively significant, CpG site associated with total LEQ scores, mapping to *POU6F2*, which has been associated with several psychiatric traits as well as intelligence and educational attainment. More specifically, genome-wide association studies have identified *POU6F2* risk variants associated with psychological distress (Koshimizu et al. 2019), feeling emotionally hurt (Nagel et al. 2018), schizophrenia (Goes et al. 2015), autism (Anney et al. 2010), educational attainment (Lee et al. 2018; Okbay et al. 2016) and intelligence (Hill et al. 2018; Davies et al. 2018). Additionally, in a longitudinal investigation of DNAm changes preceding adolescent psychotic experiences, DNAm of the CpG site cg11604728 (*POU6F2*) measured at age 15–17 was among the top 20 CpG sites indicative of psychotic experiences at age 18 (Roberts et al. 2019). Furthermore, *POU6F2* is highly expressed in the brain with the highest expression found in the frontal cortex (Additional file 1: Fig. S13) and methylation of our suggestive CpG site in blood is correlated with methylation in brain tissue across multiple brain regions. Interestingly, another of our top 20 CpG sites (cg26822318) falls in proximity to the *FER1L6* gene, of which a variant (rs4870888) has been associated with suicide attempts in a meta-analysis of major depressive disorder, schizophrenia and BD (Mullins et al. 2019). Furthermore, another GWAS reported a *FER1L6*

Table 3 Blood-brain methylation correlation for top differentially methylated CpGs associated with total LEQ scores

Probe	Blood-PFC	<i>p</i> -value	Blood-EC	<i>p</i> -value	Blood-STG	<i>p</i> -value	Blood-CER	<i>p</i> -value
cg15212455	0.721	4.16E−13	0.731	4.64E−13	0.747	1.48E−14	0.631	3.71E−09
cg05335886	− 0.086	0.467	− 0.101	0.404	− 0.145	0.213	− 0.155	0.196
cg09725915	0.576	7.86E−08	0.532	1.76E−06	0.626	1.96E−09	0.489	1.49E−05
cg21516302	0.373	0.001	0.522	3.01E−06	0.507	3.46E−06	0.307	0.009
cg05180443	0.204	0.081	0.336	0.004	0.298	0.009	0.175	0.144
cg01440452	− 0.097	0.413	0.062	0.606	− 0.119	0.309	0.020	0.870
cg26730347	0.499	6.05E−06	0.568	2.33E−07	0.562	1.51E−07	0.493	1.27E−05
cg27296293	0.131	0.265	− 0.214	0.073	0.037	0.755	0.236	0.048
cg24795825	0.016	0.894	0.299	0.011	0.247	0.033	0.121	0.317
cg18002447	0.038	0.749	0.034	0.776	0.042	0.720	− 0.262	0.028
cg07349208	0.167	0.156	0.095	0.431	− 0.112	0.338	0.068	0.571

PFC prefrontal cortex, EC entorhinal cortex, STG superior temporal gyrus, CER cerebellum

Significant correlations in italics

variant (rs10481151) suggestively associated with cognitive performance (Need et al. 2009).

At the current sample size, our study provides only minimal evidence supporting an association between methylation of individual CpGs and non-traumatic, recent stressful life events in BD. These findings, however, corroborate other reports of a limited role of DNAm with non-traumatic stress (Marzi et al. 2018). Noteworthy, a recent study reported hypermethylation of *KITLG* associated with childhood trauma in healthy controls ($n=91$) but not in bipolar patients ($n=50$) (He et al. 2018). Although the mechanistic role of DNAm in the phenotypic expression of early life adversities is well established in the literature, other mechanisms may be responsible in adulthood and in association with subsequent events. This notion aligns with theories such as Post's kindling hypothesis and the decay model which suggest a higher impact of life events on first episode than on subsequent episodes in BD (Aldinger and Schulze 2017; Kemner et al. 2015; Hillegers et al. 2004). Furthermore, it must be considered whether positive epigenetic associations with life events could be disorder-specific, genotype-dependent, associated with specific trauma exposure, age groups, sex and/or tissues measured (Marzi et al. 2018; Vinkers et al. 2015; Uddin et al. 2010; Boks et al. 2015; Smith et al. 2011; Mehta et al. 2017). While there is no gold standard for life stress measurements, differences in how to quantify stress may also have a major effect on findings (Johnson 2005; Bender and Alloy 2011; Monroe 2008; Dohrenwend 2010; Brown and Harris 2012).

The main strength of our study is its longitudinal design, allowing for repeated measures within individuals and to investigate methylation changes over time and in relation to symptomatology and stressful life events. To the best of our knowledge, this is the first study to

collect repeated epigenome-wide methylation measures in bipolar patients. Furthermore, our study paid attention to critical confounding factors which often lead to spurious findings. For example, the use of methylation-based smoking scores better controls for the extent of smoking throughout the lifetime than the use of self-reported smoking measures (Elliott et al. 2014; Shenker et al. 2013). Finally, in contrast to most other studies, we have included an exploratory epigenome-wide approach.

Despite the strengths of our study, several limitations need to be addressed. First, our study was limited by our small sample size which makes identifying subtle differences in methylation difficult. Taking power into consideration, and as an attempt to address the inherent multiple testing problem associated with EWAS, we limited our EWAS to only the most variable CpG sites according to MAD scores. While the fact that not a single site-specific association in DNAm survived correction for multiple testing could reflect the limited statistical power of our small sample, it may also be related to an overly conservative multiple testing correction considering the lack of variability in methylation at many CpGs and spatial correlation of methylation with nearby sites (Walker et al. 2016; Lunnon et al. 2015). A recent study estimated there are approximately 530,000 independent tests in a whole blood EPIC array DNAm study. Accordingly, they proposed a corrected significance threshold of 9.42×10^{-8} to be used as a standard threshold for future EWAS based on the EPIC array (Mansell et al. 2019). Furthermore, the study introduced a freely available online tool which allows users to perform power calculations to guide sample sizes, accounting for the individual properties of each DNAm site and using their empirically derived significance threshold. According to their tool, an effect size of just

1% difference between cases and controls would require a sample of 1000 participants, for only a third of methylation sites to have > 80% power. We observed an effect size below 5% in our study (based on median split) for our most significantly associated site, indicating that our study is nevertheless underpowered. Future studies should take advantage of this tool to assess, a priori, required sample sizes according to their expected effect sizes. Furthermore, complementary systems biology approaches such as weighted gene co-methylation network analysis (WGCNA) could be beneficial for studies with limited sample sizes, providing more insight into the functional role of altered DNAm (Langfelder and Horvath 2008).

Another limitation is in relation to the fact that our sample represents a cohort of chronic BD patients which likely influenced our investigation of epigenetic aging related to symptom ratings over time. The chronicity of patients may also confound our findings with regards to the heterogeneous treatments patients have received over the years. To acknowledge this critical factor, we conducted a sensitivity analysis excluding those subjects not taking psychotropic drugs at the time of testing, however, this also did not lead to significant results. One must also consider the possible recall and desirability biases associated with self-rating questionnaires like the LEQ and CTS. Lastly, little is known about the temporal stability of epigenetic markers (Byun et al. 2012; Talens et al. 2010). We cannot be sure whether the time interval of 1 year was too long or short to observe dramatic methylation changes or at what time window following exposure to stressful life events one might observe changed methylation profiles.

Conclusions

BD is a multifactorial psychiatric illness, and for many patients full interepisodic remission never occurs (Sam et al. 2019). Stressful life events have been associated with a worse course of BD (Aldinger and Schulze 2017) and there remains a need to better understand the mechanisms which allow these stressors to bring about phenotypic change. Our study provides limited evidence supporting an association between DNAm and recent, non-traumatic stressful life events in BD patients. As findings in clinical populations have been inconsistent, there is still much to be understood especially with regards to the temporal nature of environmentally induced DNA modifications. Future larger studies of well-characterized patients, longitudinal in design, are warranted.

Supplementary information

Supplementary information accompanies this paper at <https://doi.org/10.1186/s40345-019-0176-6>.

Additional file 1. Supporting methods, Figures S1–S13, and Tables S1–S3.

Abbreviations

BD: bipolar disorder; GWAS: genome-wide association studies; DNAm: DNA methylation; CpG: cytosine-guanine dinucleotides; HPA: hypothalamic–pituitary–adrenal; LEQ: Life Events Questionnaire; CTS: Childhood Trauma Screener; PANSS: Positive and Negative Syndrome Scale; GAF: Global Assessment of Functioning; YMRS: Young Mania Rating Scale; IDS-C₃₀: Inventory of Depressive Symptomatology; FunNorm: functional normalization; EWAS: epigenome-wide association analysis; BMIQ: beta-mixture quantile dilation; LMM: linear mixed-effects model; FDR: false discovery rate; MAD: median absolute deviation; AA: age acceleration; GO: gene ontology; QQ: quantile-quantile; WGCNA: weighted gene co-methylation network analysis.

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Authors' contributions

All authors made substantial contributions to the conception or design of the work, or the acquisition, analysis or interpretation of data. Authors ALC and TGS designed the study. Authors TGS and UH provided supervision. Author ALC conducted the analysis in consultation with UH, SS, DC and TA. Author ALC wrote the first draft of the manuscript. All authors contributed to the manuscript. All authors read and approved the final manuscript.

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Availability of data and materials

Participants of the PsyCourse study have consented to the sharing of their pseudonymized data with other researchers and research consortia. Thus, PsyCourse data will be made available to bona fide researchers collaborating with us given a mutually agreed written memorandum of understanding has been signed.

Ethics approval and consent to participate

The study was approved by the local ethics committee for each study center and was carried out following the rules of the Declaration of Helsinki.

Consent for publication

Not applicable.

Competing interests

Dr. Jens Reimer received honoraria from Otsuka-Lundbeck for participation in a speakers' bureau. Dr. Max Schmauß received honoraria from Otsuka-Lundbeck, Neuraxpharm, Aristo and Janssen for participation in the speakers' bureau or advisory boards. All other authors declare that they have no competing interests.

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References

- Aas M, Henry C, Andreassen OA, Bellivier F, Melle I, Etain B. The role of childhood trauma in bipolar disorders. *Int J Bipolar Disord*. 2016;4:2.
- Alsaari JS, Lagus M, Ollila HM, Toivola A, Kivimäki M, Vahtera J, et al. Environmental stress affects DNA methylation of a CpG rich promoter region of serotonin transporter gene in a nurse cohort. *PLoS ONE*. 2012;7(9):e45813.
- Aldinger F, Schulze TG. Environmental factors, life events, and trauma in the course of bipolar disorder. *Psychiatry Clin Neurosci*. 2017;71(1):6–17.
- Alloy LB, Abramson LY, Urosevic S, Walshaw PD, Nusslock R, Neeren AM. The psychosocial context of bipolar disorder: environmental, cognitive, and developmental risk factors. *Clin Psychol Rev*. 2005;25(8):1043–75.
- American Psychiatric Association. Diagnostic and statistical manual of mental disorders. 4th ed. Washington, DC: American Psychiatric Association; 2002.
- Anney R, Klei L, Pinto D, Regan R, Conroy J, Magalhaes TR, et al. A genome-wide scan for common alleles affecting risk for autism. *Hum Mol Genet*. 2010;19(20):4072–82.
- Aryee MJ, Jaffe AE, Corrada-Bravo H, Ladd-Acosta C, Feinberg AP, Hansen KD, et al. Minfi: a flexible and comprehensive bioconductor package for the analysis of Infinium DNA methylation microarrays. *Bioinformatics*. 2014;30(10):1363–9.
- Bates D, Maechler M, Bolker B, Walker S. Fitting linear mixed-effects models using lme4. *J Stat Softw*. 2015;67(1):1–48.
- Bender RE, Alloy LB. Life stress and kindling in bipolar disorder: review of the evidence and integration with emerging biopsychosocial theories. *Clin Psychol Rev*. 2011;31(3):383–98.
- Benjamini Y, Hochberg Y. Controlling the false discovery rate: a practical and powerful approach to multiple testing. *J R Stat Soc B*. 1995;57(1):289–300.
- Bernstein DP, Ahluwalia T, Pogge D, Handelsman L. Validity of the Childhood Trauma Questionnaire in an adolescent psychiatric population. *J Am Acad Child Adolesc Psychiatry*. 1997;36(3):340–8.
- Bernstein DP, Stein JA, Newcomb MD, Walker E, Pogge D, Ahluwalia T, et al. Development and validation of a brief screening version of the Childhood Trauma Questionnaire. *Child Abuse Negl*. 2003;27(2):169–90.
- Boks MP, van Mierlo HC, Rutten BP, Radstake TR, De Witte L, Geuze E, et al. Longitudinal changes of telomere length and epigenetic age related to traumatic stress and post-traumatic stress disorder. *Psychoneuroendocrinology*. 2015;51:506–12.
- Brown GW, Harris T. Social origins of depression: a study of psychiatric disorder in women. New York: Routledge; 2012.
- Budde M, Anderson-Schmidt H, Gade K, Reich-Elkelenz D, Adorjan K, Kalman JL, et al. A longitudinal approach to biological psychiatric research: the PsyCourse study. *Am J Med Genet B Neuropsychiatr Genet*. 2019;180(2):89–102.
- Byun HM, Nordio F, Coull BA, Tarantini L, Hou L, Bonzini M, et al. Temporal stability of epigenetic markers: sequence characteristics and predictors of short-term DNA methylation variations. *PLoS ONE*. 2012;7(6):e39220.
- Chen YA, Lemire M, Choufani S, Butcher DT, Grafodatskaya D, Zanke BW, et al. Discovery of cross-reactive probes and polymorphic CpGs in the Illumina Infinium HumanMethylation450 microarray. *Epigenetics*. 2013;8(2):203–9.
- Davies G, Lam M, Harris SE, Trampush JW, Luciano M, Hill WD, et al. Study of 300,486 individuals identifies 148 independent genetic loci influencing general cognitive function. *Nat Commun*. 2018;9(1):2098.
- Dohrenwend BP. Toward a typology of high-risk major stressful events and situations in posttraumatic stress disorder and related psychopathology. *Psychol Inj Law*. 2010;3(2):89–99.
- Duffy A, Goodday SM, Keown-Stoneman C, Scotti M, Maitra M, Nagy C, et al. Epigenetic markers in inflammation-related genes associated with mood disorder: a cross-sectional and longitudinal study in high-risk offspring of bipolar parents. *Int J Bipolar Disord*. 2019;7(1):17.
- Duman EA, Canli T. Influence of life stress, 5-HTTLPR genotype, and SLC6A4 methylation on gene expression and stress response in healthy Caucasian males. *Biol Mood Anxiety Disord*. 2015;5:2.
- Elliott HR, Tillin T, McArdle WL, Ho K, Duggirala A, Frayling TM, et al. Differences in smoking associated DNA methylation patterns in South Asians and Europeans. *Clin Epigenet*. 2014;6(1):4.
- Endicott J, Spitzer RL, Fleiss JL, Cohen J. The Global Assessment Scale: a procedure for measuring overall severity of psychiatric disturbance. *Arch Gen Psychiatry*. 1976;33(6):766–71.
- Fortin J-P, Labbe A, Lemire M, Zanke BW, Hudson TJ, Fertig EJ, et al. Functional normalization of 450k methylation array data improves replication in large cancer studies. *Genome Biol*. 2014;15(11):503.
- Fries GR, Li Q, McAlpin B, Rein T, Walss-Bass C, Soares JC, et al. The role of DNA methylation in the pathophysiology and treatment of bipolar disorder. *Neurosci Biobehav Rev*. 2016;68:474–88.
- Geeleher P, Hartnett L, Egan LJ, Golden A, Raja Ali RA, Seoighe C. Gene-set analysis is severely biased when applied to genome-wide methylation data. *Bioinformatics*. 2013;29(15):1851–7.
- Glaesmer H, Schulz A, Hauser W, Freyberger HJ, Brahler E, Grabe HJ. The childhood trauma screener (CTS)—development and validation of cut-off-scores for classificatory diagnostics. *Psychiatr Prax*. 2013;40(4):220–6.
- Goes FS, McGrath J, Avramopoulos D, Wolyniec P, Pirooznia M, Ruczinski I, et al. Genome-wide association study of schizophrenia in Ashkenazi Jews. *Am J Med Genet Part B Neuropsychiatr Genet*. 2015;168(8):649–59.

- Goodwin FK, Jamison KR. Manic-depressive illness: bipolar disorders and recurrent depression. 2nd ed. New York: Oxford University Press; 2007.
- Grabe HJ, Schulz A, Schmidt CO, Appel K, Driessen M, Wingenfeld K, et al. Ein Screeninginstrument für Missbrauch und Vernachlässigung in der Kindheit: der Childhood Trauma Screener (CTS). *Psychiatr Prax*. 2012;39(03):109–15.
- He Y, Vinkers CH, Houtepen LC, de Witte LD, Boks MP. Childhood adversity is associated with increased KILG methylation in healthy individuals but not in bipolar disorder patients. *Front Psychiatry*. 2018;9:743.
- Hill WD, Marioni RE, Maghazian O, Ritchie SJ, Hagenaars SP, McIntosh AM, et al. A combined analysis of genetically correlated traits identifies 187 loci and a role for neurogenesis and myelination in intelligence. *Mol Psychiatry*. 2018;24(2):169–81.
- Hillegers MH, Burger H, Wals M, Reichart CG, Verhulst FC, Nolen WA, et al. Impact of stressful life events, familial loading and their interaction on the onset of mood disorders: study in a high-risk cohort of adolescent offspring of parents with bipolar disorder. *Br J Psychiatry*. 2004;185:97–101.
- Horvath S. DNA methylation age of human tissues and cell types. *Genome Biol*. 2013;14(10):R115.
- Houseman EA, Accomando WP, Koestler DC, Christensen BC, Marsit CJ, Nelson HH, et al. DNA methylation arrays as surrogate measures of cell mixture distribution. *BMC Bioinform*. 2012;13:86.
- Jaworska-Andrzejewska P, Rybakowski JK. Childhood trauma in mood disorders: neurobiological mechanisms and implications for treatment. *Pharmacol Rep*. 2019;71(1):112–20.
- Johnson SL. Life events in bipolar disorder: towards more specific models. *Clin Psychol Rev*. 2005;25(8):1008–27.
- Johnson WE, Li C, Rabinovic A. Adjusting batch effects in microarray expression data using empirical Bayes methods. *Biostatistics*. 2007;8(1):118–27.
- Kay SR, Fiszbein A, Opler LA. The positive and negative syndrome scale (PANSS) for schizophrenia. *Schizophr Bull*. 1987;13(2):261–76.
- Kemner SM, van Haren NE, Bootsman F, Eijkemans MJ, Vonk R, van der Schot AC, et al. The influence of life events on first and recurrent admissions in bipolar disorder. *Int J Bipolar Disord*. 2015;3:6.
- Kobeissy F, Alawieh A, Mondello S, Boustany RM, Gold MS. Biomarkers in psychiatry: how close are we? *Front Psychiatry*. 2012;3:114.
- Koshimizu H, Nogawa S, Asano S, Ikeda M, Iwata N, Takahashi S, et al. Genome-wide association study identifies a novel locus associated with psychological distress in the Japanese population. *Transl Psychiatry*. 2019;9(1):52.
- Kular L, Kular S. Epigenetics applied to psychiatry: clinical opportunities and future challenges. *Psychiatry Clin Neurosci*. 2018;72(4):195–211.
- Langfelder P, Horvath S. WGCNA: an R package for weighted correlation network analysis. *BMC Bioinform*. 2008;9(1):559.
- Lee KWK, Pausova Z. Cigarette smoking and DNA methylation. *Front Genet*. 2013;4:132.
- Lee JJ, Wedow R, Okbay A, Kong E, Maghazian O, Zacher M, et al. Gene discovery and polygenic prediction from a genome-wide association study of educational attainment in 1.1 million individuals. *Nat Genet*. 2018;50(8):1112–21.
- Lex C, Bazner E, Meyer TD. Does stress play a significant role in bipolar disorder? A meta-analysis. *J Affect Disord*. 2017;208:298–308.
- Li Y, Camarillo C, Xu J, Arana TB, Xiao Y, Zhao Z, et al. Genome-wide methylome analyses reveal novel epigenetic regulation patterns in schizophrenia and bipolar disorder. *Biomed Res Int*. 2015;2015:201587.
- Luborsky L. Clinicians' judgments of mental health: a proposed scale. *Arch Gen Psychiatry*. 1962;7(6):407–17.
- Ludwig B, Dwivedi Y. Dissecting bipolar disorder complexity through epigenomic approach. *Mol Psychiatry*. 2016;21(11):1490–8.
- Lunnun K, Smith RG, Cooper I, Greenbaum L, Mill J, Beerli MS. Blood methylomic signatures of presymptomatic dementia in elderly subjects with type 2 diabetes mellitus. *Neurobiol Aging*. 2015;36(3):1600.e1–e4.
- Mansell G, Gorrie-Stone TJ, Bao Y, Kumari M, Schalkwyk LS, Mill J, et al. Guidance for DNA methylation studies: statistical insights from the Illumina EPIC array. *BMC Genomics*. 2019;20(1):366.
- Marzi SJ, Sugden K, Arseneault L, Belsky DW, Burrage J, Corcoran DL, et al. Analysis of DNA methylation in young people: limited evidence for an association between victimization stress and epigenetic variation in blood. *Am J Psychiatry*. 2018;175(6):517–29.
- Matosin N, Cruceanu C, Binder EB. Preclinical and clinical evidence of DNA methylation changes in response to trauma and chronic stress. *Chronic Stress*. 2017. <https://doi.org/10.1177/2470547017710764>.
- McGowan PO, Sasaki A, D'Alessio AC, Dymov S, Labonte B, Szyf M, et al. Epigenetic regulation of the glucocorticoid receptor in human brain associates with childhood abuse. *Nat Neurosci*. 2009;12(3):342–8.
- Meana JJ, Mollinedo-Gajate I. Biomarkers in psychiatry: between myth and clinical reality. *Revista de Psiquiatría y Salud Mental*. 2017;10(4):183–4 (English Edition).
- Mehta D, Bruenig D, Carrillo-Roa T, Lawford B, Harvey W, Morris CP, et al. Genomewide DNA methylation analysis in combat veterans reveals a novel locus for PTSD. *Acta Psychiatr Scand*. 2017;136(5):493–505.
- Monk C, Spicer J, Champagne FA. Linking prenatal maternal adversity to developmental outcomes in infants: the role of epigenetic pathways. *Dev Psychopathol*. 2012;24(4):1361–76.
- Monroe SM. Modern approaches to conceptualizing and measuring human life stress. *Annu Rev Clin Psychol*. 2008;4:33–52.
- Mullins N, Bigdeli TB, Borglum AD, Coleman JRI, Demontis D, Mehta D, et al. GWAS of suicide attempt in psychiatric disorders and association with major depression polygenic risk scores. *Am J Psychiatry*. 2019;176(8):651–60.
- Myaki K, Suzuki T, Song Y, Tsutsumi A, Kawakami N, Takahashi M, et al. Epigenetic changes caused by occupational stress in humans revealed through noninvasive assessment of DNA methylation of the tyrosine hydroxylase gene. *J Neurol Neuro Disord*. 2015;2(2):201.
- Nagel M, Watanabe K, Stringer S, Posthuma D, van der Sluis S. Item-level analyses reveal genetic heterogeneity in neuroticism. *Nat Commun*. 2018;9(1):905.
- Need AC, Attix DK, McEvoy JM, Cirulli ET, Linney KL, Hunt P, et al. A genome-wide study of common SNPs and CNVs in cognitive performance in the CANTAB. *Hum Mol Genet*. 2009;18(23):4650–61.
- Norbeck JS. Modification of life event questionnaires for use with female respondents. *Res Nurs Health*. 1984;7(1):61–71.
- Okbay A, Beauchamp JP, Fontana MA, Lee JJ, Pers TH, Rietveld CA, et al. Genome-wide association study identifies 74 loci associated with educational attainment. *Nature*. 2016;533(7604):539–42.
- Paykel ES. Life events and affective disorders. *Acta Psychiatr Scand*. 2003;108(s418):61–6.
- Pishva E, Kenis G, van den Hove D, Lesch KP, Boks MP, van Os J, et al. The epigenome and postnatal environmental influences in psychotic disorders. *Soc Psychiatry Psychiatr Epidemiol*. 2014;49(3):337–48.
- R Core Team. R: a language and environment for statistical computing. Vienna: R Foundation for Statistical Computing; 2014. <https://www.R-project.org/>.
- Roberts S, Suderman M, Zammitt S, Watkins SH, Hannon E, Mill J, et al. Longitudinal investigation of DNA methylation changes preceding adolescent psychotic experiences. *Transl Psychiatry*. 2019;9(1):69.
- Rowland TA, Marwaha S. Epidemiology and risk factors for bipolar disorder. *Ther Adv Psychopharmacol*. 2018;8(9):251–69.
- Sam SP, Nisha A, Varghese PJ. Stressful life events and relapse in bipolar affective disorder: a cross-sectional study from a tertiary care center of southern India. *Indian J Psychol Med*. 2019;41(1):61–7.
- Sarason IG, Johnson JH, Siegel JM. Assessing the impact of life changes: development of the Life Experiences Survey. *J Consult Clin Psychol*. 1978;46(5):932–46.
- Sharma S, Powers A, Bradley B, Ressler KJ. Gene x environment determinants of stress- and anxiety-related disorders. *Annu Rev Psychol*. 2016;67:239–61.
- Shenker NS, Polidoro S, van Veldhoven K, Sacerdote C, Ricceri F, Birrell MA, et al. Epigenome-wide association study in the European Prospective Investigation into Cancer and Nutrition (EPIC-Turin) identifies novel genetic loci associated with smoking. *Hum Mol Genet*. 2013;22(5):843–51.
- Smith AK, Conneely KN, Kilari V, Mercer KB, Weiss TE, Bradley B, et al. Differential immune system DNA methylation and cytokine regulation in post-traumatic stress disorder. *Am J Med Genet B Neuropsychiatr Genet*. 2011;156b(6):700–8.
- Song Y, Miyaki K, Suzuki T, Sasaki Y, Tsutsumi A, Kawakami N, et al. Altered DNA methylation status of human brain derived neurotrophin factor gene could be useful as biomarker of depression. *Am J Med Genet B Neuropsychiatr Genet*. 2014;165b(4):357–64.
- Stahl EA, Breen G, Forstner AJ, McQuillin A, Ripke S, Trubetskoy V, et al. Genome-wide association study identifies 30 loci associated with bipolar disorder. *Nat Genet*. 2019;51(5):793–803.

- Talens RP, Boomsma DI, Tobi EW, Kremer D, Jukema JW, Willemsen G, et al. Variation, patterns, and temporal stability of DNA methylation: considerations for epigenetic epidemiology. *FASEB J*. 2010;24(9):3135–44.
- Teschendorff AE, Marabita F, Lechner M, Bartlett T, Tegner J, Gomez-Cabrero D, et al. A beta-mixture quantile normalization method for correcting probe design bias in Illumina Infinium 450k DNA methylation data. *Bioinformatics*. 2013;29(2):189–96.
- Trivedi MH, Rush AJ, Ibrahim HM, Carmody TJ, Biggs MM, Suppes T, et al. The Inventory of Depressive Symptomatology, Clinician Rating (IDS-C) and Self-Report (IDS-SR), and the Quick Inventory of Depressive Symptomatology, Clinician Rating (QIDS-C) and Self-Report (QIDS-SR) in public sector patients with mood disorders: a psychometric evaluation. *Psychol Med*. 2004;34(1):73–82.
- Uddin M, Aiello AE, Wildman DE, Koenen KC, Pawelec G, de Los Santos R, et al. Epigenetic and immune function profiles associated with posttraumatic stress disorder. *Proc Natl Acad Sci USA*. 2010;107(20):9470–5.
- Vinkers CH, Kalafateli AL, Rutten BP, Kas MJ, Kaminsky Z, Turner JD, et al. Traumatic stress and human DNA methylation: a critical review. *Epigenomics*. 2015;7(4):593–608.
- Walker RM, Sussmann JE, Whalley HC, Ryan NM, Porteous DJ, McIntosh AM, et al. Preliminary assessment of pre-morbid DNA methylation in individuals at high genetic risk of mood disorders. *Bipolar Disord*. 2016;18(5):410–22.
- Weaver IC, Cervoni N, Champagne FA, D'Alessio AC, Sharma S, Seckl JR, et al. Epigenetic programming by maternal behavior. *Nat Neurosci*. 2004;7(8):847–54.
- Young RC, Biggs JT, Ziegler VE, Meyer DA. A rating scale for mania: reliability, validity and sensitivity. *Br J Psychiatry*. 1978;133:429–35.
- Zeilinger S, Kühnel B, Klopp N, Baurecht H, Kleinschmidt A, Gieger C, et al. Tobacco smoking leads to extensive genome-wide changes in DNA methylation. *PLoS ONE*. 2013;8(5):e63812.

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3.2 Original Article 2: Supplementary material

Supplementary methods

Batch effects. We used R version 3.4.4 to randomly draw samples from selected individuals stratified by groups (age, sex, and exposures of interest) and to assign them to batches (e.g. different microarray chips). Standard statistical tests were then used to assess if the putative technical batches (e.g. plates, microarray chips, chip rows/columns) were significantly associated with age, gender or exposure of interest. This procedure was repeated for up to 10000 permutations and the combination of samples that produced the least significant associations was selected. As within-individual variability over time was explored, baseline and visit 3 samples for each individual were loaded onto the same chip. Post hoc tests were then run on the selected combination of samples to confirm no significant associations with other variables, namely diagnosis (Bipolar I/II) or disease severity.

Blood-brain methylation correlation. The freely available Blood Brain DNA Methylation Comparison Tool (<http://epigenetics.iop.kcl.ac.uk/bloodbrain/>) (Hannon, Lunnon, Schalkwyk, & Mill, 2015) was used to determine the likely correlation between DNAm in blood with four different brain regions (prefrontal cortex, entorhinal cortex, superior temporal gyrus and cerebellum) for the most suggestive CpGs associated with total LEQ scores.

Table S1. Reported childhood trauma type

Childhood trauma type		n (%)
Abuse only		22 (45.8)
Neglect only		6 (12.5)
Number of traumas		
1		22 (45.8)
2		11 (22.9)
3		8 (16.7)
4		1 (2.1)
5		1 (2.1)
Missing complete CTS		5 (10.4)
CTS single items	Trauma threshold	
"... I had the feeling to be loved"	"Seldom" or "Not at all"	19 (39.6)
"... persons in my family hit me so hard that I bruised"	"Sometimes" to "Very often"	16 (33.3)
"... I had the feeling someone in my family hated me"	"Sometimes" to "Very often"	24 (59.0)
"... someone harassed me sexually"	"Seldom" to "Very often"	15 (31.2)
"... there was someone who took me to the doctor when I needed it"	"Seldom" or "Not at all"	10 (20.8)

Table S2. Probe filtering

CpGs probes EPIC	866,238
Low detection p-values	6529
Bead count < 3	17315
X and Y probes	19096
CpGs with SNPs	29310
Cross-reactive probes	37968
Non-specific probes	2769
CpGs after filtering	753,251

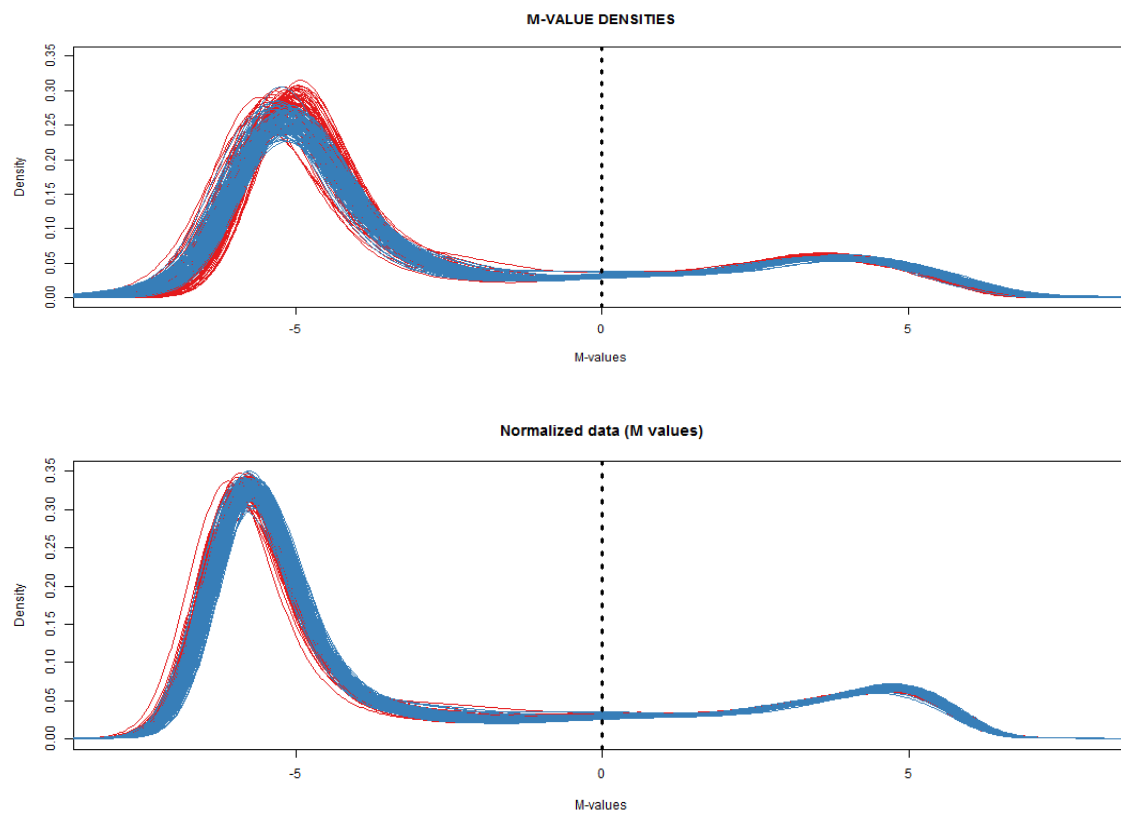


Figure S1. M-Value densities before and after functional normalization.

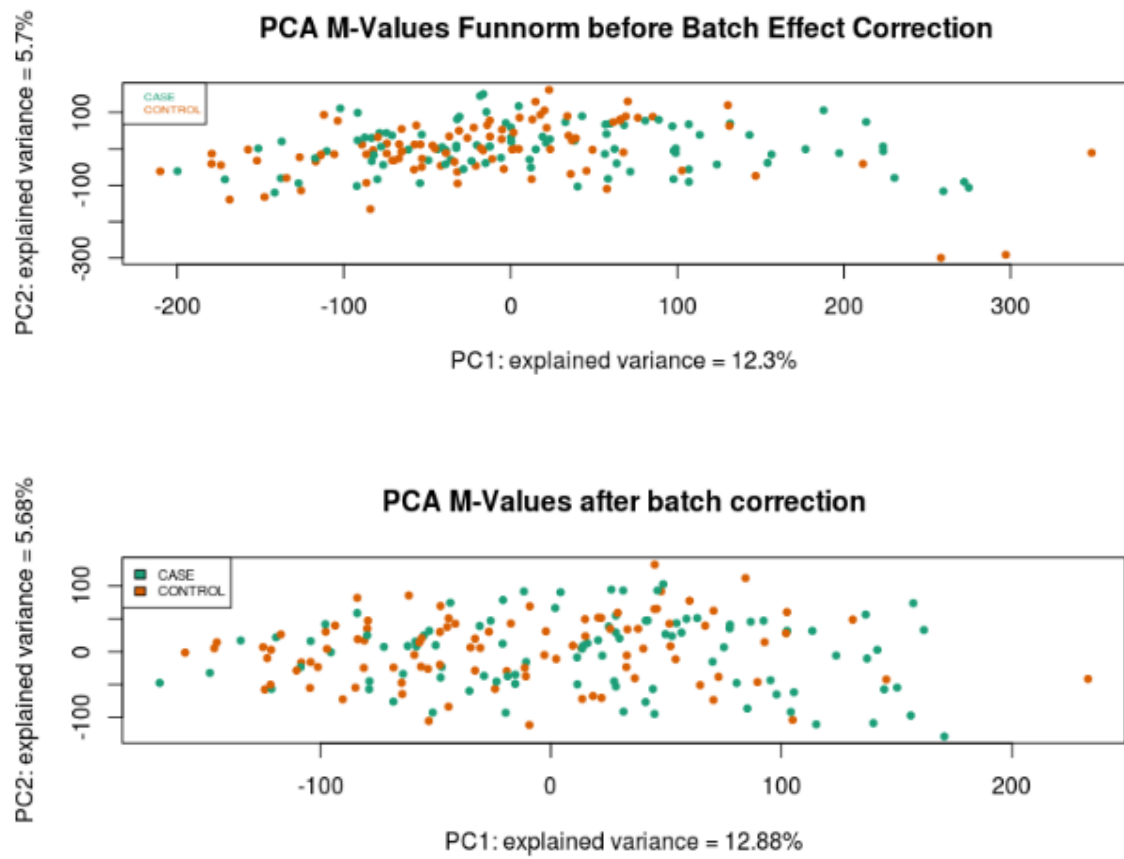


Figure S2. Inspection of batch effects through principle component analysis plots before and after batch correction with *ComBat*

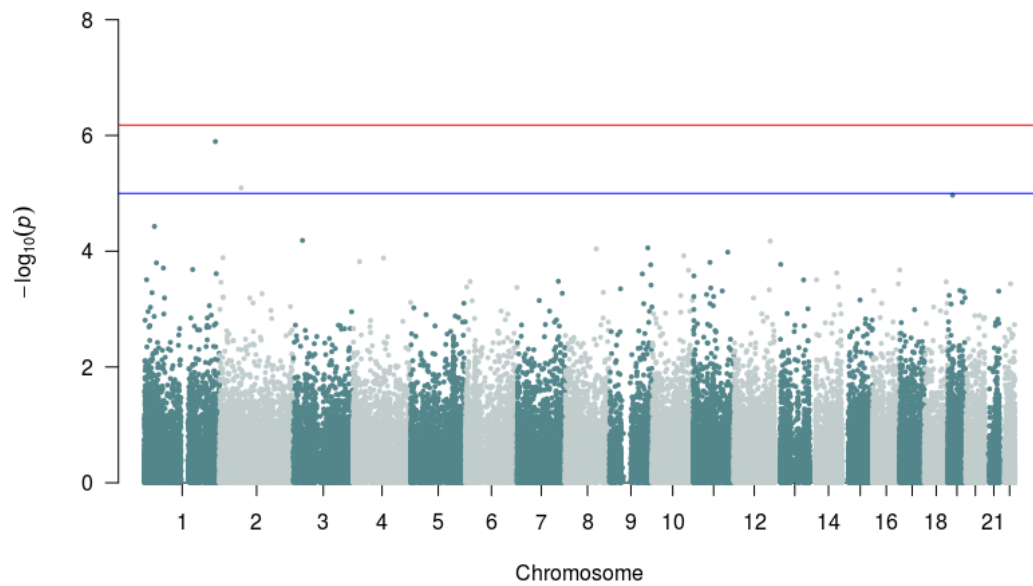


Figure S3. Manhattan plot. The Manhattan plot depicts the association between DNA methylation and LEQ “good” scores ($n = 191$). The horizontal red line represents the epigenome-wide significant threshold for this study ($p < 6.6 \times 10^{-7}$) and the blue line represents a suggestive significance threshold ($p < 1.0 \times 10^{-5}$).

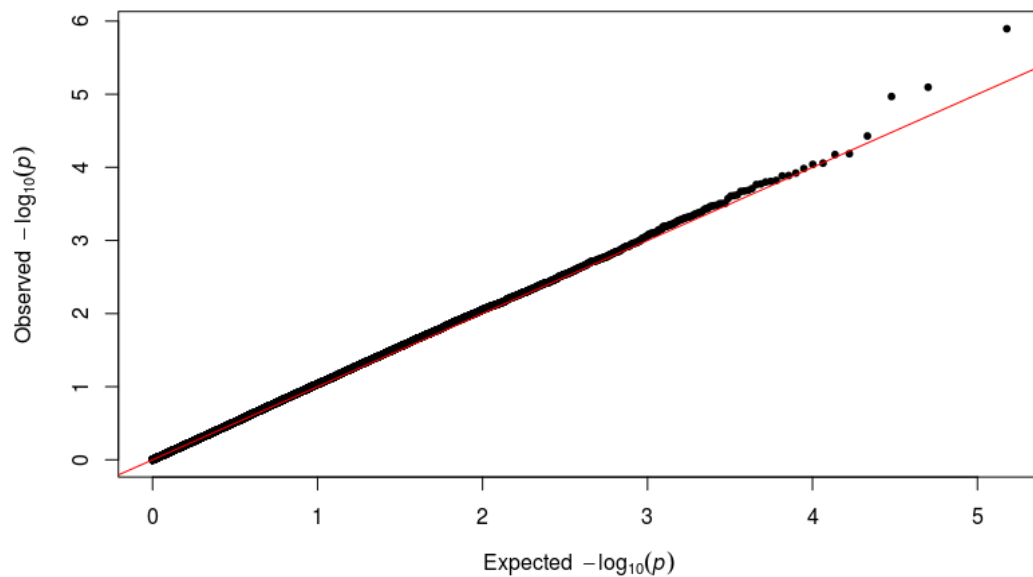


Figure S4. QQ plot. The QQ plot shows no evidence for inflation or bias in the association analysis of DNA methylation with “good” LEQ scores ($\Lambda = 1.04$).

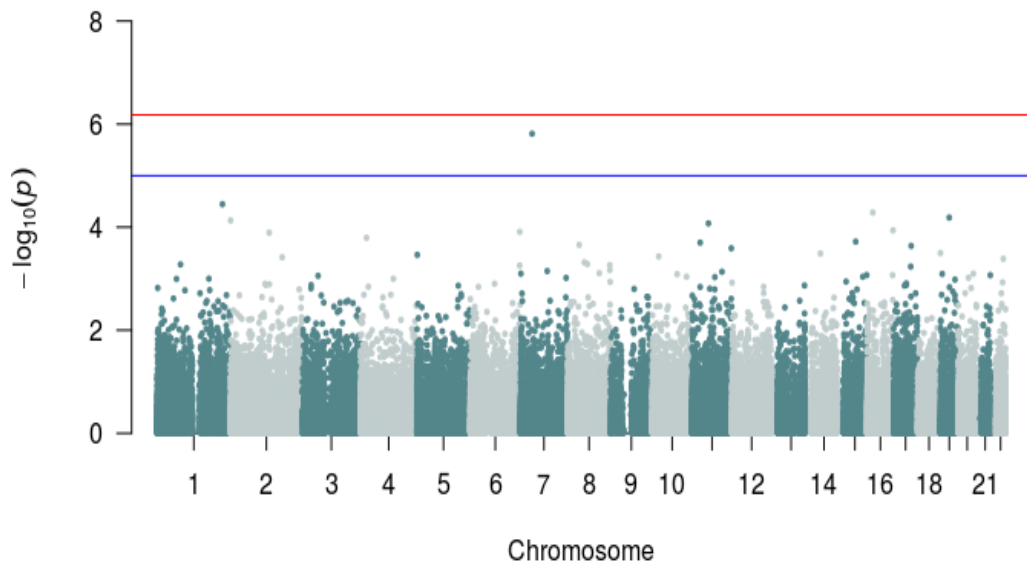


Figure S5. Manhattan plot. The Manhattan plot depicts the association between DNA methylation and “bad” LEQ scores ($n = 191$). The horizontal red line represents the epigenome-wide significant threshold for this study ($p < 6.6 \times 10^{-7}$) and the blue line represents a suggestive significance threshold ($p < 1.0 \times 10^{-5}$).

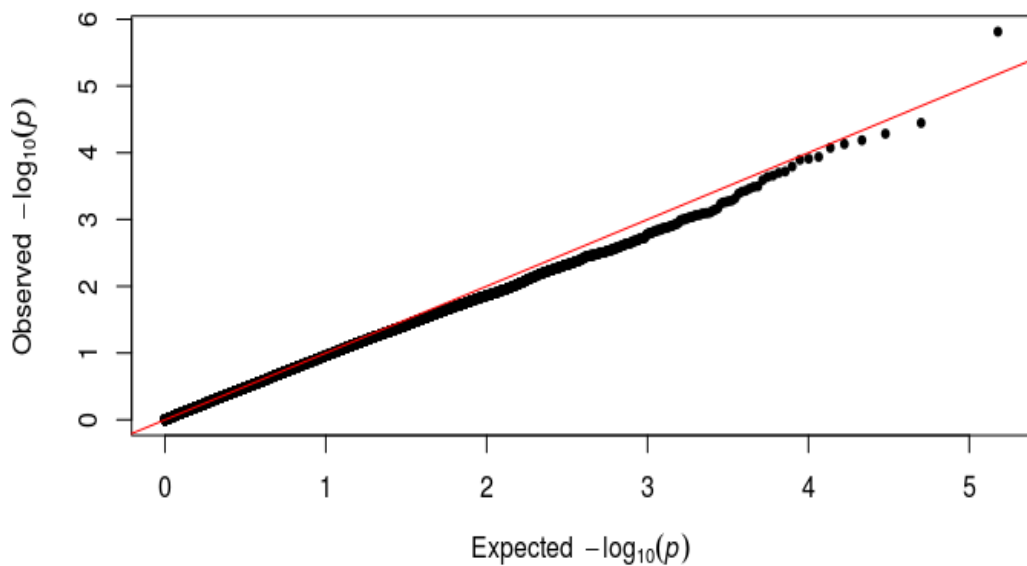


Figure S6. QQ plot. The QQ plot shows no evidence for inflation or bias in the association analysis of DNA methylation with “bad” LEQ scores ($\Lambda = 0.96$).

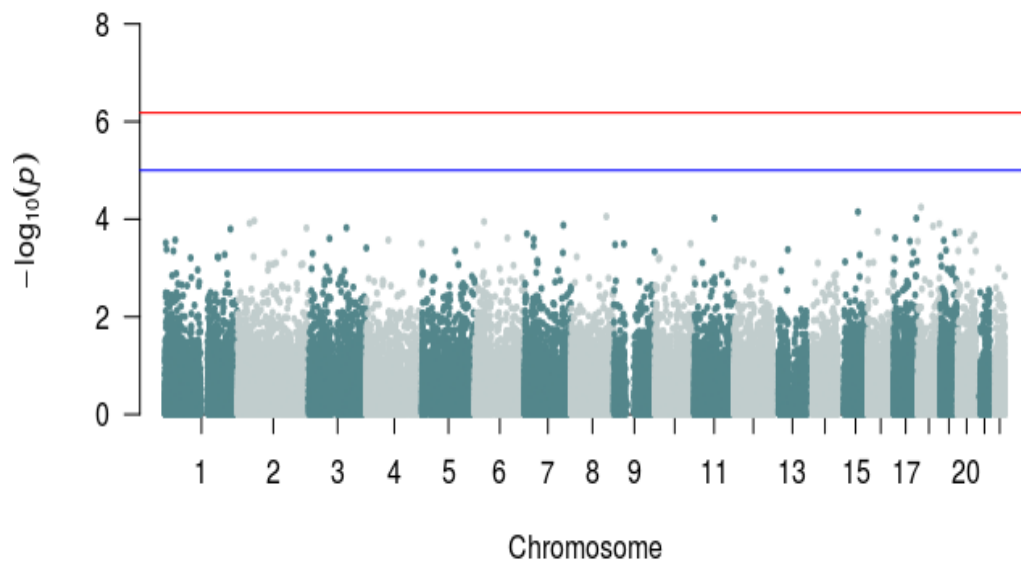


Figure S7. Manhattan plot. The Manhattan plot depicts the association between DNA methylation and the interaction between childhood trauma and total LEQ scores ($n = 191$). The horizontal red line represents the epigenome-wide significant threshold for this study ($p < 6.6 \times 10^{-7}$) and the blue line represents the suggestive significance threshold ($p < 1.0 \times 10^{-5}$).

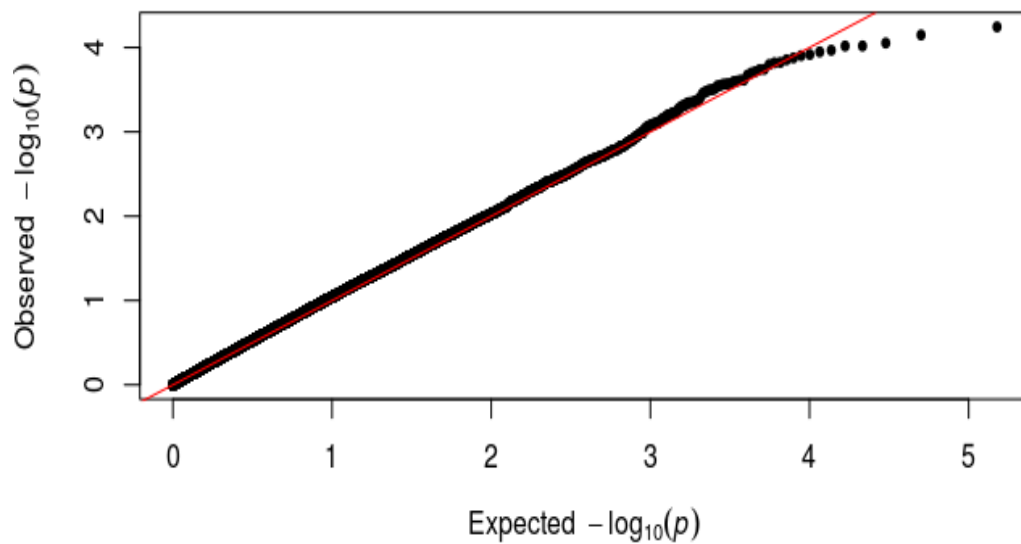


Figure S8. QQ plot. The QQ plot shows no evidence for inflation or bias in the association analysis of DNA methylation with the interaction between childhood trauma and total LEQ scores (Lambda = 1.10).

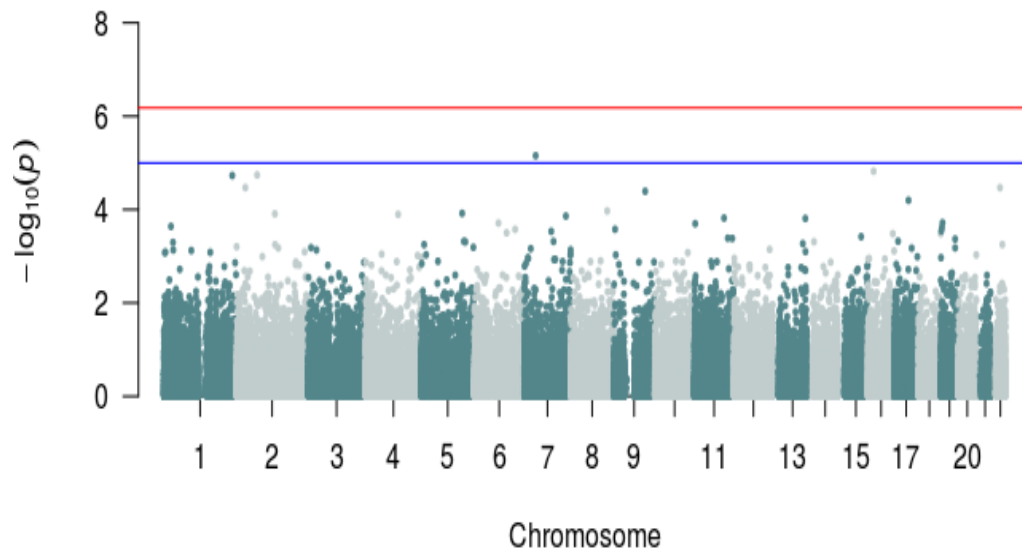


Figure S9. Manhattan plot. The Manhattan plot depicts a sensitivity analysis testing the association between DNA methylation and total LEQ scores after removing patients not taking psychotropic drugs at the time of testing ($n = 186$). The horizontal red line represents the epigenome-wide significant threshold for this study ($p < 6.6 \times 10^{-7}$) and the blue line represents the suggestive significance threshold ($p < 1.0 \times 10^{-5}$).

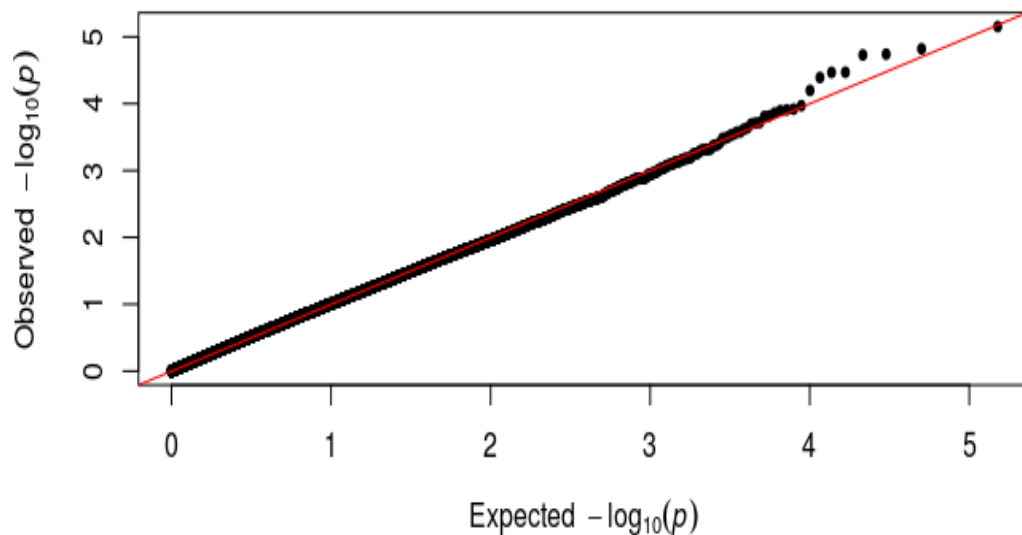


Figure S10. QQ plot. The QQ plot shows no evidence for inflation or bias in the association analysis of DNA methylation with total LEQ scores after removal of patients not on taking psychotropic drugs at the time of testing ($\Lambda = 1.06$).

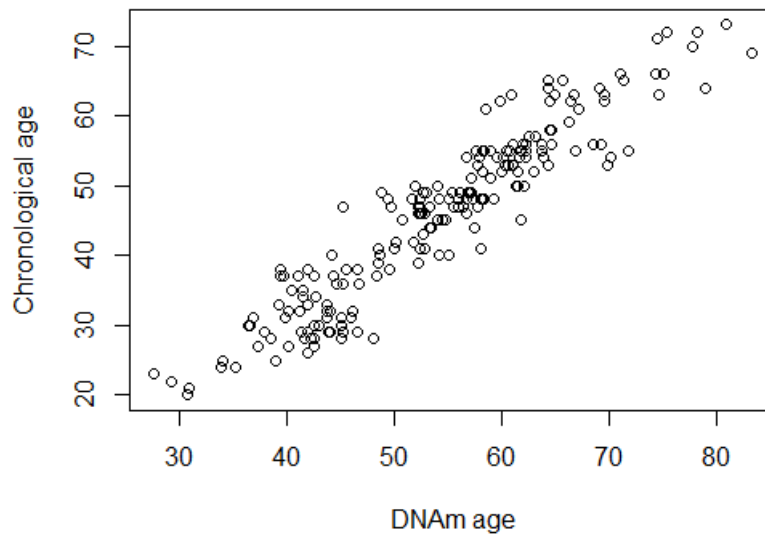


Figure S11. Scatterplot. The scatterplot illustrates the significant and positive correlation between DNA methylation age (DNAm age in years, calculated based on the Horvath algorithm) and chronological age (years) at baseline (Spearman's $\rho = 0.94$).

Table S3. GOMeth enrichment analysis

Biological process	N	DE	P.DE	FDR
homophilic cell adhesion via plasma membrane adhesion molecules	136	54	7.28E-05	1
spindle midzone assembly	8	6	0.001	1
cell-cell adhesion via plasma-membrane adhesion molecules	199	64	0.001	1
leukotriene biosynthetic process	10	6	0.002	1
exploration behavior	22	11	0.003	1
maintenance of unfolded protein	3	3	0.003	1
maintenance of unfolded protein involved in ERAD pathway	3	3	0.003	1
regulation of epithelial cell migration	154	41	0.004	1
myelination	87	26	0.004	1
vesicle targeting, trans-Golgi to endosome	2	2	0.004	1
regulation of endothelial cell migration	110	30	0.005	1
negative regulation of osteoclast development	5	4	0.006	1
ensheathment of neurons	90	26	0.006	1
axon ensheathment	90	26	0.006	1
positive regulation by host of viral transcription	9	5	0.007	1
cellular response to arsenic-containing substance	12	5	0.007	1
germinal center B cell differentiation	2	2	0.007	1
hormone-mediated apoptotic signaling pathway	3	3	0.007	1
positive regulation of axon extension	26	11	0.007	1
nervous system development	1747	369	0.008	1
Abbreviations: N – number of genes in the GO; DE – number of genes that are differentially methylated; P.DE – <i>p</i> -value for the over-representation of the GO term				

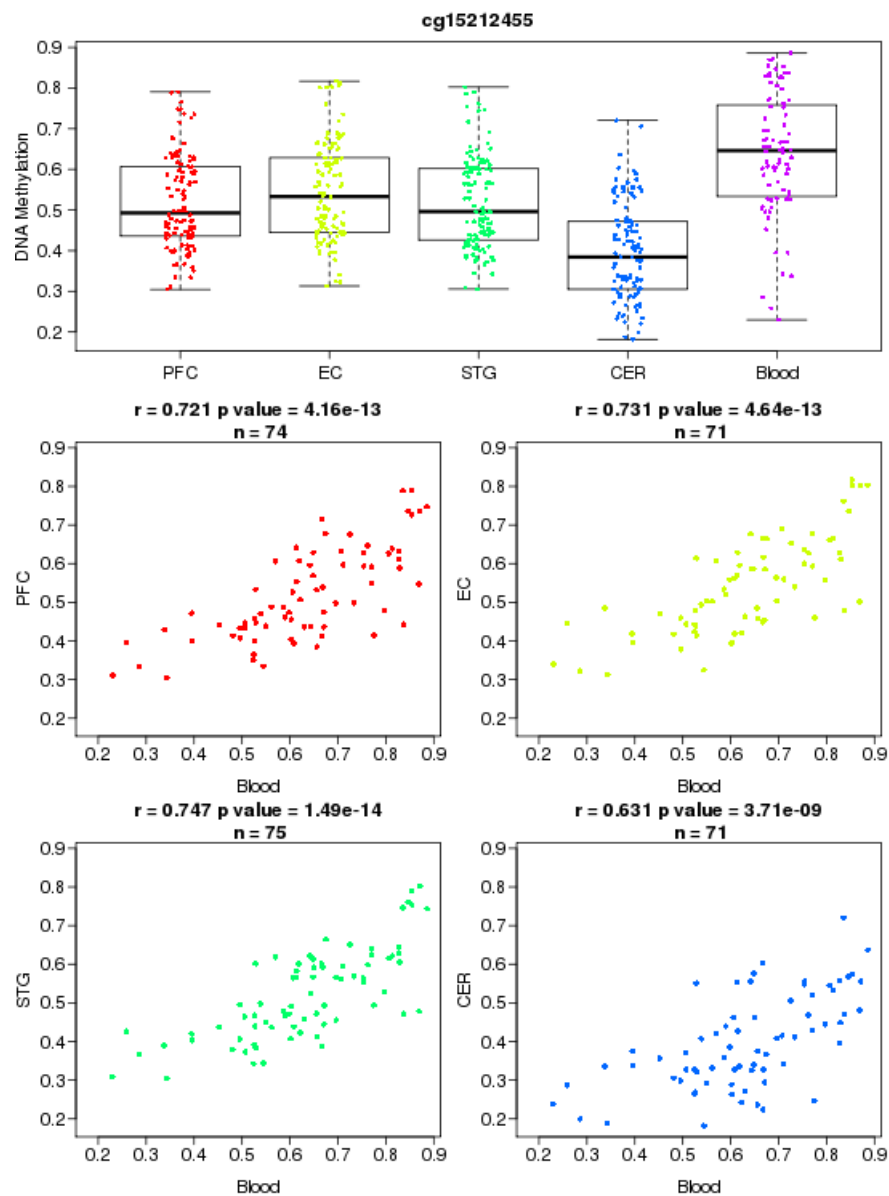


Figure S12. Blood-brain methylation correlation. Publicly available data from Hannon et al. was used to assess cg15212455 DNA methylation patterns across blood, the prefrontal cortex (PFC), entorhinal cortex (EC), superior temporal gyrus (STG), and cerebellum (CER).

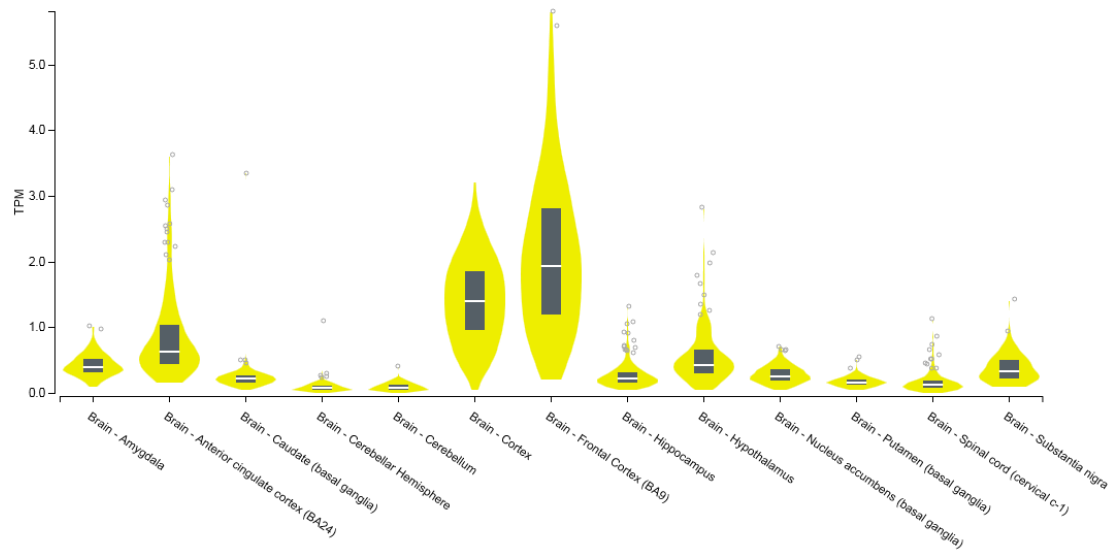


Figure S13. Gene expression patterns of *POU6F2* across multiple brain regions. The freely available Genotype-Tissue Expression (GTEx) Project Database portal (www.gtexportal.org) was used to determine the expression patterns of *POU6F2* across multiple tissues. The figure shows expression levels across multiple brain regions, showing the highest expression of *POU6F2* in the frontal cortex.

Original Article 2: Supplementary references

Hannon, E., Lunnon, K., Schalkwyk, L., & Mill, J. (2015). Interindividual methylomic variation across blood, cortex, and cerebellum: Implications for epigenetic studies of neurological and neuropsychiatric phenotypes. *Epigenetics*, 10(11), 1024-1032. doi:10.1080/15592294.2015.1100786

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- Dwyer, D.B., Kalman, J.K., Budde, M., [...], **Comes, A.L.**, [...], Falkai, P., Schulze, T.G., Koutsouleris, N. (2020). An investigation of psychosis subgroups with prognostic validation and exploration of genetic underpinnings: The PsyCourse Study. *JAMA Psychiatry*, doi:10.1001/jamapsychiatry.2019.4910. [Epub ahead of print]
- Comes, A.L.**, Senner, F., Budde, M., [...], Falkai, P., Schulze, T.G., Papiol, S. (2019). The genetic relationship between educational attainment and cognitive performance in major psychiatric disorders. *Transl Psychiatry*, 9(1), 210. doi:0.1038/s41398-019-0547-x.
- Anderson-Schmidt, H., Gade, K., Malzahn, D., [...], **Comes, A.L.**, [...], Wiltfang, J., Falkai, P., Schulze, T. G. (2019). The influence of religious activity and polygenic schizophrenia risk on religious delusions in schizophrenia. *Schizophr Res*, 210, 255-261. doi:10.1016/j.schres.2018.12.025
- Budde, M., Anderson-Schmidt, H., Gade, K., [...], **Comes, A.L.**, [...], Falkai, P., Schulze, T.G., Heilbronner, U. (2019). A longitudinal approach to biological psychiatric research: The PsyCourse study. *Am J Med Genet B Neuropsychiatr Genet*, 180(2), 89-102. doi:10.1002/ajmg.b.32639
- Budde, M., Friedrichs, S., Alliey-Rodriguez, N., [...], **Comes, A.L.**, [...], Rietschel, M., Schulze, T.G., Malzahn, D. (2019). Efficient region-based test strategy uncovers genetic risk factors for functional outcome in bipolar disorder. *Eur Neuropsychopharmacol*, 29(1), 156-170. doi:10.1016/j.euroneuro.2018.10.005
- Kalman, J.L., Papiol, S., Forstner, A.J., [...], **Comes, A.L.**, [...], Nöthen, M., Rietschel, M., Schulze, T.G. (2019). Investigating polygenic burden in age at disease onset in bipolar disorder: Findings from an international multicentric study. *Bipolar Disord*, 21(1), 68-75. doi:10.1111/bdi.12659
- Comes, A.L.**, Papiol, S., Mueller, T., Geyer, P.E., Mann, M., Schulze, T.G. (2018). Proteomics for blood biomarker exploration of severe mental illness: pitfalls of the past and potential for the future. *Transl Psychiatry*, 8(1), 160. doi:10.1038/s41398-018-0219-2